## 1 Vertical stratification-driven nutrient ratios regulate phytoplankton

## 2 community structure in the oligotrophic western Pacific Ocean

- 3 Zhuo Chen<sup>1,3</sup>, Jun Sun<sup>2,3\*</sup>, Ting Gu<sup>3</sup>, Guicheng Zhang<sup>3</sup>, Yuqiu Wei<sup>4</sup>
- 4 <sup>1</sup> College of Biotechnology, Tianjin University of Science and Technology, Tianjin 300457, China;
- <sup>2</sup> College of Marine Science and Technology, China University of Geosciences (Wuhan), Wuhan, Hubei 430074, China;
- 6 <sup>3</sup> Research Centre for Indian Ocean Ecosystem, Tianjin University of Science and Technology, Tianjin 300457, China;
- <sup>4</sup> Yellow Sea Fisheries Research Institute, Chinese Academy of Fishery Sciences, Qingdao, 266071, China
- 8 \*Correspondence: <u>phytoplankton@163.com</u>
- 9

10 Abstract: Vertical stratification determines the variability of temperature and nutrient distribution in 11 upper seawater, thereby affecting ocean primary production. Nutrients in the oligotrophic region 12 vary in time and space, and thus, phytoplankton vary in their vertical distribution. However, 13 differences in the vertical distribution of phytoplankton have not been studied systematically. This 14 study investigated the spatial distribution pattern and diversity of phytoplankton communities in the 15 western Pacific Ocean (WPO) in the autumn of 2016, 2017, and 2018 and the local hydrological 16 and nutritional status. The Utermöhl method was used to analyze the ecological characteristics of 17 phytoplankton in the surveyed sea area. In the three cruises investigated, we show universal 18 relationships between phytoplankton and (1) vertical stratification, (2) N:P ratio, and (3) temperature 19 and salinity. The potential influencing factors of physical and chemical parameters on phytoplankton 20 abundance were analyzed using a structural equation model (SEM), which determined that the 21 vertical stratification index was the most important influencing factor affecting phytoplankton 22 abundance and indirectly affecting phytoplankton abundance by dissolved inorganic nitrogen (DIN) 23 and dissolved inorganic phosphorus (DIP). Vertical stratification determines the vertical distribution 24 of the phytoplankton community structure in the WPO. The areas with strong vertical stratification 25 (Groups A and B) are more conducive to the growth of cyanobacteria, and the areas with weak 26 vertical stratification (Groups C and D) are more conducive to the bloom of diatoms and 27 dinoflagellates.

28

29

Keywords: Vertical stratification; phytoplankton community; western Pacific Ocean; N:P ratio

30

31 1. Introduction

32 Phytoplankton account for more than half of global marine primary production and can actively 33 maintain the stability of the entire ecosystem (Sun, 2011). Phytoplankton form the most important 34 level of in the marine food chain (Qian et al., 2005), and changes in their community structure could 35 alter the entire food web. They are widely distributed in various aquatic ecosystems and are marginally affected by fishing activities. Therefore, they are good indicators of marine environment 36 health and climate change (Tang et al., 2017), and studying their community structure is crucial to 37 38 marine ecology. Phytoplankton growth is closely related to nutrient concentration, and in the stable 39 upper water, the thermocline prevents the upward replenishment of nutrients. Therefore, vertical 40 stratification has a substantial effect on phytoplankton.

41

42

As the world's largest and deepest ocean, the Pacific Ocean covers a vast area and has a

1 complex geographic topography, with the deepest trenches on Earth and the highest absolute peaks 2 (Hu et al., 2016). The study area is located in the western Pacific Ocean (WPO) because the 3 equatorial current flows from east to west. Furthermore, warm seawater in the surface layer flows 4 with the current to the WPO, and in the equatorial region, strong solar irradiation heats the seawater 5 year-round. Under the influence of the dual factors, the average temperature of the ocean surface in 6 the WPO region is higher than 28 °C throughout the year. Through heated seawater, radiant heat, 7 and evaporative heat generated by heated seawater, radiative heat, and latent heat, the WPO is higher 8 than the equatorial eastern Pacific by 3–6 °C, and it has a profound impact on global climate change, 9 especially in China and Southeast Asia (Gordon et al., 1996; Hu et al., 2012; Hu et al., 2015). In 10 addition, the surface primary productivity is low, which is typical of oceans with high temperatures, 11 low salinity, and poor nutrition (Messié et al., 2006). The subtropical Pacific is stratified vertically 12 due to typhoons, upwellings, and various physical mixing processes (Emery et al., 1982). It also 13 causes the sea area to have a 100-150 m thermocline, and the high-temperature seawater in the 14 surface layer transfers heat to the atmosphere through sea-air interaction, which generates large 15 disturbances to the atmosphere, which is the area with the most tropical storms and typhoons formed 16 worldwide (Yan et al., 1992; Wang et al., 2012). Perennial tropical storms and typhoons cause heavy 17 rainfall in the sea, thereby reducing the nutrient content in the sea surface layer and restricting the 18 upwelling of nutrient-rich seawater in the lower layer through the thick thermocline layer and 19 making it difficult to reach the upper water column, which has the typical characteristics of high 20 temperature, high salinity, low nutrient, and low primary productivity (Messié et al., 2006; 21 Kawahata et al., 2002). Due to the Coriolis effect, after the trade wind current reaches the WPO, it 22 encounters the barrier of continental topography and changes its path again, some of which joins 23 the Equatorial Counter Current. However, most of it flows along the continental margin toward 24 higher latitudes and becomes the boundary between the nearshore and oceanic water systems. The 25 Philippine waters at 14–15°N are divided into the southward-moving Mindanao Current (MC) and the northward-moving Kuroshio Current (KC) (Dong et al., 2012; Zhao, 2015). The Kuroshio 26 27 Current brings high-temperature and high-salinity seawater from the Pacific Ocean to a wide range 28 of offshore sea areas, which greatly impacts the ocean, meteorology, and hydrology of these sea 29 areas. These impacts could result in the movement of fishing grounds, waxing and waning of sea 30 fog, ice conditions in the Bohai and Yellow seas, and even flooding conditions in eastern China, 31 which are all important and related this phenomena (Christian et al., 2004; Gordon et al., 1996). 32 Phytoplankton brought by the Kuroshio water affect the structure of the biotic community in the 33 nearshore area under the effect of multiple factors. During phytoplankton blooms, the subsurface 34 chlorophyll maximum (SCM) usually occurs near or at the bottom of the light-permeable layer of 35 stable seawater (Yentsch, 1965). SCM distribution is closely related to the depth and intensity of the 36 thermocline, and mixing caused by solar radiation and wind is the driving force for regional 37 consistency and latitudinal differences in the thermocline.

38

Most ocean waters in the global oceans are oligotrophic. With global warming and increased stratification of seawater, these zones are expected to expand, leading to decreases in marine nutrient fluxes and primary productivity (Capotondi et al., 2012; Falkowski et al., 2007; Gruber, 2011). Eutrophic zones with intermittent or irregular nutrient pulses alter the phytoplankton community structure and are ideal for studying changes in phytoplankton community structure dynamics (Lozier et al., 2011; Siokou-Frangou et al., 2010). Changes in seawater stratification and vertical 1 mixing may affect phytoplankton species composition, abundance, size structure, spatial 2 distribution, phenology, and productivity (Behrenfeld et al., 2006; Daufresne et al., 2009; Edwards 3 et al., 2004). These, in turn, affect the function and biogeochemistry of marine ecosystems 4 (Beaugrand et al., 2009; Hoegh-Guldberg et al., 2010). Therefore, studying the ecological and 5 physiological mechanisms that control changes in the phytoplankton community structure within 6 vertical gradients is essential to assess the response of marine systems to global climate change 7 (Richardson et al., 2004).

