

Vertical stratification-driven nutrient ratios regulate phytoplankton community structure in the oligotrophic western Pacific Ocean

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Abstract: The stratification of the upper oligotrophic ocean have a direct impact on biogeochemistry by regulating the components of the upper-ocean environment that are critical to biological productivity, such as light availability for photosynthesis and nutrient supply from the deep ocean. We investigated the spatial distribution pattern and diversity of phytoplankton communities in the western Pacific Ocean (WPO) in the autumn of 2016, 2017, and 2018. Our results showed the phytoplankton community structure mainly consisted of cyanobacteria, diatoms, and dinoflagellates, while the abundance of Chrysophyceae was negligible. Phytoplankton abundance was high from the equatorial region to 10 °N, and decreased with increasing latitude in spatial distribution. Phytoplankton also showed a strong variation in the vertical distribution. The potential influences of physicochemical parameters on phytoplankton abundance were analyzed by Structural Equation Model (SEM) to determine nutrient ratios driven by vertical stratification to regulate phytoplankton community structure in the typical oligotrophic ocean. Regions with strong vertical stratification were more favorable for cyanobacteria, whereas weak vertical stratification was more conducive to diatoms and dinoflagellates. Our study shows that stratification is a major determinant of phytoplankton community structure; and highlights that physical process in the ocean control phytoplankton community structure by driving the balance of chemical elements, providing a data base to better predict models of changes in phytoplankton community structure under future ocean scenarios.

Keywords: Vertical stratification; phytoplankton community; western Pacific Ocean; N:P ratio

1. Introduction

Phytoplankton contribute nearly half of global primary production (Field et al., 1998) and represents an important part of biogeochemical cycling and transformation (Falkowski et al., 1998). Marine phytoplankton link the cycling of different elements through their demand for multiple nutrients such as nitrogen (N), phosphorus (P) or iron and their relative availability (Hillebrand et al., 2013). The Redfield ratio is probably the most powerful generalization and cornerstone of the marine biogeochemical cycle (Redfield et al., 1963; Schindler, 2003). The nutrient requirements of phytoplankton are limited by the environmental conditions in which they grow, and nutrient limitation increases the N: P ratio of primary production (Carlson, 2002; Fogg, 1983; Karl et al., 1998). Nitrogen fixation by phytoplankton may deplete phosphorus from the upper ocean, causing an increase to the N: P ratios (Karl et al., 2001). The photosynthesis does not cease, even when there are not enough nutrients to grow (Bertilsson et al., 2003; Geider et al., 1998; Goldman et al., 1979).

Upper-ocean stratification plays an important role in the climate system and in many marine

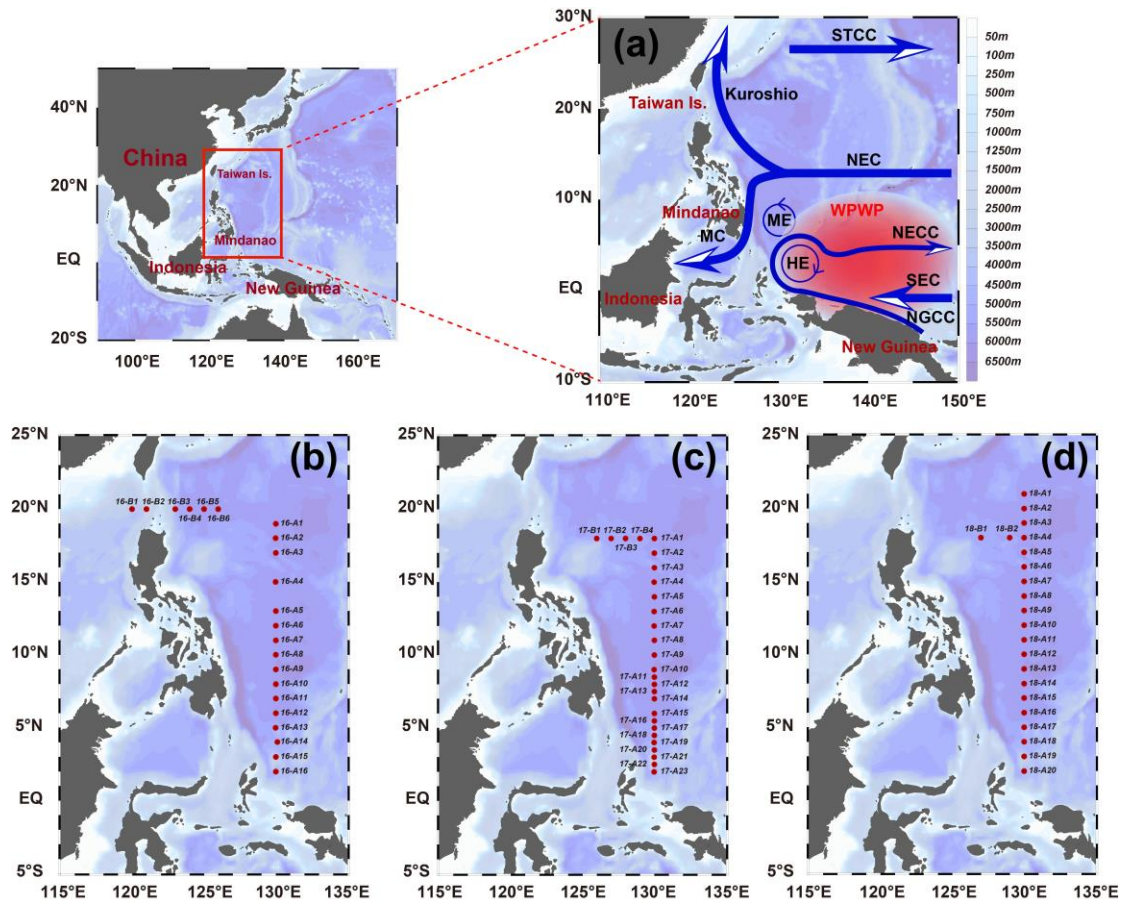
1 biogeochemical processes. The degree of vertical mixing is controlled by the strength of nearsurface
2 density stratification (Cronin et al., 2013; Qiu et al., 2004), which impacts the formation of the
3 surface mixed layer (ML) and the entrainment process at the ML's base. The ML depth directly
4 modulates the oceanic reaction to atmospheric forcing and the ocean ventilation process that
5 includes the sinking of water masses into the ocean interior, accompanied by heat, carbon, and
6 oxygen. Upper ocean stratification can directly affect important processes such as biogeochemistry
7 and primary production by regulating the light supply for photosynthesis and nutrient supply from
8 the subsurface ocean (Yamaguchi and Suga, 2019). While the strengthened stratification may
9 produce better light availability for the phytoplankton community, it will also prevent vertical
10 nutrient supply to the euphotic zone from the deep sea (Doney, 2006). Previous studies have shown
11 that net primary production (NPP) shows a stronger linear decrease with stronger vertical
12 stratification and a significant decrease in surface nitrate and phosphate concentrations. The
13 decrease in NPP can be partly explained by the increase in vertical stratification that leads to changes
14 in nutrient concentration (Yamaguchi and Suga, 2019).

15 In the present study, we focused on the vertical structure of the change in the ocean temperature
16 and salinity, that is, the change in the density stratification. The vertical stratification index (VSI)
17 used in this study is the potential density difference between the surface layer and the depth of 200
18 meters ($\Delta\rho_{200}$), which can quantify the strength of the upper ocean stratification well (Mena et al.,
19 2019). The purpose of this study is to determine the community composition mechanisms that drive
20 phytoplankton in oligotrophic region. These mechanisms are related to vertical stratification and
21 nutrient ratios. We explored how vertical stratification affects the composition of phytoplankton
22 communities. We hypothesized that vertical stratification might regulate phytoplankton abundance
23 and community composition by driving the ratio of nutrients.