8

9 Currently, most studies on phytoplankton communities focus on the horizontal distribution at 10 the regional scale, while the vertical stratification of phytoplankton communities has been less 11 studied, and the factors affecting the vertical stratification of phytoplankton remain unclear. The 12 WPO is a typical oligotrophic zone with severe vertical stratification, and seawater stratification has 13 an important influence on the distribution of phytoplankton; therefore, it is necessary to study the 14 vertical stratification of phytoplankton in this region. We investigated how phytoplankton 15 abundance and community composition are related to vertical stratification along a latitudinal gradient in the WPO during 2016–2018. Comparisons between different geographical regions with 16 17 different vertical density distributions offer a unique opportunity to study how phytoplankton 18 dynamics change as stratification develops.

19

20 2. Materials and methods

21 2.1. Study area and sampling

This study relied on the shared voyage of the WPO (0–20 °N, 120–130 °E), commissioned by the National Natural Science Foundation of China. Physical, biological, chemical, and geological surveys were carried out on the RV "*kexue*" from September to November in 2016, 2017, and 2018. The sampling stations used in this study are shown in Figure 1; the sampling layers were 5, 25, 50, 75, 100, 150, and 200 m. Phytoplankton samples from different water layers were placed in 1 L PE bottles, fixed in formaldehyde solution (3%), and stored in dark. Nutrient samples from different layers were placed in PE bottles, frozen, and stored at –20 °C for laboratory nutrient analysis.



Figure 1. Stations in the western Pacific Ocean (WPO) of three cruises. (a): Current systems of the
WPO; (b), (c), and (d): sampling stations of 2016, 2017 and 2018 cruises, respectively. The station
at 130°E forms the section A, and the station at 20°N forms the section B. Map of the WPO shows
the major geographic names and the surface currents, including the Subtropical Counter Current
(STCC), the North Equatorial Current (NEC), the Northern Equatorial Counter Current (NECC),
the South Equatorial Current (SEC), the New Guinea Coastal Current (NGCC), the Mindanao
Current (MC), the Mindanao Eddy (ME), the Halmahera Eddy (HE).

1

10 2.2. Identification of Phytoplankton

11 After returning to the laboratory, the Utermöhl method was applied for phytoplankton analysis. 12 A 1 L subsample was allowed to stand for 48 h; then 800 mL supernatant was removed carefully by 13 siphoning through a catheter, taking care to prevent the catheter from touching the bottom of the 14 bottle. Thereafter, the remaining 200 mL liquid was gently mixed and half of which was further 15 concentrated with a 100 mL sedimentation column (Utermöhl method) for 48 h sedimentation. The phytoplankton species were identified and enumerated under an inverted microscope (AE2000, 16 17 Motic, Xiamen, China) at 400× (or 200×) magnification. Phytoplankton identification was 18 conducted as described by Jin et al. (1965), Isamu Y (1991), and Sun et al. (2002). The World 19 Register of Marine Species (http://www.marinespecies.org). Species identification was as close as 20 possible to the species level.

21

22 2.3. Nutrient Analysis

23 The AA3 (SEAL, German) was used for the analysis and determination nutrient. Soluble

inorganic phosphorus (PO<sub>4</sub>-P) was determined by the phosphomolybdenum blue method with the 1 limit of detection of 0.02 µmol L<sup>-1</sup>; dissolved silicate (SiO<sub>3</sub>-Si) was determined by the silicon 2 molybdenum blue method with the limit of detection of 0.02 µmol L<sup>-1</sup>; nitrate (NO<sub>3</sub>-N) was 3 determined by the cadmium column method with the limit of detection of 0.01 µmol L<sup>-1</sup>; nitrite 4 5 (NO<sub>2</sub>-N) was determined by the naphthalene ethylenediamine method with the limit of detection of 6 0.01 µmol L<sup>-1</sup> (Dai et al., 2008). Ammonia (NH<sub>4</sub>-N) was determined by the sodium salicylate 7 method with the limit of detection of 0.03 µmol L<sup>-1</sup> (Guo et al., 2014; Pai et al., 2001). Nitrogen-tophosphorous (N:P) ratio was calculated by dividing nitrogen concentration (NO3<sup>-</sup>+NO2<sup>-</sup>) by 8 9 phosphate concentration.

10

14

11 2.4. Analysis and methods

12 A SBE911 CTD sensor and standard Sea-Bird Electronics methods were used to process 13 recorded hydrological parameters. The depth of the mixed layer (ML) is calculated as

$$(S, T) = (Sref, Tref-\Delta T)$$

15 S and T are the average salinity and temperature, respectively, and Sref and Tref are the temperature 16 and salinity at 5 m,  $\Delta$ T is equal to 0.5 °C.

We calculated the vertical stratification index (VSI) to indicate the degree of verticalstratification of the water column:

VSI= $\Sigma [\delta_{\theta}(m+1) - \delta_{\theta}(m)]$ 

20 where  $\delta_{\theta}$  is the potential density anomaly, and m is the depth from 5 to 200 m.

The abundance of phytoplankton cells in water column was calculated through the trapezoidal
 integral method (Zhu et al., 2019):

23 
$$P = \left\{ \sum_{i=l}^{n-l} \frac{P_{i+l} + P_i}{2} (D_{i+l} - D_i) \right\} / (D_n - D_1)$$

where P is the average value of phytoplankton abundance in water column, P*i* is the abundance value of phytoplankton in layer *i*, i + 1 is the layer i + 1, Dn is the maximum sampling depth, D*i* is the depth of layer *i*, and n is the sampling level.

27

28 We clustered all species based on Bray-Curtis similarity distance for three years, and the results 29 showed four distinct regions using the Primer (version 6). Distance-based Redundancy analysis (db-30 RDA) and Principal Co-ordinates Analysis (PCoA) were performed using the R package vegan 31 (version 2.5-7) (Oksanen et al., 2020) to explain the relationship between the environmental 32 parameters (temperature, salinity, depth, VSI, Dissolved inorganic nitrogen (DIN) and Dissolved 33 inorganic phosphorus (DIP) and Dissolved silicate (DSi)) and phytoplankton community structure. 34 The results were visualized using the R package ggplot2 (version 3.3.2). SEM was used to assess 35 the relative direct and indirect impact of physical and chemical parameters on phytoplankton abundance. The chi-square test ( $\chi^2$ ), comparative fit index (CFI), and goodness fit index (GFI) were 36 used to assess the model fit. 37

38

39 3. Results

40 3.1 Hydrographic features of the study area during the sampling years

The surface temperature and salinity of the surveyed sea area in 2016, 2017, and 2018 are shown in Figure 2. In general, the temperature increased with decreasing latitude, and the stations

1 near the equator exhibited the highest temperature; in constrast, the salinity showed an opposite 2 trend as that of temperature, with a high value from 15 °N to 20 °N. The surface temperature (Fig. 3 2) of the surveyed area in 2016 ranged from 28.58 °C (station 16-B1) to 30.14 °C (station 16-A16), with an average of 29.43 °C. The surface salinity (Fig. 2) of the surveyed area in 2016 ranged from 4 5 33.80 (station 16-B2) to 34.65 (station 16-A2), with an average of 34.32. The surface temperature 6 (Fig. 2) of the surveyed area in 2017 ranged from 27.91 °C (station 17-A4) to 30.19 °C (station 17-7 A20), with an average of 29.26 °C. The surface salinity (Fig. 2) of the surveyed area in 2017 ranged 8 from 33.38 (station 17-A16) to 34.64 (station 17-B4), with an average of 33.94. The surface 9 temperature (Fig. 2) of the surveyed sea area in 2018 ranged from 26.33 °C (station 18-B1) to 10 29.79 °C (station 18-A17), with an average of 28.83 °C. The surface salinity (Fig. 2) of the surveyed 11 sea area in 2018 ranged from 33.77 (station 18-A14) to 34.64 (station 18-B1), with an average of 12 34.21.

13

14 The profile distribution of temperature and salinity based on the cross-sectional data of 15 different water layers at each station obtained from the survey is shown in Figure 2. The temperature of the shallow water column (0-100 m) is higher than that of the deep-water column (100-200 m). 16 17 The salinity values of the deep-water bodies (100-200 m) were higher than those of the shallow 18 water bodies (0-100 m). The values of temperature and salinity in 2016, 2017, and 2018 did not 19 change significantly. The temperature of the section in 2016 ranged from 12.16 °C (200 m at station 20 16-A11) to 30.14 °C (5 m at station 16-A16), with an average of 25.74 °C. The salinity of the section 21 in 2016 ranged from 33.80 (5 m at station 16-B2) to 35.39 (150 m at station 16-A16), with an average of 34.61 °C. The temperature of the section in 2017 ranged from 11.16 °C (200 m at station 22 23 17-A13) to 30.19 °C (5 m at station 17-A20), with an average of 25.18 °C. The salinity of the section 24 in 2017 ranged from 33.38 (5 m at station 17-A16) to 35.24 (150 m at station 17-A23), with an 25 average of 34.46. The temperature of the section in 2018 ranged from 9.65 °C (200 m at station 18-A14) to 29.79 °C (5 m at station 18-A17), with an average of 24.22 °C. The salinity of the section 26 27 in 2018 ranged from 33.77 °C (5 m at station 18-A14) to 35.39 °C (150 m at station 18-A17), with 28 an average of 34.57.



2 3

5

1

Figure 2. The temperature and salinity distribution in the WPO from three cruises. (a–c) surface temperature in 2016, 2017, 2018 respectively, (d–f) surface salinity in 2016, 2017, 2018 respectively, (g–i) vertical distribution of temperature in 2016, 2017, 2018 respectively, (j–l) vertical distribution of salinity in 2016, 2017, 2018 respectively.

6 7

8 The distribution of the VSI in latitude for the three cruises is shown in Figure 3. Overall, the 9 VSI showed the same distribution pattern in the three cruises, with the highest value occurring at 7– 10 8 °N and a decreasing trend with increasing latitude. In the 2016 cruise (Figure 3-a), the minimum 11 value of VSI (2.54) appeared in the station at 20 °N (station 16-B4), and the maximum value (4.94) 12 appeared in the station at 7 °N (station 16-A11), with an average of  $3.90 \pm 0.76$ . In the 2017 cruise 13 (Figure 3-b), a minimum value of VSI (2.85) appeared in the station at 18 °N (station 17-B4), and

- 1 the maximum value (5.54) appeared in the station at 7 °N. The maximum value (5.54) occurred in
- 2 the station at 7 °N (station 17-A14) with an average of  $4.30 \pm 0.82$ . In the 2018 cruise (Figure 3-c),
- 3 the minimum value of VSI (2.50) occurred in the station at 18 °N (station 18-B1), and the maximum
- 4 value (5.48) occurred in the station at 8 °N (station 18-A14) with an average of  $4.01 \pm 0.95$ .
- 5 Interestingly, the variation in VSI varied significantly across latitudinal regions; the VSI was high
- 6 from the equator to 10 °N, while it was low at 10–20 °N.



Figure 3. Linear fits of the vertical stratification index with latitude (a) in 2016, (b) in 2017, (c) in
2018. The black dots are the VSI of each station.

7

12 3.2. Interannual variability of phytoplankton communities

13 Figures 4a, b, and c show the horizontal distribution of surface phytoplankton abundance from 14 2016 to 2018. The interannual variation in phytoplankton was relatively stable, and the sampling 15 area and sampling time from 2016 to 2018 were generally consistent. Most phytoplankton species 16 showed a relatively uniform distribution. Phytoplankton distribution showed a trend of decreasing 17 abundance from the equator to the north, extending in latitude, especially between the equator to 18 10 °N, where high values of abundance were concentrated. The abnormally high phytoplankton 19 abundance in this region is associated with the predominance of Trichodesmium. However, affected 20 by coastal currents, high abundance patches dominated by diatoms were observed in the Luzon 21 Strait area in southern Taiwan, which were carried to the surface by upwelling currents and 22 accounted for more than 67.76% of the abundance at this station. Relatively high abundances were 23 observed at stations in the Kuroshio extension region, consisting mainly of cosmopolitan and warm 24 water species. Phytoplankton abundance was the lowest in the high latitude region.





Figure 4. Horizontal distribution of phytoplankton abundance in the WPO. a. 2016, surface layer; b.

- 27 2017, surface layer; and c. 2018, surface layer.
- 28



1 Figure 5 shows the vertical distribution of the phytoplankton. As can be seen from the figure, 2 the overall trend in the WPO was consistent across the three cruises in 2016 (a), 2017 (b), and 2018 3 (c), with the phytoplankton distribution showing regional variations in latitude and differences in vertical distribution at depth. In terms of latitude, high phytoplankton value areas were concentrated 4 5 near the equator (0 °E–8 °E), and phytoplankton abundance gradually decreased with increasing 6 latitude. Vertical distribution of phytoplankton indicated that the plankton-abundant areas occurred 7 from 0-50 m, and the phytoplankton abundance gradually decreased with the increase in depth. 8 Vertical distribution of phytoplankton abundance differed significantly across different areas. In the 9 areas near the equator affected by Halmahera Eddy (HE) and Mindanao Eddy (ME), phytoplankton 10 abundance was mainly concentrated in the upper water column (0-50 m) and consisted mainly of 11 cyanobacteria. In the northern area affected by Kuroshio (KC), the phytoplankton abundance was 12 lower than that in the equatorial stations, while the phytoplankton species composition was mostly 13 dominated by diatoms and dinoflagellates.



14

Figure 5. Vertical distribution of phytoplankton abundance (Log10 cells  $L^{-1}$ ) in the WPO in 2016 (a); 2017 (b) and 2018 (c).

17

18 3.4. Phytoplankton community structure

19 Since there was little difference in interannual changes between species, we clustered all 20 species based on Bray-Curtis similarity distance for stations, and the results showed four distinct 21 regions (Figure 6). Cluster analysis divided the phytoplankton communities at the sampling sites for 22 three years into four groups. Cyanobacteria (>90%) were the dominant species in groups A and B. 23 The ratio of diatoms to dinoflagellates in Group A (4.8) was higher than that in Group B (1.4). 24 Cyanobacteria were the dominant (66%) phytoplankton at the stations of Group C, while diatoms 25 (18%) and dinoflagellates (14%) constituted 32% of the population in this group. Diatoms (43%) 26 and dinoflagellates (49%) dominated the stations in Group D, accounting for approximately 92% of 27 the total phytoplankton. The proportion of Chrysophyceae was low in all four groups (Table 1). The 28 dendrogram showed that these populations were grouped into four groups, which were essentially identical to those determined by PCoA analysis (Figure 7). The horizontal and vertical axes explain 29 30 51.87% and 21.41% of the phytoplankton community structure, respectively.