24 25 2. Materials and methods

26 2.1. Study area and sampling

27 This study relied on the shared voyage of the WPO (0–20 °N, 120–130 °E), commissioned by
28 the National Natural Science Foundation of China. Physical, biological, chemical, and geological
29 surveys were carried out from September to November in 2016, 2017, and 2018 aboard the R/V
30 *Kexue*. The sampling stations used in this study are shown in Figure 1; the sampling layers were 5,
31 25, 50, 75, 100, 150, and 200 m. Phytoplankton samples from different water layers were placed in
32 1 L polyethylene bottles, fixed in formaldehyde solution (3%), and stored in dark. Nutrient samples
33 from different layers were placed in PE bottles, frozen, and stored at –20 °C for laboratory nutrient
34 analysis.



1
2 Figure 1. Stations in the western Pacific Ocean (WPO) of three cruises. (a): Current systems of the
3 WPO; (b), (c), and (d): sampling stations of 2016, 2017 and 2018 cruises, respectively. The station
4 at 130°E forms the section A, and the station at 20 °N forms the section B. Map of the WPO shows
5 the major geographic names and the surface currents, including the Subtropical Counter Current
6 (STCC), the North Equatorial Current (NEC), the Northern Equatorial Counter Current (NECC),
7 the South Equatorial Current (SEC), the New Guinea Coastal Current (NGCC), the Mindanao
8 Current (MC), the Mindanao Eddy (ME), the Halmahera Eddy (HE).

9
10 **2.2. Identification of Phytoplankton**

11 After returning to the laboratory, the Utermöhl method was applied for phytoplankton analysis.
12 A 1 L subsample was allowed to stand for 48 h; then 800 mL supernatant was removed carefully by
13 siphoning through a catheter, taking care to prevent the catheter from touching the bottom of the
14 bottle. Thereafter, the remaining 200 mL liquid was gently mixed and half of which was further
15 concentrated with a 100 mL sedimentation column (Utermöhl method) for 48 h sedimentation (Sun
16 et al., 2002a). The phytoplankton species were identified and enumerated under an inverted
17 microscope (AE2000, Motic, Xiamen, China) at 400× (or 200×) magnification. Phytoplankton
18 identification was conducted as described by Jin et al. (1965), Yamaji (1991), and Sun et al. (2002b),
19 and the World Register of Marine Species (<http://www.marinespecies.org>). Species identification
20 was as close as possible to the species level. The minimum size of the organisms identified and
21 counted is 20 μm.

22
23 **2.3. Laboratory Nutrient Analysis**

1 The Technicon AA3 Auto-Analyzer (Bran + Luebbe, Norderstedt, Germany) based on classical
 2 colorimetric methods was used for the analysis and determination nutrient (Grasshoff et al., 2009).
 3 Soluble inorganic phosphorus (PO₄-P) was determined by the phosphomolybdenum blue method
 4 with the limit of detection of 0.02 μmol L⁻¹; dissolved silicate (SiO₃-Si) was determined by the
 5 silicon molybdenum blue method with the limit of detection of 0.02 μmol L⁻¹; nitrate (NO₃-N) was
 6 determined by the cadmium column method with the limit of detection of 0.01 μmol L⁻¹; nitrite
 7 (NO₂-N) was determined by the naphthalene ethylenediamine method with the limit of detection of
 8 0.01 μmol L⁻¹ (Dai et al., 2008). Ammonia (NH₄-N) was determined by the sodium salicylate
 9 method with the limit of detection of 0.03 μmol L⁻¹ (Guo et al., 2014; Pai et al., 2001). Nitrogen-to-
 10 phosphorous (N: P) ratio was calculated by dividing nitrogen concentration (NO₃⁻+NO₂⁻) by
 11 phosphate concentration.

12 2.4. Analysis and methods

13 A SBE911 CTD sensor and standard Sea-Bird Electronics methods were used to process
 14 recorded hydrological parameters. The depth of the mixed layer (ML) is calculated as

$$15 (S, T) = (S_{ref}, T_{ref} - \Delta T)$$

16 S and T are the salinity and temperature, respectively, and S_{ref} and T_{ref} are the temperature and
 17 salinity at 5 m, ΔT is equal to 0.5 °C.

18 We calculated the vertical stratification index (VSI) to indicate the degree of vertical
 19 stratification of the water column:

$$20 VSI = \sum [\delta_{\theta}(m+1) - \delta_{\theta}(m)]$$

21 where δ_θ is the potential density anomaly, and m is the depth from 5 to 200 m.

22 The abundance of phytoplankton cells in water column was calculated through the trapezoidal
 23 integral method (Zhu et al., 2019):

$$24 P = \left\{ \sum_{i=1}^{n-1} \frac{P_{i+1} + P_i}{2} (D_{i+1} - D_i) \right\} / (D_n - D_1)$$

25 where P is the average value of phytoplankton abundance in water column, P_i is the abundance
 26 value of phytoplankton in layer *i*, *i* + 1 is the layer *i* + 1, D_n is the maximum sampling depth, D_i
 27 is the depth of layer *i*, and n is the sampling level.

28 We clustered all species based on Bray-Curtis similarity distance the three years, and the results
 29 showed four distinct regions using the Primer (version 6). Distance-based Redundancy analysis (db-
 30 RDA) and Principal Co-ordinates Analysis (PCoA) were performed using the R package vegan
 31 (version 2.5-7) (Oksanen et al., 2020) to explain the relationship between the environmental
 32 parameters (temperature, salinity, depth, VSI, Dissolved inorganic nitrogen (DIN) and Dissolved
 33 inorganic phosphorus (DIP) and Dissolved silicate (DSi)) and phytoplankton community structure.
 34 The results were visualized using the R package ggplot2 (version 3.3.2). SEM was used to assess
 35 the relative direct and indirect impact of physical and chemical parameters on phytoplankton
 36 abundance. The chi-square test (χ²), comparative fit index (CFI), and goodness fit index (GFI) were
 37 used to assess the model fit.

38 3. Results

39 3.1 Hydrographic features of the study area during the sampling years

1 The surface temperature and salinity of the surveyed sea area in 2016, 2017, and 2018 are
2 shown in Figure 2. In general, the temperature increased with decreasing latitude, and the stations
3 near the equator exhibited the highest temperature; in contrast, the salinity showed an opposite
4 trend as that of temperature, with a high value from 15 °N to 20 °N. The surface temperature (Fig.
5 2) of the surveyed area in 2016 ranged from 28.58 °C (station 16-B1) to 30.14 °C (station 16-A16),
6 with an average of 29.43 °C. The surface salinity (Fig. 2) of the surveyed area in 2016 ranged from
7 33.80 (station 16-B2) to 34.65 (station 16-A2), with an average of 34.32. The surface temperature
8 (Fig. 2) of the surveyed area in 2017 ranged from 27.91 °C (station 17-A4) to 30.19 °C (station 17-
9 A20), with an average of 29.26 °C. The surface salinity (Fig. 2) of the surveyed area in 2017 ranged
10 from 33.38 (station 17-A16) to 34.64 (station 17-B4), with an average of 33.94. The surface
11 temperature (Fig. 2) of the surveyed sea area in 2018 ranged from 26.33 °C (station 18-B1) to
12 29.79 °C (station 18-A17), with an average of 28.83 °C. The surface salinity (Fig. 2) of the surveyed
13 sea area in 2018 ranged from 33.77 (station 18-A14) to 34.64 (station 18-B1), with an average of
14 34.21.

15 The profile distribution of temperature and salinity based on the cross-sectional data of
16 different water layers at each station obtained from the survey is shown in Figure 2. The temperature
17 of the shallow water column (0–100 m) is higher than that of the deep-water column (100–200 m).
18 The salinity values of the deep-water bodies (100–200 m) were higher than those of the shallow
19 water bodies (0–100 m). The values of temperature and salinity in 2016, 2017, and 2018 did not
20 change significantly. The temperature of the section in 2016 ranged from 12.16 °C (200 m at station
21 16-A11) to 30.14 °C (5 m at station 16-A16), with an average of 25.74 °C. The salinity of the section
22 in 2016 ranged from 33.80 (5 m at station 16-B2) to 35.39 (150 m at station 16-A16), with an
23 average of 34.61 °C. The temperature of the section in 2017 ranged from 11.16 °C (200 m at station
24 17-A13) to 30.19 °C (5 m at station 17-A20), with an average of 25.18 °C. The salinity of the section
25 in 2017 ranged from 33.38 (5 m at station 17-A16) to 35.24 (150 m at station 17-A23), with an
26 average of 34.46. The temperature of the section in 2018 ranged from 9.65 °C (200 m at station 18-
27 A14) to 29.79 °C (5 m at station 18-A17), with an average of 24.22 °C. The salinity of the section
28 in 2018 ranged from 33.77 °C (5 m at station 18-A14) to 35.39 °C (150 m at station 18-A17), with
29 an average of 34.57.

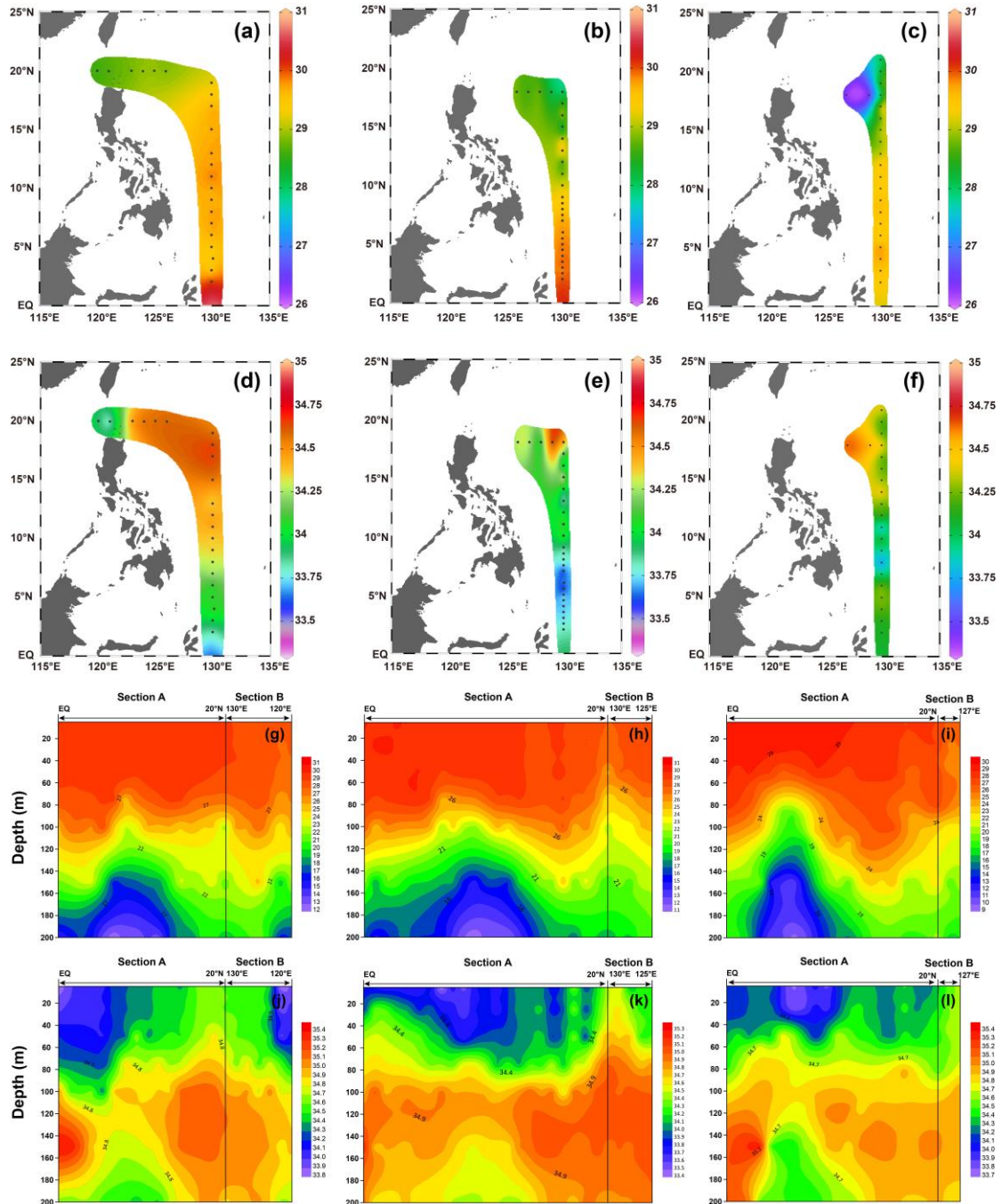
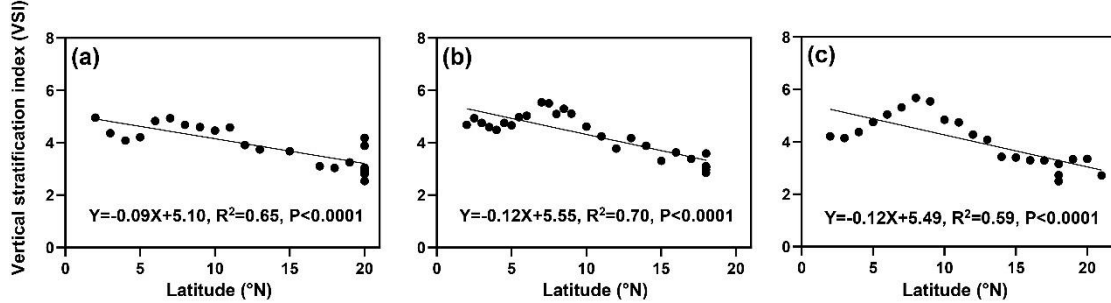


Figure 2. The temperature and salinity distribution in the WPO from three cruises. (a–c) surface temperature in 2016, 2017, 2018 respectively, (d–f) surface salinity in 2016, 2017, 2018 respectively, (g–i) vertical distribution of temperature in 2016, 2017, 2018 respectively, (j–l) vertical distribution of salinity in 2016, 2017, 2018 respectively.