Figure 6. Bray-Curtis similarity-based dendrogram showing averaged phytoplankton community
 composition and abundance for each station across the 3 cruises. For each station, community
 composition is indicated with bar plots, and phytoplankton abundance is represented with black bars.

Table 1. The percentages (%) (average ± standard deviations) of diatoms, dinoflagellates,
cyanobacteria and Chrysophyceae in the four groups.

5	5 1 5	υī		
Species	Group A	Group B	Group C	Group D
Diatoms	1.09±0.79	4.25±1.57	21.83±11.45	43.71±10.12
Dinoflagellates	0.44±0.42	3.41±3.30	17.26±12.45	48.38±11.61
Cyanobacteria	98.45±1.10	92.08±4.79	59.05±20.38	6.06±4.93
Chrysophyceae	0.02±0.01	0.26±0.10	1.86±1.99	1.85±1.66



Figure 7. Principal Coordinates Analysis for groups. Triangles, circles, and squares represent 2016,
2017, and 2018 stations, respectively. P < 0.05. Different colors represent different groups.</li>
Percentages of total variance are explained by coordinates 1 and 2, accounting for 51.87% and
21.41%, respectively.

6

## 7 3.5. Relationships between phytoplankton and environmental factors

8 The relationship between phytoplankton and environmental factors was analyzed using RDA. 9 We obtained a two-dimensional distribution map of the species, sample distribution, and 10 environmental factors (Figure 8). The results showed that different phytoplankton classes were 11 correlated differently with environmental factors. Cyanobacteria showed negative correlations with 12 temperature and salinity and positive correlations with VSI and nutrient concentration, indicating 13 that waters with high VSI are suitable for the growth of cyanobacteria (mostly Trichodesmium). 14 Diatoms and methanogens showed the opposite trend, exhibiting positive correlations with 15 temperature and salinity and negative correlations with VSI and nutrient concentration, indicating 16 that diatoms and dinoflagellates prefer waters with low VSI.

17

18 There were four distinct phytoplankton communities in the WPO: Group A was distributed in 19 the equatorial region with clear vertical stratification. This community is characterized by high 20 abundance and is dominated by Trichodesmium species such as T. thiebautii, T. hildebrandtii, and 21 T. erythraeum, which are positively correlated with high concentrations of DIN, phosphate, and 22 silicate. Group B was located near 8°N and is mainly influenced by the NECC and mesoscale eddy 23 influence; the phytoplankton community was represented by warm water species, similar to that of 24 Group A. Group C was mainly distributed in the 15 °N region and was strongly influenced by the 25 NEC. Group D was mainly distributed in the 20°N region, where it was directly influenced by the 26 Kuroshio Current; the phytoplankton community was positively correlated with temperature and 27 salinity.



Figure 8. Redundancy analysis of the (a) phytoplankton and environmental parameters, (b) groups
and environmental parameters in the WPO. Colored dots represent sampling sites, triangles
represent phytoplankton species, and arrows represent environmental factors.

5

6

3.4 Temperature, salinity, and vertical stratification index

7 The temperature, salinity, and VSI of the four groups are shown in Figure 9. The temperature 8 and salinity (T-S) box diagram of the sample from 5 m above depicts the four main water masses in 9 the WPO. Groups A (average 29.8 °C) and B (average 29.6 °C) had high temperatures, but the 10 salinities of Groups A (average 33.9 °C) and B (average 33.8 °C) was low. The temperature of Groups C (average 28.9 °C) and D (average 28.9 °C) was low, but the salinity of Groups C (average 11 12 34.2) and D (average 34.4) was high (Fig.9-a). The strong spatial variability of T-S was evident from 13 the characteristics of salinity and temperature. We also calculated the vertical stratification index of the four groups (Fig.9-b). Compared with Groups C (average 3.86) and D (average 3.54), the number 14 15 of VSIs in Groups A (average 4.69) and B (average 4.86) was markedly higher, and Group A had 16 the highest VSI. There were obvious differences between the four groups; that is, the stratification 17 of the first two groups was more pronounced (Table 2).

The vertical stratification index was linearly fitted to temperature (Fig.9-a) and salinity (Fig.9b). The fitting results show that the temperature is positively correlated with the vertical stratification index. The VSI of all groups was negatively correlated with salinity. It can be noted that the changes in temperature and salinity were more pronounced in the vertical direction. In Groups A and B with a high stratification index, the changes in temperature and salinity within the group were small. However, the temperature and salinity changed significantly within Groups C and D, with a small stratification index.



1

2 Figure 9 Surface temperature and salinity (a), and vertical stratification index (b) of the four groups.

5

4 Table 2. Average (±standard deviations) values for nutrients (µmol L<sup>-1</sup>), temperature (°C), salinity

for each phytoplankton community group were identified by the cluster analysis in the WPO.					
	Group A	Group B	Group C	Group D	
Temperature	25.30±1.06	24.45±1.85	24.92±1.32	25.41±1.23	
Salinity	34.45±0.14	34.40±0.07	34.56±0.16	34.68±0.20	
DIP	$0.28 \pm 0.07$	0.18±0.13	0.16±0.13	0.13±0.10	
DIN	4.49±1.76	5.43±2.71	2.62±1.89	$1.80{\pm}1.08$	
DSi	2.93±1.05	4.13±2.15	$1.90 \pm 1.47$	1.44±0.95	
VSI	4.69±0.39	4.86±0.45	3.86±0.84	$3.54 \pm 0.82$	

6

7 3.6. Direct vs. indirect effects of environmental parameter on phytoplankton abundance

The causal relationships between measured phytoplankton abundance and relevant physical 8 9 and chemical parameters were examined using SEM, using interactions between temperature, 10 salinity, VSI, DIN, and DIP (Fig.10), as theoretical and experimental data indicated the importance 11 of these variables. The model results showed that temperature, DIP, and DIN had a direct effect on phytoplankton abundance, with temperature having the largest direct effect on phytoplankton 12 13 abundance (0.38), followed by DIN (0.28) and DIP (0.24). Temperature, salinity, and VSI had 14 indirect effects on phytoplankton abundance, with temperature and salinity having negative indirect 15 effects on phytoplankton abundance (-0.17 and -0.30) and VSI having positive indirect effects (0.31) 16 (Figure 10). From the results of the total effect, only salinity had a negative effect on phytoplankton 17 abundance (-0.30), while both temperature and VSI had positive effects on phytoplankton 18 abundance (0.20 and 0.312), with VSI having the largest total effect. Although the direct effect of 19 temperature on phytoplankton abundance was significant, it was partially offset by the indirect negative effect, while VSI had no direct effect on phytoplankton abundance, but its larger indirect 20 21 effect resulted in the largest total effect. Both DIN and DIP had positive effects on phytoplankton 22 abundance, but the effect of DIN was greater. Since the vertical distribution of DIN and DIP 23 exhibited stronger variability, more specific analyses of DIN and DIP will be conducted later.



Chi-square=8.385, p=0.211, CFI=0.989, GFI=0.963

1

Figure 10. Structural Equation Model (SEM) analysis examining the effects of temperature, salinity,
 VSI, DIN and DIP on phytoplankton abundance. Solid black and red lines indicate significant
 positive and negative effects at p < 0.05, black and red dashed lines indicated insignificant effects.</li>
 R<sup>2</sup> values associated with response variables indicate the proportion of variation explained by
 relationships with other variables. Values associated with arrows represent standardized path
 coefficients.