The distribution of the VSI in latitude for the three cruises is shown in Figure 3. Overall, the VSI showed a similar distribution pattern in the three cruises, with the highest value occurring at 7–8 °N and a decreasing trend with increasing latitude. In the 2016 cruise (Figure 3-a), the minimum value of VSI (2.54) appeared in the station at 20 °N (station 16-B4), and the maximum value (4.94) appeared in the station at 7 °N (station 16-A11), with an average of 3.90 ± 0.76 . In the 2017 cruise (Figure 3-b), a minimum value of VSI (2.85) appeared in the station at 18 °N (station 17-B4), and

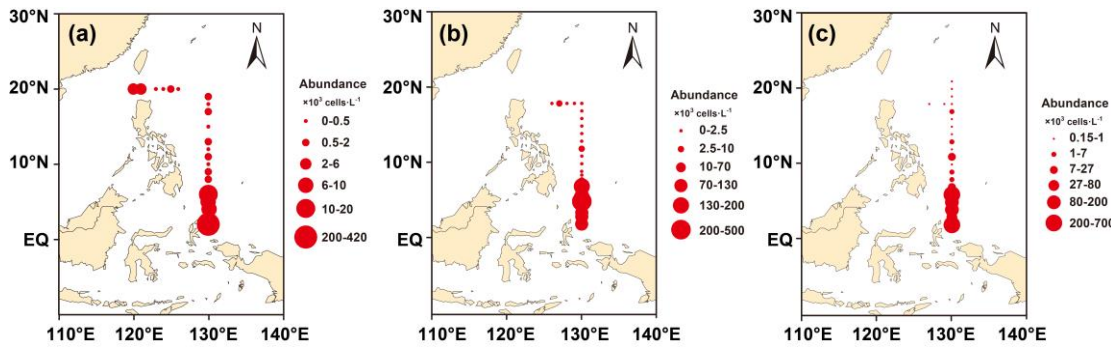
1 the maximum value (5.54) appeared in the station at 7 °N (station 17-A14) with an average of 4.30
 2 ± 0.82. In the 2018 cruise (Figure 3-c), the minimum value of VSI (2.50) occurred in the station at
 3 18 °N (station 18-B1), and the maximum value (5.48) occurred in the station at 8 °N (station 18-
 4 A14) with an average of 4.01 ± 0.95. Interestingly, the VSI varied significantly across latitudinal
 5 regions; the VSI was high from the equator to 10 °N, while it was low at 10–20 °N.



6
 7 Figure 3. Linear fits of the vertical stratification index with latitude (a) in 2016, (b) in 2017, (c) in
 8 2018. The black dots are the VSI of each station.

9
 10 3.2 Interannual variation of phytoplankton communities

11 Figures 4a, b, and c show the horizontal distribution of surface phytoplankton abundance from
 12 2016 to 2018. The interannual variation in phytoplankton was relatively stable, and the sampling
 13 area and sampling time from 2016 to 2018 were generally consistent. Most phytoplankton species
 14 varied little from year to year in their distribution. Phytoplankton distribution showed a trend of
 15 decreasing abundance from the equator to the north with a minor abundance peak at about 10°N.
 16 This abundance peak was associated with the predominance of *Trichodesmium*. However, affected
 17 by coastal currents, high abundance patches dominated by diatoms were observed also in the Luzon
 18 Strait area south of Taiwan, which were carried to the surface by upwelling currents and accounted
 19 for more than 67.76% of the abundance at this station. Relatively high abundances were observed
 20 at stations in the Kuroshio extension region, consisting mainly of cosmopolitan and warm water
 21 species. Phytoplankton abundance was the lowest in the high latitude region.

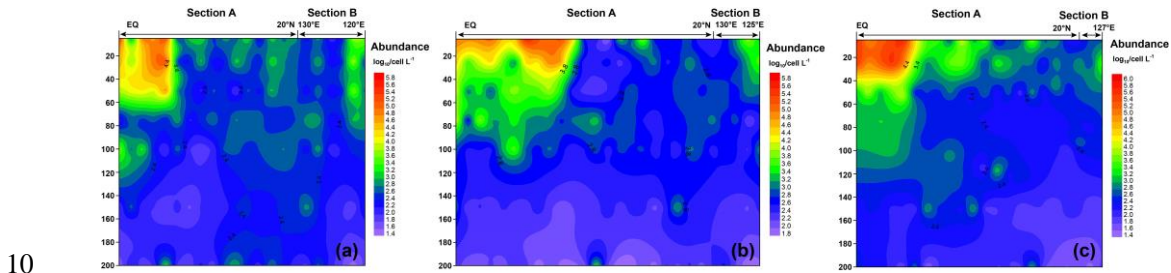


22
 23 Figure 4. Horizontal distribution of phytoplankton abundance in the WPO. a. 2016, surface layer; b.
 24 2017, surface layer; and c. 2018, surface layer.

25
 26 3.3 Vertical distribution of phytoplankton abundance

27 Figure 5 shows the vertical distribution of the phytoplankton. The overall trend in the WPO
 28 was consistent across the three cruises in 2016 (a), 2017 (b), and 2018 (c), with the phytoplankton
 29 distribution showing variations with latitude and differences in vertical distribution at depth. In

1 terms of latitude, high phytoplankton value areas were concentrated near the equator (0 °E–8 °E).
 2 Vertical distribution of phytoplankton indicated that the plankton-abundant areas occurred from 0–
 3 50 m, and the phytoplankton abundance gradually decreased with the increase in depth. Vertical
 4 distribution of phytoplankton abundance differed significantly across different areas. In the areas
 5 near the equator affected by Halmahera Eddy (HE) and Mindanao Eddy (ME), phytoplankton
 6 abundance was mainly concentrated in the upper water column (0–50 m) and consisted mainly of
 7 cyanobacteria. In the northern area affected by Kuroshio, the lower phytoplankton abundance was
 8 mostly dominated by the equatorial stations, while the phytoplankton species composition was
 9 mostly dominated by diatoms and dinoflagellates.

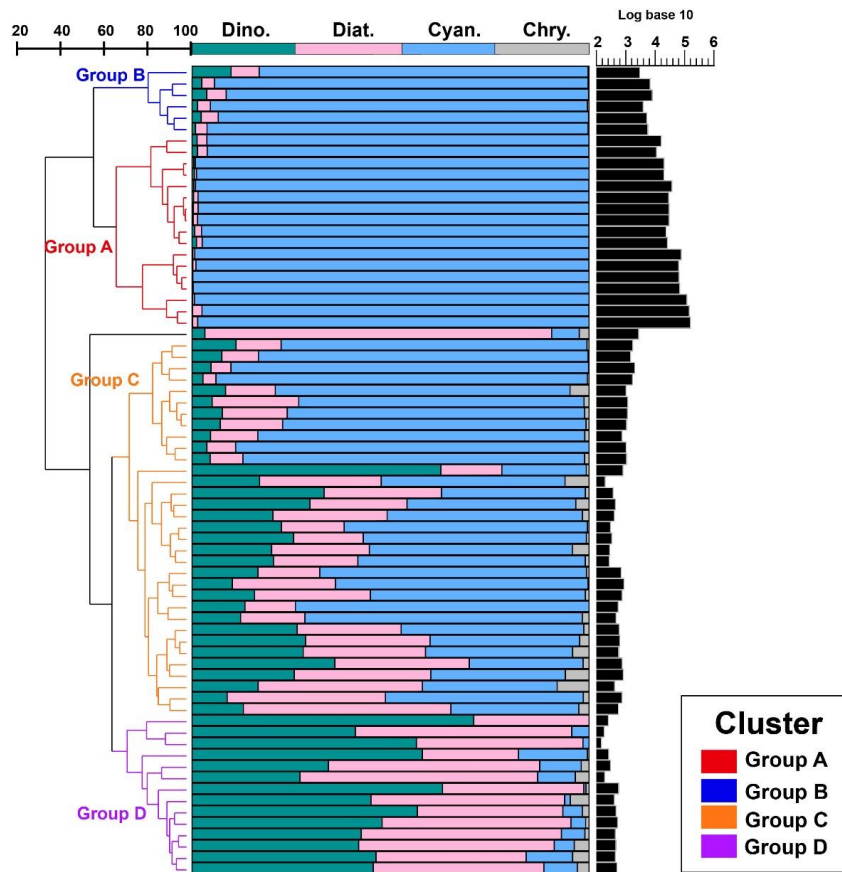


10
 11 Figure 5. Vertical distribution of phytoplankton abundance ($\text{Log}_{10} \text{ cells L}^{-1}$) in the WPO in 2016
 12 (a); 2017 (b) and 2018 (c).

13 3.4 Phytoplankton community structure

14 Since there was little interannual difference between species, we clustered all species based on
 15 Bray-Curtis similarity distance for stations, and the results showed four distinct regions (Figure 6).
 16 Cluster analysis divided the phytoplankton communities at the sampling sites for three years into
 17 four groups. Cyanobacteria (>90%) were the dominant species in Groups A and B. The species ratio
 18 of diatoms to dinoflagellates in Group A (dias: dinos = 4.8) was higher than that in Group B (dias:
 19 dinos = 1.4). Cyanobacteria were the dominant (66%) phytoplankton at the stations of Group C,
 20 while diatoms (18%) and dinoflagellates (14%) constituted 32% of the population in this group.
 21 Diatoms (43%) and dinoflagellates (49%) dominated the stations in Group D, accounting for
 22 approximately 92% of the total phytoplankton. The proportion of Chrysophyceae was low in all four
 23 groups (Table 1). The dendrogram showed that these populations were grouped into four groups,
 24 which were essentially identical to those determined by PCoA analysis (Figure 7).
 25

26



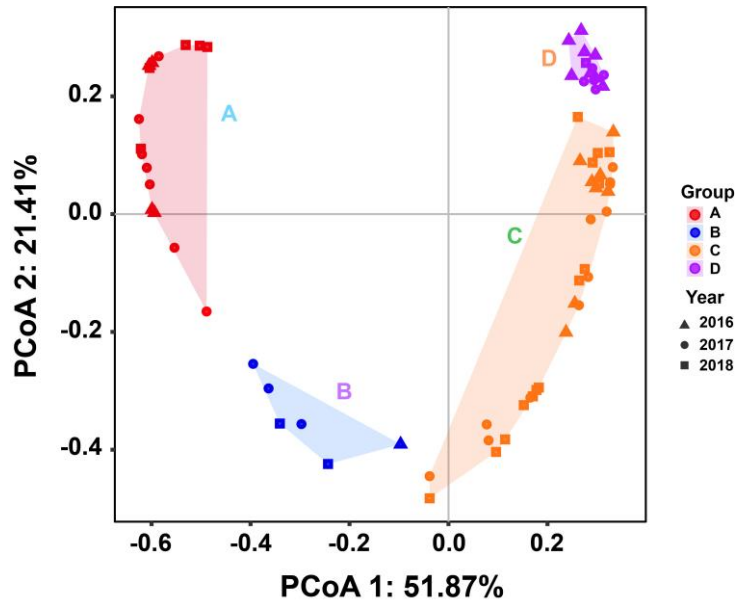
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2 Figure 6. Bray-Curtis similarity-based dendrogram showing averaged phytoplankton community
 3 composition and abundance for each station across the 3 cruises. For each station, community
 4 composition is indicated with bar plots, and phytoplankton abundance is represented with black bars.

5

6 Table 1. The percentages (%) (average \pm standard deviations) of diatoms, dinoflagellates,
 7 cyanobacteria and chrysophyceae in the four groups.

Species	Group A	Group B	Group C	Group D
Diatoms	1.09 \pm 0.79	4.25 \pm 1.57	21.83 \pm 11.45	43.71 \pm 10.12
Dinoflagellates	0.44 \pm 0.42	3.41 \pm 3.30	17.26 \pm 12.45	48.38 \pm 11.61
Cyanobacteria	98.45 \pm 1.10	92.08 \pm 4.79	59.05 \pm 20.38	6.06 \pm 4.93
Chrysophyceae	0.02 \pm 0.01	0.26 \pm 0.10	1.86 \pm 1.99	1.85 \pm 1.66



1

2 Figure 7. Principal Coordinates Analysis for groups. Triangles, circles, and squares represent 2016,
 3 2017, and 2018 stations, respectively. $P < 0.05$. Different colors represent different groups.
 4 Percentages of total variance are explained by coordinates 1 and 2, accounting for 51.87% and
 5 21.41%, respectively.

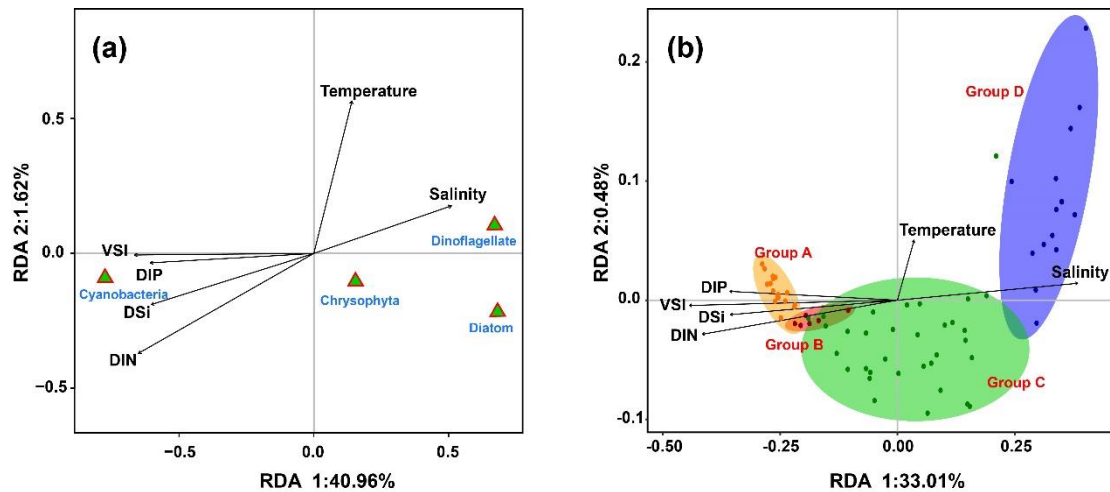
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7 3.5 Relationships between phytoplankton and environmental factors

8 The relationship between phytoplankton and environmental factors was analyzed using RDA.
 9 We obtained a two-dimensional distribution map of the species, sample distribution, and
 10 environmental factors (Figure 8). The results showed that different phytoplankton classes were
 11 correlated differently with environmental factors. Cyanobacteria showed negative correlations with
 12 temperature and salinity and positive correlations with VSI and nutrient concentration, indicating
 13 that waters with high VSI are suitable for the growth of cyanobacteria (mostly *Trichodesmium*).
 14 Diatoms and dinoflagellates exhibiting positive correlations with temperature and salinity and
 15 negative correlations with VSI and nutrient concentration, indicating that diatoms and
 16 dinoflagellates prefer waters with low VSI.

17 There were four distinct phytoplankton communities in the WPO: Group A was distributed in
 18 the equatorial region with clear vertical stratification. This community is characterized by high
 19 abundance and is dominated by *Trichodesmium* species such as *T. thiebautii*, *T. hildebrandtii*, and
 20 *T. erythraeum*, which are positively correlated with high concentrations of DIN, phosphate, and
 21 silicate. Group B was located near 8°N and is mainly influenced by the NECC and mesoscale eddy
 22 influence; the phytoplankton community was represented by warm water species, similar to that of
 23 Group A. Group C was mainly distributed in the 15 °N region and was strongly influenced by the
 24 NEC. Group D was mainly distributed in the 20 °N region, where it was directly influenced by the
 25 Kuroshio Current; here, the phytoplankton community was positively correlated with temperature
 26 and salinity.