8

9 We analyzed the N:P ratio of the surface layer, SCM, and 200 m. The N:P ratio in the surface 10 layer (N:P>16:1) indicates phosphorus limitation, which is consistent with the SEM analysis 11 (Fig.11). The trophic structure of the SCM layer changed, N:P <16:1 indicated nitrogen limitation, 12 and the depth continued to increase to the bottom of the euphotic layer and stabilized around N:P 13 =16:1, indicating that at the bottom of the euphotic layer, as phytoplankton abundance decreased 14 and interspecific competition decreased, the trophic ratio approached the Redfield ratio.





17 m. The dashed line indicates the Redfield ratio N:P = 16:1.

- 18
- 19 4. Discussion
- 20 4.1. Comparison with historical data

1 Kuroshio and WPWP are key areas of the WPO sea-air interaction (Zhang, 1999). Previous 2 surveys have provided less knowledge of the phytoplankton community structure in this study area (Table 3). Previously, samples were collected by net, and net-collected samples reduced 3 4 phytoplankton abundance in small volumes, thereby underestimating the phytoplankton abundance 5 in the ocean under investigation. In the present study, phytoplankton samples were collected from 6 water samples, which better reflected the phytoplankton community structure and abundance. Sun 7 et al. (1997) and Liu et al. (1997) further investigated the species composition and abundance 8 distribution of phytoplankton diatoms and dinoflagellates in the Ryukyu Islands and nearby waters. 9 Li et al. (2008) conducted a study on phytoplankton in the tropical and subtropical Pacific oceanic 10 zones with response mechanisms to the limitation of nitrogen and iron. Chen et al. (2018) 11 investigated the phytoplankton community structure and mesoscale eddies in the western boundary 12 current. A total of 199 species in 61 genera belonging to four phytoplankton families were identified, 13 among which the abundance of Trichodesmium species was high. Previous studies have mostly 14 focused on the vertical trawl and the horizontal distribution of phytoplankton throughout the water 15 column while ignoring the effect of vertical stratification on phytoplankton.

16

17 Table 3. Historical data of the phytoplankton commu	nity in the WPO.
--	------------------

Date	Sampling areas	Layer /m	Number of species	Sampling types	References
2018.10	2°–20°N, 120°–130°E	0–200	305	Water samples	This study
2017.10	2°–18°N, 126°–130°E	0–200	339	Water samples	This study
2017.08	10.3°–10.9°N, 139.8°–140.4°E	0–200	147	Water samples	Dai et al.,2020
2017.08	21°-42°N, 118°-156°E	0–200	235	Water and net samples	Lin et al., 2020
2017.05	21°-42°N, 118°-156°E	0–200	248	Water and net samples	Lin et al., 2020
2016.09	2°–21°N, 127°–130°E	0–200	269	Water samples	This study
2016.09	0°–20°N, 120°–130°E	0–200	243	Net samples	Chen et al., 2018b
2014.08	0°–21.5°N, 121°–135.5°E	0–300	199	Net samples	Chen et al., 2018a
1997.07	23°30′–29°30′N,	0–200	227	Net samples	Sun et al., 2000
	122°30′–130°30′E				
1997.07	23°30′–29°30′N,	0–200	251	Net samples	Liu et al., 2000
	122°30′–130°30′E				

18 19

4.2. Relationship between N:P ratio and vertical distribution of phytoplankton

20 Research on the factors that control the structure of the phytoplankton community has been 21 carried out for decades, but the hypothesis of nutrient concentration limits and ratios has not been 22 fully explained in terms of affecting the structure of the phytoplankton community (Gao et al., 2019). 23 As diatoms and dinoflagellates show great differences in cell morphology, structure, and nutrition 24 mode, they differ greatly in their acquisition of nutrient strategies. Several studies have revealed 25 that dinoflagellates have the ability of mixotrophy, and the mixotrophic modes of dinoflagellates include direct engulfment of prey, peduncle feeding, and pallium feeding, and phosphorus limitation 26 27 is a common factor stimulating dinoflagellates to ingest particulate nutrients (Huang et al., 2005; 28 Smayda, 1997; Stoecker, 1999). The variation in phytoplankton community structure was always correlated with fluctuations in physicochemical environmental parameters. 29

30

31

In the four groups we studied, surface seawater N:P>16:1 indicated that phosphorus in surface

1 seawater was limited, but Trichodesmium relied on its own nitrogen fixation function and was highly 2 abundant in oligotrophic waters (Fig.11). The relationship between Trichodesmium and nitrogen 3 fixation has been demonstrated several years ago (Grosskopf et al., 2012; Luo et al., 2012; Zehr, 4 2011). The presence of slight nitrogen limitation in surface seawater in Group D was consistent with 5 the low abundance of Trichodesmium, which was consistent with studies on the abundance of 6 Trichodesmium in the region (Chen et al., 2019; Sohm et al., 2011). In the WPO, the most 7 oligotrophic ocean around the world (Hansell et al., 2000), nutrients have become an important 8 factor that determines the distribution of phytoplankton. Under nutrition-limited conditions, diatoms 9 and dinoflagellates were more susceptible, especially under phosphorus limitation (Egge, 1998), 10 which corresponds to the high abundance of Group D diatoms and dinoflagellates. In the present 11 study, the vertical pattern of N:P ratios indicated differences in nutrient composition across the 12 vertical gradient. The N:P ratio of the surface layer (N:P>16:1) indicated phosphorus limitation, the 13 structure of nutrients in the SCM layer changed, and (N:P<16:1) indicated nitrogen limitation: the 14 depth continued to increase to the bottom of the euphotic layer and was stable near (N:P=16:1), 15 indicating that at the bottom of the euphotic layer, with decreasing phytoplankton abundance, 16 interspecific competition reduced and the nutrient ratio approached the Redfield ratio. The 17 differences in nutrients partly affected the vertical distribution patterns of phytoplankton abundance. 18 Diatoms have higher phosphorus requirements than other phytoplankton groups, which may be 19 reflected by the lower N:P ratio in diatoms than in other groups (Hillebrand et al., 2013).

20 21

## 4.3. Vertical stratification determined the vertical distribution of phytoplankton

22 With global climate change, marine oligotrophic regions continue to expand, and seawater 23 stratification is intensified, which is the main problem affecting the marine phytoplankton 24 community structure. The WPO is a typical oligotrophic area with severe stratification. We found 25 that the interannual variation of phytoplankton in stable oligotrophy was not significant, and the 26 intensity of vertical stratification adapted to different environmental changes (nutrients, temperature, 27 and salinity), thus forming four contrasting environments with varying degrees of limiting the 28 community structure of phytoplankton. Comparative analysis of the phytoplankton community 29 composition of the four groups showed that the phytoplankton was mainly strongly affected by the 30 vertical stratification, which corresponds to previous research (Bouman et al., 2011; Hidalgo et al., 31 2014; Mojica et al., 2015). Vertical stratification limits the replenishment of nutrients in the deep 32 layer and aggravates the formation of the thermocline, which affects the N:P ratio, thereby 33 restricting vertical migration of phytoplankton or affecting the physiology of heat-driven 34 phytoplankton growth and mortality variety (Gupta et al., 2020).

35

36 In the present study, Trichodesmium was the dominant cyanobacterial species. Marine 37 Trichodesmium has been considered the most critical autotrophic nitrogen-fixing cyanobacteria 38 since the 1960s (Dugdale et al., 1961). Trichodesmium can be divided into two forms: clusters and 39 free filaments. Trichodesmium is suitable for living in waters above 20 °C, and has a special cellular 40 air sac structure that allows it to move vertically within the upper 100 m of the ocean water column 41 (Laroche et al., 2005). In the process of water blooms formed by Trichodesmium, a large amount of 42 nitrogen is often fixed in a relatively short period of time. Therefore, the study of the nitrogen 43 fixation rate of *Trichodesmium* is crucial for estimating the rate of nitrogen fixation in the ocean 44 (Karl et al., 2002). Previous studies have not clarified which factors are the main causes of

1 Trichodesmium growth (possibly temperature, wind, iron, phosphorus, etc.) (Gobler, 2001; Karl et 2 al., 1997; Capone et al., 1997; Chang et al., 2000). Many researchers believe that temperature is the 3 most important factor affecting the growth of Trichodesmium (Capone et al., 1999; Adam et al., 4 2002). However, we believe that there is no single positive correlation between temperature and 5 Trichodesmium growth, which is consistent with the study of Chang (2000). In the tropical WPO, 6 where the temperature was not restricted, the abundance of Trichodesmium in areas with higher 7 temperatures (Groups A and B) was higher than that at relatively low temperatures (Groups C and 8 D). However, when temperature was no longer the main limiting factor for the growth of 9 Trichodesmium, the abundance of Trichodesmium in the higher temperature Group A was not higher 10 than the relatively lower temperature Group B as expected, and the high-value area of 11 Trichodesmium appeared in Group B. For this reason, we believe that the vertical stratification of 12 Group B was the highest, and the abundance of *Trichodesmium* was also the highest, and the vertical 13 stratification of the four groups was in the following order: Group B > A > C > D (Fig. 9), which 14 was consistent with the high and low abundance of *Trichodesmium* in the four groups. SEM analysis 15 (Fig. 10) revealed that vertical stratification does not directly affect phytoplankton abundance, but indirectly affects phytoplankton growth and abundance by driving the nutrient ratio (N:P). Therefore, 16 17 we suggest that temperature is not the main factor limiting phytoplankton growth in tropical 18 oligotrophic waters but rather drives nutrient ratios through vertical stratification, which affects 19 phytoplankton growth. This is reflected by the fact that growth of cyanobacteria, diatoms, and 20 dinoflagellates is higher in areas with severe vertical stratification, whereas diatoms and 21 dinoflagellates exhibited superior growth in areas with weak vertical stratification.

22

23 Previous models and field experiments have shown that the species composition of 24 phytoplankton communities is significantly affected by vertical turbulent mixing changes (Huisman 25 et al., 2004). A strong coupling exists between the nutrient supply rate and the photosynthetic performance of phytoplankton (Bouman et al., 2006) and phytoplankton biomass and primary 26 27 production in eutrophic areas are high (Richardson et al., 2019), which directly limits nutrient supply. 28 The vertical stratification index reflects the potential causes of vertical stratification in various 29 physical and chemical processes, such as regulating the utilization of light and nutrients in the ocean, 30 which in turn affects phytoplankton dynamics. The results of the present study showed that from the 31 equator to the north, the VSI decreases as the latitude increases, and the phytoplankton community 32 structure changes from cyanobacteria to diatoms. Phytoplankton abundance was significantly 33 different in the water layer above the SCM. The water layer below the SCM tended to be stable. The 34 surface phytoplankton abundance was usually greater than that of the SCM layer, which was related 35 to the surface layer of Trichodesmium. Our results demonstrated that the highly stratified region was more suitable for the growth of Trichodesmium, while the region with low vertical stratification 36 37 seems to be more conducive to the survival of diatoms and dinoflagellates (Fig. 6 and 8). Due to 38 their poor activity and high potential growth rate, diatoms can reproduce rapidly in the circulation 39 and water with high nutrient content (Tilman et al., 1986). The weak vertical stratification of Group 40 C and D regions leads to the homogeneity of temperature, salinity, density, and nutrients in the upper 41 part of 200 m in the vertical direction. The frequency and abundance of dinoflagellates in Groups C 42 and D were higher, which is consistent with the environment where they are more inclined to vertical 43 stratification and weaker (Perez et al., 2006). The vertical distribution of zooplankton has shown 44 that vertical stratification can hinder the migration of small zooplankton populations and indicate

different grazing pressures (Long et al., 2021; Aditee et al., 2005). Further research should consider the difference in predation pressure of different zooplankton predators on the composition of the phytoplankton community in different regions. Phytoplankton stratification may cause thin-layer algal blooms and other phenomena, which was not discussed in this article, and the influence of phytoplankton stratification can be investigated in future studies.

- 6
- 7 5. Conclusions

8 This study investigated the phytoplankton community structure of the WPO in the autumn of 9 2016, 2017, and 2018. The WPO is a typical oligotrophic ocean with a weak water exchange 10 capacity owing to the thermocline and severe stratification in the upper seawater layer. The 11 phytoplankton community structure mainly consisted of cyanobacteria, diatoms, and dinoflagellates, 12 while the abundance of Chrysophyceae was low. In terms of spatial distribution, phytoplankton 13 abundance was high from the equatorial region to 10 °N, and decreased with increasing latitude. 14 Phytoplankton showed a high variability in the vertical distribution. The potential influences of 15 physicochemical parameters on phytoplankton abundance were analyzed by SEM to determine 16 nutrient ratios driven by vertical stratification to regulate phytoplankton community structure in a 17 typical oligotrophic. Regions with strong vertical stratification (Groups A and B) were more 18 favorable for cyanobacteria, whereas weak vertical stratification (Groups C and D) was more 19 conducive to diatoms and dinoflagellates.

20

Funding: This research was financially supported by the National Key Research and Development
Project of China (2019YFC1407805), the National Natural Science Foundation of China (41876134,
41676112 and 41276124), the Tianjin 131 Innovation Team Program (20180314), and the
Changjiang Scholar Program of Chinese Ministry of Education (T2014253) to Jun Sun.

25

Acknowledgments: Thank the Natural Science Foundation for its support of the Northwest Pacific voyage for sampling and field experiments. Samples were collected onboard of R/V *Kexue* implementing the open research cruise (voyage number: NORC2016-09, NORC2017-09 and NORC2018-09) supported by NSFC Shiptime Sharing Project. Thank you to all the staff of "*Kexue*" for their help. Thanks for the CTD data provided by Dongliang Yuan Physical Oceanography Research Group, Institute of Oceanography, Chinese Academy of Sciences.

- 32
- 33 References
- Aditee, M. and Kevin, F. J.: Predator-prey interactions: is 'ecological stoichiometry' sufficient when
   good food goes bad?, Journal of Plankton Research, 27 (5), 393-399, doi:
   10.1093/plankt/fbi022, 2005.
- Beaugrand, G.: Decadal changes in climate and ecosystems in the North Atlantic Ocean and adjacent
  seas, Deep-Sea Research Part II, 56 (8-10), 656-673, doi: 10.1016/j.dsr2.2008.12.022, 2009.
- Behrenfeld, M. J., O'Malley, R. T., Siegel, D. A., McClain, C. R., Sarmiento, J. L., Feldman, G. C.,
  Milligan, A. J., Falkowski, P. G., Letelier, R. M., and Boss, E. S.: Climate-driven trends in
  contemporary ocean productivity, Nature, 444 (7120), 752-755, doi: 10.1038/nature05317,
  2006.
- Bouman, H. A., Ulloa, O., Barlow, R., Li, W., Platt, T., Zwirglmaier, K., Scanlan, D. J., and
  Sathyendranath, S.: Water-column stratification governs the community structure of