27

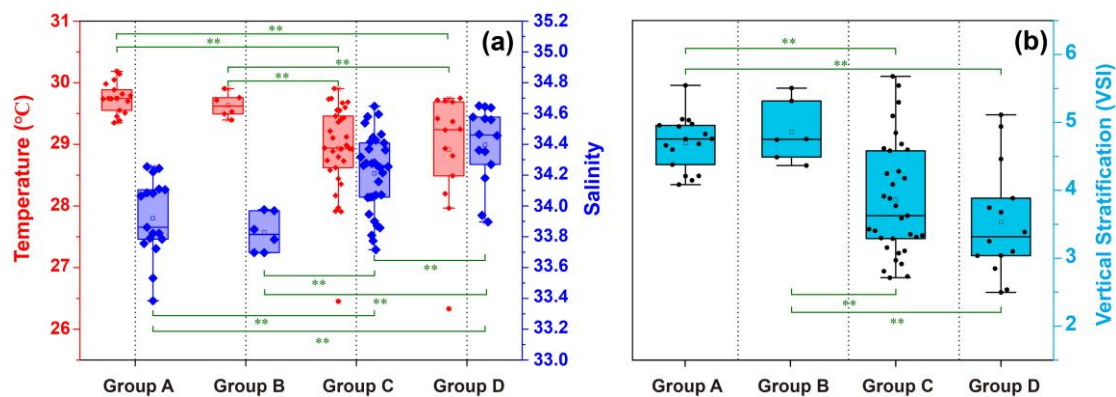


1
2 Figure 8. Redundancy analysis of the (a) phytoplankton and environmental parameters, (b) groups
3 and environmental parameters in the WPO. Colored dots represent sampling sites, triangles
4 represent phytoplankton species, and arrows represent environmental factors.

6 3.6 Temperature, salinity, and vertical stratification index

7 The temperature, salinity, and VSI of the four groups are shown in Figure 9. The temperature
8 and salinity (T-S) box diagram depicts the four main water masses in the WPO. Groups A (average
9 29.8 °C) and B (average 29.6 °C) had high temperatures, but the salinities of Groups A (average
10 33.9 °C) and B (average 33.8 °C) was low. The temperature of Groups C (average 28.9 °C) and D
11 (average 28.9 °C) was low, but the salinity of Groups C (average 34.2) and D (average 34.4) was
12 high (Fig. 9-a). Figure 9 shows clear variation in T-S, we also calculated the vertical stratification
13 index of the four groups (Figure 9-b). Compared with Groups C (average 3.86) and D (average 3.54),
14 the values of VSI in Groups A (average 4.69) and B (average 4.86) were markedly higher, and Group
15 A had the highest VSI. The stratification of the first two groups was more pronounced (Table 2).

16 The vertical stratification index was related to temperature (Figure 9-a) and salinity (Figure 9-
17 b). Temperature is positively correlated with the vertical stratification index. The VSI of all groups
18 was negatively correlated with salinity. The changes in temperature and salinity were most
19 pronounced in the vertical direction. In Groups A and B with a high stratification index, the changes
20 in temperature and salinity within the group were small. However, the temperature and salinity
21 changed significantly within Groups C and D, with a small stratification index.



23
24 Figure 9 Surface temperature and salinity (a), and vertical stratification index (b) of the four groups.

1

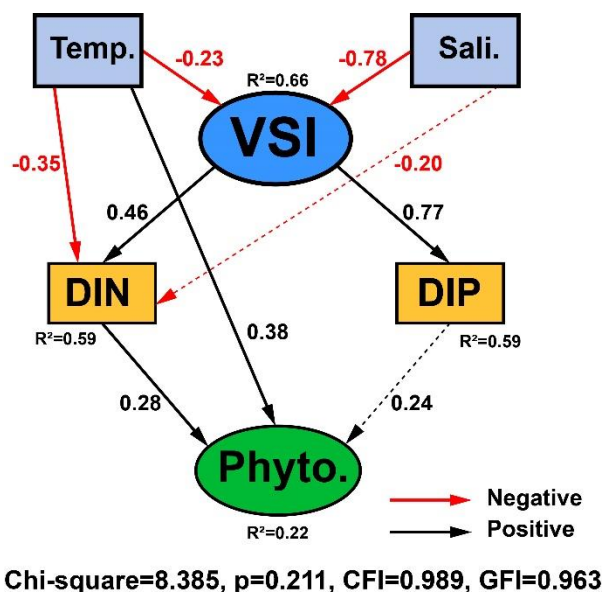
2 Table 2. Average (\pm standard deviations) values for nutrients ($\mu\text{mol L}^{-1}$), temperature ($^{\circ}\text{C}$), salinity
 3 for each phytoplankton community group were identified by the cluster analysis in the WPO.

	Group A	Group B	Group C	Group D
Temperature	25.30 \pm 1.06	24.45 \pm 1.85	24.92 \pm 1.32	25.41 \pm 1.23
Salinity	34.45 \pm 0.14	34.40 \pm 0.07	34.56 \pm 0.16	34.68 \pm 0.20
DIP	0.28 \pm 0.07	0.18 \pm 0.13	0.16 \pm 0.13	0.13 \pm 0.10
DIN	4.49 \pm 1.76	5.43 \pm 2.71	2.62 \pm 1.89	1.80 \pm 1.08
DSi	2.93 \pm 1.05	4.13 \pm 2.15	1.90 \pm 1.47	1.44 \pm 0.95
VSI	4.69 \pm 0.39	4.86 \pm 0.45	3.86 \pm 0.84	3.54 \pm 0.82

4

5 3.7 Direct vs. indirect effects of environmental parameters on phytoplankton abundance

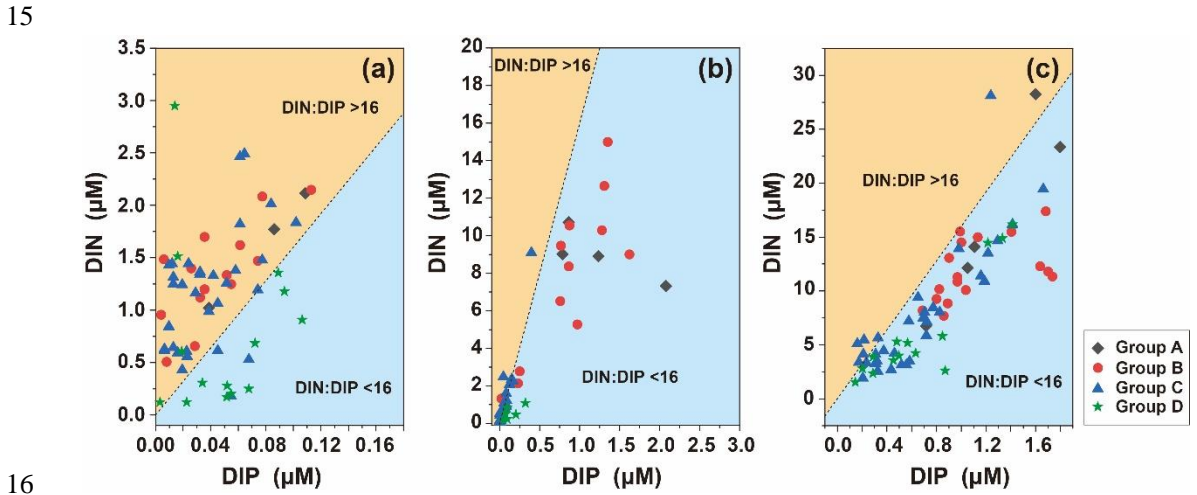
6 The causal relationships between measured phytoplankton abundance and relevant physical
 7 and chemical parameters were examined using SEM, using interactions between temperature,
 8 salinity, VSI, DIN, and DIP (Figure 10), as theoretical and experimental data indicated the
 9 importance of these variables. The model results showed that temperature, DIP, and DIN had a direct
 10 effect on phytoplankton abundance, with temperature having the largest direct effect on
 11 phytoplankton abundance (0.38), followed by DIN (0.28) and DIP (0.24). Temperature, salinity, and
 12 VSI had indirect effects on phytoplankton abundance, with temperature and salinity having negative
 13 indirect effects on phytoplankton abundance (-0.17 and -0.30) and VSI having positive indirect
 14 effects (0.31) (Figure 10). From the results of the total effect, only salinity had a negative effect on
 15 phytoplankton abundance (-0.30), while both temperature and VSI had positive effects on
 16 phytoplankton abundance (0.20 and 0.312), with VSI having the largest total effect. Although the
 17 direct effect of temperature on phytoplankton abundance was significant, it was partially offset by
 18 the indirect negative effect, while VSI had no direct effect on phytoplankton abundance, but its
 19 larger indirect effect resulted in the largest total effect. Both DIN and DIP had positive effects on
 20 phytoplankton abundance, but the effect of DIN was greater. Since the vertical distribution of DIN
 21 and DIP exhibited stronger variability, more specific analyses of DIN and DIP will be conducted
 22 later.