- subtropical marine picophytoplankton. Environmental microbiology reports, 3, 473-482, doi:
   10.1111/j.1758-2229.2011.00241.x, 2011.
- Bouman, H. A., Ulloa, O., Scanlan, D. J., Zwirglmaier, K., Li, W., Platt, T., Stuart, V., Barlow, R.,
  Leth, O., Clementson, L., Lutz, V., Fukasawa, M., Watanabe, S., and Sathyendranath, S.:
  Oceanographic basis of the global surface distribution of Prochlorococcus ecotypes, Science,
  312 (5775), 918-921, doi: 10.1126/science.1122692, 2006.
- Capone, D., Carpente, E.: Nitrogen fixation by marine cyanobacteria: historical and global
   perspectives, Bulletin De Linstitut Océanographique, 19: 235-256, 1999.
- 9 Capone, D. G., Zehr, J. P., Paerl, H. W., Bergman, B., Carpenter, E. J.: Trichodesmium, a globally
  10 significant marine Cyanobacterium, Science, 276: 1221-1229, doi:
  11 10.1126/science.276.5316.1221, 1997.
- Capotondi, A., Alexander, M., Bond, N., Curchitser, E., and Scott, J.: Enhanced upper ocean
  stratification with climate change in the CMIP3 models, Journal of Geophysical Research
  (Oceans), 117, 4031, doi: 10.1029/2011JC007409, 2012.
- Chang, J., Chiang, K. P., Gong, G. C.: Seasonal variation and cross-shelf distribution of the nitrogen fixing cyanobacterium, Trichodesmium, in southern East China Sea, Continental Shelf
   Research, 20: 479-492, doi: 10.1016/S0278-4343(99)00082-5, 2000.
- Chen, M., Lu, Y., Jiao, N., Tian, J., Kao, S. J., and Zhang, Y.: Biogeographic drivers of diazotrophs
  in the western Pacific Ocean: Pacific Ocean diazotroph biogeography, Limnology and
  Oceanography, 64 (3), 1403-1421, doi: 10.1002/lno.11123, 2019.
- Chen, Y. Y., Sun, X. X., and Zhu, M. L.: Net-phytoplankton communities in the Western Boundary
   Currents and their environmental correlations, Chinese Journal of Oceanology and Limnology,
   doi: 10.1007/s00343-017-6261-8, 2018a.
- Chen, Z., Sun, J., and Zhang, G. C.: Netz-phytoplankton community structure of the tropical
  Western Pacific Ocean in summer 2016, Marine science, 42 (7), 114-130, doi:
  10.11759//hykx20180331002, 2018b.
- Christian, J. R., Murtugudde, R., Ballabrera-Poy, J., and Mcclain, C. R.: A ribbon of dark water:
  phytoplankton blooms in the meanders of the Pacific north equatorial countercurrent, DeepSea Research Part II, 51 (1): 209-228, doi: 10.1016/j.dsr2.2003.06.002, 2004.
- Dai, M. H., Wang, L., Guo, X., Zhai, W., Li, Q., He, B., and Kao, S. J.: Nitrification and inorganic
   nitrogen distribution in a large perturbed river/estuarine system: the Pearl River Estuary, China,
   Biogeosciences, 5, 1545-1585, doi: 10.5194/bg-5-1227-2008, 2008.
- Dai, S., Zhao, Y. F., Li, X. G., Wang, Z. Y., Zhu, M. L., Liang, J. H., Liu, H. J., Tian, Z. Y., and Sun,
   X. X.: The seamount effect on phytoplankton in the tropical western Pacific. Marine
   Environmental Research, 162, doi: 10.1016/j.marenvres.2020.105094, 2020.
- Daufresne, M., Lengfellner, K., and Sommer, U: Global warming benefits the small in aquatic
  ecosystems, Proceedings of the National Academy of Sciences, 106 (31), 12788-12793, doi:
  10.1073/pnas.0902080106, 2009.
- Dong, L., Li, L., Wang, H., He, J., and Wei, Y. L.: Phytoplankton distribution in surface water of
  western Pacific during winter, 2008: a study of molecular organic geochemistry, Marine
  Geology & Quaternary Geology, doi: CNKI:SUN:HYDZ.0.2012-01-012, 2012.
- Dugdale, R. C., Menzel, D. W., Ryther, J. H.: Nitrogen fixation in the Sargasso Sea, Deep Sea
  Research, 7 (4): 297-300 doi: 10.1016/0146-6313(61)90051-X, 1961.
- 44 Edwards, M. and Richardson, A.: Impact of climate change on marine pelagic phenology and trophic

1	mismatch, Nature, 430 (7002), 881-884, doi: 10.1038/nature02808, 2004.
2	Egge, J. K.: Are diatoms poor competitors at low phosphate concentrations?, Journal of Marine
3	Systems, 16 (3-4), 191-198, doi: 10.1016/S0924-7963(97)00113-9, 1998.
4	Emery, W. J. and Dewar, J. S: Mean temperature-salinity, salinity-depth and temperature-depth
5	curves for the North Atlantic and the North Pacific, Progress in Oceanography, 11 (3), 257-256,
6	doi: 10.1016/0079-6611(82)90015-5, 1982.
7	Falkowski, P. G. and Oliver, M. J.: Mix and match: how climate selects phytoplankton, Nature
8	Reviews Microbiology, 5 (10), 813-819, doi: 10.1038/nrmicro1751, 2007.
9	Gao, K., Beardall, J., Hder, D. P., Hall-Spencer, J. M., and Hutchins, D. A.: Effects of ocean
10	acidification on marine photosynthetic organisms under the con-current influences of warming,
11	UV radiation, and deoxygenation, Frontiers in Marine Science, 6, doi:
12	10.3389/fmars.2019.00322, 2019.
13	Raven, J., Capone, D. G., Carpenter, E. J., Hutchins, D. A., Burn, J.: Phosphorus limitation of
14	nitrogen fixation by Trichodesmium in the central Atlantic Ocean, Nature, 411 (6833): 66-69,
15	doi: 10.1038/35075041, 2001.
16	Gordon, A. L., and Fine, R. A.: Pathways of water between the Pacific and Indian oceans in the
17	Indonesian seas, Nature, 379 (6561), 146-149, doi: 10.1038/379146a0, 1996.
18	Grosskopf, T., Mohr, W., Baustian, T., Schunck, H., Gill, D., Kuypers, M., Lavik, G., Schmitz, R.
19	A., Wallace, D., and Laroche, J.: Doubling of marine dinitrogen-fixation rates based on direct
20	measurements, Nature, 488 (7411), 361-364, doi: 10.1038/nature11338, 2012.
21	Gruber, N.: Warming up, turning sour, losing breath: ocean biogeochemistry under global change,
22	Philo Trans A Math Phys Eng, 369 (1943), 1980-1996, doi: 10.1098/rsta.2011.0003, 2011.
23	Guo, S. J., Feng, Y. Y., Lei, W., Dai, M., Liu, Z., Yan, B., and Sun, J.: Seasonal variation in the
24	phytoplankton community of a continental-shelf sea: the East China Sea, Marine Ecology
25	Progress, 103-126, doi: 10.3354/meps10952, 2014.
26	Gupta, A. S., Thomsen, M., Benthuysen J. A., Hobday, A. J., and Dan, A. S.: Drivers and impacts
27	of the most extreme marine heatwaves events, Scientific reports, 10 (1), doi: 10.1038/s41598-
28	020-75445-3, 2020.
29	Hansell, D. A. and Feely, R. A.: Atmospheric Intertropical Convergence impacts surface ocean
30	carbon and nitrogen biogeochemistry in the western tropical Pacific, Geophysical Research
31	Letters, 27 (7), 1013-1016, doi: 10.1029/1999gl002376, 2000.
32	Hidalgo, M., Reglero, P., Álvarez-Berastegui, D., Torres, A. P., Álvarez, I., Rodriguez, J. M.,
33	Carbonell, A., Zaragoza, N., Tor, A., Goñi, R., Mallol, S., Balbín, R., and Alemany, F.:
34	Hydrographic and biological components of the seascape structure the meroplankton
35	community in a frontal system, Marine Ecology Progress Series, 505, 65-80, doi:
36	10.3354/meps10763, 2014.
37	Hillebrand, H., Steinert, G., Boersma, M., Malzahn, A., Meunier, C. L., Plum, C., and Ptacnik, R.:
38	Goldman revisited: Faster-growing phytoplankton has lower N: P and lower stoichiometric
39	flexibility, Limnology and Oceanography, 58 (6), 2076-2088, doi:10.4319/lo.2013.58.6.2076,
40	2013.
41	Hoegh-Guldberg, O. and Bruno, J. F.: The impact of climate change on the world's marine
42	ecosystems, Science, 328 (5985), 1523-1528, doi: 10.1126/science.1189930, 2010.
43	Hu, D. X., Wu, L. X., Cai, W. J., Gupta, A. S., Ganachaud, A., Qiu, B., Gordon, A. L., Lin, X.P.,
44	Chen, Z. H., Hu, S. J., Wang, G. J., Wang, Q. Y., Sprintall, J., Qu, T. D., Kashino, Y. J., Wang,