23

1 Figure 10. Structural Equation Model (SEM) analysis examining the effects of temperature, salinity,
 2 VSI, DIN and DIP on phytoplankton abundance. Solid black and red lines indicate significant
 3 positive and negative effects at $p < 0.05$, black and red dashed lines indicated insignificant effects.
 4 R^2 values associated with response variables indicate the proportion of variation explained by
 5 relationships with other variables. Values associated with arrows represent standardized path
 6 coefficients.

8 We analyzed the N:P ratio of the surface layer, SCM, and 200 m. The N:P ratio in the surface
 9 layer (N:P>16:1) indicates phosphorus limitation, which is consistent with the SEM analysis
 10 (Fig.11). The trophic structure of the SCM layer changed, N:P <16:1 indicated nitrogen limitation,
 11 and the depth continued to increase to the bottom of the euphotic layer and stabilized around N:P
 12 =16:1, indicating that at the bottom of the euphotic layer, as phytoplankton abundance decreased
 13 and interspecific competition decreased, the trophic ratio approached the Redfield ratio and growth
 14 may have become increasingly limited by light.



16 Figure 11. Distribution of phytoplankton community in DIN and DIP. (a): 5 m, (b): SCM, (b): 200
 17 m. The dashed line indicates the Redfield ratio N:P = 16:1.

19 4. Discussion

20 4.1 Comparison with historical data

21 The Kuroshio and WPWP are key areas of the WPO sea-air interaction and climate modulation
 22 (Zhang, 1999). Previous surveys have provided less knowledge of the phytoplankton community
 23 structure in this study area (Table 3). Previously, samples were collected by net, and net-collected
 24 samples reduced phytoplankton abundance in small volumes, thereby underestimating the
 25 phytoplankton abundance in the ocean under investigation. In the present study, phytoplankton
 26 samples were collected from water samples, which better reflected the phytoplankton community
 27 structure and abundance. Sun et al. (2000) and Liu et al. (2000) further investigated the species
 28 composition and abundance distribution of phytoplankton diatoms and dinoflagellates in the
 29 Ryukyu Islands and nearby waters. Li et al. (2015) conducted a study on phytoplankton in the
 30 tropical and subtropical Pacific oceanic zones with response mechanisms to the limitation of
 31 nitrogen and iron. Chen et al. (2018a) investigated the phytoplankton community structure and
 32 mesoscale eddies in the western boundary current. A total of 199 species in 61 genera belonging to
 33 four phytoplankton families were identified, among which the abundance of *Trichodesmium* species
 34

1 was high. Previous studies have mostly focused on vertical hauls and the horizontal distribution of
 2 phytoplankton throughout the water column while ignoring the effect of vertical stratification on
 3 phytoplankton.

4

5 Table 3. Historical data of the phytoplankton community in the WPO.

Month	Sampling areas	Layer /m	Number of species	Sampling types	References
2018.10	2°–20°N, 120°–130°E	0–200	305	Water samples	This study
2017.10	2°–18°N, 126°–130°E	0–200	339	Water samples	This study
2017.08	10.3°–10.9°N, 139.8°–140.4°E	0–200	147	Water samples	Dai et al., 2020
2017.08	21°–42°N, 118°–156°E	0–200	235	Water and net samples	Lin et al., 2020
2017.05	21°–42°N, 118°–156°E	0–200	248	Water and net samples	Lin et al., 2020
2016.09	2°–21°N, 127°–130°E	0–200	269	Water samples	This study
2016.09	0°–20°N, 120°–130°E	0–200	243	Net samples	Chen et al., 2018b
2014.08	0°–21.5°N, 121°–135.5°E	0–300	199	Net samples	Chen et al., 2018a
1997.07	23°30′–29°30′N, 122°30′–130°30′E	0–200	227	Net samples	Sun et al., 2000
1997.07	23°30′–29°30′N, 122°30′–130°30′E	0–200	251	Net samples	Liu et al., 2000

6

7 4.2 Relationship between N:P ratio and vertical distribution of phytoplankton

8 Research on the factors that control the structure of the phytoplankton community has been
 9 carried out for decades, but the hypothesis of nutrient concentration limits and ratios has not been
 10 fully explained in terms of affecting the structure of the phytoplankton community (Gao et al., 2019).
 11 As diatoms and dinoflagellates show great differences in cell morphology, structure, and nutrition
 12 mode, they differ greatly in their nutrient acquisition strategies. Several studies have revealed that
 13 dinoflagellates use mixotrophy, engulfing prey as well as feeding using peduncles and palia, while
 14 phosphorus limitation is a common factor stimulating dinoflagellates to ingest particulate nutrients
 15 (Huang et al., 2005; Smayda, 1997; Stoecker, 1999). The variation in phytoplankton community
 16 structure is always correlated with fluctuations in physicochemical environmental parameters.

17 In the four groups we studied, surface seawater N:P>16:1 indicated that phosphorus in surface
 18 seawater was limited, but *Trichodesmium* relied on its own nitrogen fixation function and was highly
 19 abundant in oligotrophic waters (Figure 6). The relationship between *Trichodesmium* and nitrogen
 20 fixation has already been demonstrated (Grosskopf et al., 2012; Luo et al., 2012; Zehr, 2011). The
 21 virtual absence nitrogen limitation in surface seawater in Group D was consistent with the low
 22 abundance of *Trichodesmium*, which was consistent with studies on the abundance of
 23 *Trichodesmium* in the region (Chen et al., 2019; Sohm et al., 2011). In the WPO, the most
 24 oligotrophic ocean around the world (Hansell et al., 2000), nutrients have become an important
 25 factor that determines the distribution of phytoplankton. Under nutrition-limited conditions, diatoms
 26 and dinoflagellates are more affected, especially under phosphorus limitation (Egge, 1998), which
 27 corresponds to the high abundance of Group D diatoms and dinoflagellates. In the present study, the
 28 vertical pattern of N: P ratios indicated differences in nutrient composition along the vertical
 29 gradient. The N: P ratio of the surface layer (N: P>16: 1) indicates phosphorus limitation, the
 30 structure of nutrients in the SCM layer changed, and (N: P<16: 1) indicates nitrogen limitation: the
 31 depth continued to increase to the bottom of the euphotic layer and was stable near (N: P=16: 1),

1 indicating that at the bottom of the euphotic layer, with decreasing phytoplankton abundance,
2 interspecific competition is reduced as light limitation kicks in, and the nutrient ratio approaches
3 the Redfield ratio. The differences in nutrient ratios thus affect the vertical distribution patterns of
4 phytoplankton abundance. Diatoms have higher phosphorus requirements than other phytoplankton
5 groups, which may be reflected by the lower N: P ratio in diatoms than in other groups (Hillebrand
6 et al., 2013). Iron is essential for the synthesis of nitrogen-fixing enzymes in *Trichodesmium*, and
7 *Trichodesmium* have a higher demand for iron than other planktonic organisms. The main source of
8 iron in open ocean is atmospheric deposition. Duce et al. showed that the flux of iron deposition is
9 higher in the WPO, so iron is an important environmental limiting factor for the growth of
10 *Trichodesmium* after temperature (Duce and Tindale, 1991). And we suggest that some of the
11 sampled phytoplankton may have recently sunk from the upper layers and therefore represent
12 nutrient rationing and T-S in the water layers. Directly sinking phytoplankton cells are major
13 contributors to surface carbon export and an important component of ocean carbon sink (Boyd and
14 Newton, 1999). The phytoplankton cells can regulate their sinking rates in a variety of ways, such
15 as the physiological state of themselves (Eppley et al., 1967), morphology of themselves (Pitcher et
16 al., 1989), light (Bienfang, 1981) and environmental factors such as temperature and nutrients
17 (Titman and Kilham, 1976).

18 19 4.3 Vertical stratification determined the vertical distribution of phytoplankton

20 The WPO is a oligotrophic area with strong stratification. We found that the interannual
21 variation of phytoplankton was not significant. It remained stably oligotrophic, and the vertical
22 stratification structure determined that of environmental resources such as nutrients, thus forming
23 four contrasting environments, each with its characteristic phytoplankton community structure.
24 Comparative analysis of the phytoplankton community composition of the four groups showed that
25 the phytoplankton was mainly strongly affected by the vertical stratification, which corresponds to
26 previous research (Bouman et al., 2011; Hidalgo et al., 2014; Mojica et al., 2015). Vertical
27 stratification limits the replenishment of nutrients from the deep layer below the thermocline, which
28 affects the N: P ratio, and restricts vertical migration as well as physiologically affecting the vertical
29 structure of phytoplankton growth and mortality (Gupta et al., 2020).

30 In the present study, *Trichodesmium* was the dominant cyanobacterial species. Marine
31 *Trichodesmium* is considered the most critical autotrophic nitrogen-fixing cyanobacteria (Dugdale
32 et al., 1961). *Trichodesmium* can be divided into two forms: clusters and free filaments.
33 *Trichodesmium* thrives in waters above 20 °C, and has a special cellular air sac structure that allows
34 it to move vertically within the upper 100 m of the ocean water column (Laroche et al., 2005). In
35 the process of water blooms formed by *Trichodesmium*, a large amount of nitrogen is often fixed in
36 a relatively short period of time. Therefore, the study of the nitrogen fixation rate of *Trichodesmium*
37 is crucial for estimating the rate of nitrogen fixation in the ocean (Karl et al., 2002). Previous studies
38 have not clarified which factors are the main causes of *Trichodesmium* growth (possibly temperature,
39 wind, iron, phosphorus, etc.) (Capone et al., 1997; Chang et al., 2000; Sañudo-Wilhelmy et al., 2001;
40 Karl et al., 1997). Many researchers have proposed that temperature is the most important factor
41 affecting the growth of *Trichodesmium* (Capone et al., 1999; Kustka et al., 2002). However, we
42 suggest that there is no single positive correlation between temperature and *Trichodesmium* growth,
43 which also is consistent with the study of Chang (2000). In the tropical WPO, where the surface
44 temperatures all exceeded 20 °C, the abundance of *Trichodesmium* in areas with higher temperatures

1 (Groups A and B) was higher than in those with lower temperatures (Groups C and D). Temperature
2 not only directly affected phytoplankton growth, but also indirectly affected phytoplankton growth
3 and abundance by regulating VSI to drive the nutrient ratio (N: P) (Figure 10).

4 Previous models and field experiments have shown that the species composition of
5 phytoplankton communities is significantly affected by vertical turbulent mixing changes (Huisman
6 et al., 2004). A strong coupling exists among the nutrient supply rate, the photosynthetic
7 performance of phytoplankton (Bouman et al., 2006), the phytoplankton biomass and primary
8 production, particularly in eutrophic areas (Richardson et al., 2019). The vertical stratification index
9 reflects the potential effects of vertical stratification on various physical and chemical processes,
10 such as regulating the utilization of light and nutrients in the ocean, which in turn affects
11 phytoplankton dynamics. The results of the present study showed that from the equator to the north,
12 the VSI decreases as the latitude increases, and the phytoplankton community structure changes
13 from cyanobacteria to diatoms. Phytoplankton abundance was significantly different in the water
14 layer above the SCM. The water layer below the SCM tended to be stable. The surface
15 phytoplankton abundance was usually greater than that of the SCM layer, and was related to the
16 surface layer of *Trichodesmium*. Our results demonstrate that the highly stratified region was more
17 suitable for the growth of *Trichodesmium*, while the region with low vertical stratification seems to
18 be more conducive to diatoms and dinoflagellates (Figures 6 and 8). Due to their low mobility and
19 high potential growth rate, diatoms can reproduce rapidly in mixed water with high nutrient content
20 (Tilman et al., 1986). The weak vertical stratification of Group C and D regions (Figure 9b) leads
21 to relative homogeneity of temperature, salinity, density, and nutrients in the upper part of 200 m in
22 the vertical direction (Perez et al., 2006). The vertical distribution of zooplankton has shown that
23 vertical stratification can hinder the migration of small zooplankton populations and indicate
24 different grazing pressures (Mitra et al., 2005; Long et al., 2021). Further research should consider
25 the difference in predation pressure of different zooplankton predators on the composition of the
26 phytoplankton community in different regions. Phytoplankton stratification may cause thin-layer
27 algal blooms and other phenomena, and the influence of phytoplankton stratification can be
28 investigated in further investigated.

30 5 Conclusions

31 This study investigated the phytoplankton community structure of the WPO in the autumn of
32 2016, 2017, and 2018. The WPO is an oligotrophic ocean with a weak water exchange capacity owing
33 to the thermocline and severe stratification in the upper seawater layer. The phytoplankton
34 community structure mainly consisted of cyanobacteria, diatoms, and dinoflagellates, while the
35 abundance of Chrysophyceae was low. In terms of spatial distribution, phytoplankton abundance
36 was high from the equatorial region to 10 °N, and decreased with increasing latitude. Phytoplankton
37 showed a high variation in the vertical distribution. The potential influences of physicochemical
38 parameters on phytoplankton abundance were analyzed by Structural Equation Model (SEM) to
39 determine nutrient ratios driven by vertical stratification to regulate phytoplankton community
40 structure in a typical oligotrophic sea area. Regions with strong vertical stratification (Groups A and
41 B) were more favorable for cyanobacteria, whereas weak vertical stratification (Groups C and D)
42 was more conducive to diatoms and dinoflagellates.

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