1 F., and Kessler, W. S.: Pacific western boundary currents and their roles in climate, Nature, 522 2 (7556), 299-308, doi: 10.1038/nature14504, 2015. 3 Hu, S. J. and Hu, D. X.: Heat center of the western Pacific warm pool, Chinese Journal of 4 Oceanology and Limnolog, 30 (1), 169-176, doi: 10.1007/s00343-012-1193-9, 2012. 5 Hu, S. J. and Hu, D. X.: Review on Western Pacific Warm Pool Study, Studia Marina Sinica, 37-48, 6 doi: 10.12036/hykxjk20160724001, 2016. 7 Huang, B. Q., Ou, L. J., Hong, H. S., and Wang, D.: Bioavailability of dissolved organic phosphorus 8 compounds to typical harmful dinoflagellate Prorocentrum donghaiense Lu, Marine Pollution 9 Bulletin, 51: 838-844, doi: 10.1016/j.marpolbul.2005.02.035, 2005. 10 Huisman, J., Sharples, J., Stroom, J. M., Visser, P. M., Kardinaal, W., Verspagen, J., and Sommeijer, 11 B.: Changes in turbulent mixing shift competition for light between phytoplankton species, 12 Ecology, 85 (11), 2960-2970, doi: 10.1890/03-0763, 2004. 13 Isamu, Y.: Illustrations of the Marine Plankton of Japan, Tpkyo: Hoikusha Press, 1-158, 1991. 14 Jin, D. X., and Chen, J. H.: Chinese Marine Planktonic Diatoms, Shanghai Scientific & Technical 15 Press, 1-230, 1965. 16 Karl, D., Michaels, A., Bergman, B., Capone, D., Carpenter, E., Letelier, R., Lipschultz, F., Paerl, 17 H.: Dinitrogen fixation in the world's oceans, Biogeochemistry, 57-58 (1): 47-98 doi: 18 10.1007/978-94-017-3405-9 2, 2002. 19 Karl, D. M., and Tien, G.: Temporal variability in dissolved phosphorus concentrations in the 20 subtropical North Pacific Ocean. Marine Chemistry, 56: 77-96, doi: 10.1016/S0304-21 4203(96)00081-3, 1997. 22 Kawahata, H., Nishimura, A., and Gagan, M. K., Seasonal change in foraminiferal production in the 23 western equatorial Pacific warm pool: evidence from sediment trap experiments, Deep Sea 24 Research Part II: Topical Studies in Oceanography, 49 (13-14): 2783-2800, doi: 25 10.1016/S0967-0645(02)00058-9, 2002. 26 Kustka, A., Carpenter, E. J., Sanudo-Wilhelmy, S. A.: Iron and marine nitrogen fixation:progress 27 and future directions, Research in Microbiology, 153: 255-262, doi: 10.1016/S0923-2508(02)01325-6, 2002. 28 29 Laroche, J., and Breitbarth, E.: Importance of the diazotrophs as a source of new nitrogen in the 30 ocean, Journal of Sea Research, 53 (1): 67-91, doi: 10.1016/j.seares.2004.05.005, 2005. 31 Li, Q., Legendre, L., Jiao, N. Z.: Phytoplankton responses to nitrogen and iron limitation in the 32 tropical and subtropical Pacific Ocean, Journal of Plankton Research, 37 (2): 306-319, doi: 33 10.1093/plankt/fbv008, 2015. 34 Liu, D. Y., Sun, J., and Qian, S. B.: Planktonic dinoflagellate in Ryukyu Islands and its adjacent 35 waters-species composition and abundance distribution in summer 1997, 2000. 36 Lin, G., Chen, Y., Huang, J., Wang, Y., and Yang, Q.: Regional disparities of phytoplankton in 37 relation to different water masses in the Northwest Pacific Ocean during the spring and summer 38 of 2017. Acta Oceanologica Sinica, 39 (5), doi: 10.1007/s13131-019-1511-6, 2020. 39 Long, Y., Noman, A., Chen, D. W., Wang, S. H., Yu, H., Chen, H. T., Wang, M., and Sun, J.: Western 40 pacific zooplankton community along latitudinal and equatorial transects in autumn 2017 (northern hemisphere), Diversity, 13 (2), 58, doi: 10.3390/d13020058, 2021. 41 42 Lozier, M. S., Dave, A. C., Palter, J. B., Gerber, L. M., and Barber, R. T.: On the relationship between 43 stratification and primary productivity in the North Atlantic, Geophysical Research Letters, 38, 44 doi: 10.1029/2011GL049414, 2011.

- Luo, Y. W., Doney, S. C., Anderson, L. A., Benavides, M., and Mara<sup>n</sup>on, E.: Database of
   diazotrophs in global ocean: abundance, biomass and nitrogen fixation rates, Earth System
   Science Data, 4 (1), 47-73, doi: 10.5194/essd-4-47-2012, 2012.
- Messié, M. and Radenac, M. H.: Seasonal variability of the surface chlorophyll in the western
  tropical Pacific from SeaWiFS data, Deep Sea Research Part I: Oceanographic Research Papers,
  53 (10), 1581-1600, doi: 10.1016/j.dsr.2006.06.007, 2006.
- Mojica, K., van de Poll, W., Kehoe, M., Huisman, J., Timmermans, K., Buma, A., Woerd, H., HahnWoernle, L., and Dijkstra, H.: Phytoplankton community structure in relation to vertical
  stratification along a north-south gradient in the Northeast Atlantic Ocean, Limnology and
  Oceanography, 60, doi: 10.1002/lno.10113, 2015.
- Oksanen, J. F., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P. R.,
   O'Hara, R. B., Simpson, G. L., Solymos, P., Stevens, M. H., Szoecs, E., and Wagner, H.: vegan:
   Community Ecology Package, 2020.
- Pai, S.C., Tsau, Y.J., and Yang, T.I.: PH and buffering capacity problems involved in the
  determination of ammonia in saline water using the indophenol blue spectrophotometric
  method, Analytica Chimica Acta, 434 (2), 209-216, doi: 10.1016/S0003-2670(01)00851-0,
  2001.
- Perez, V., Fernandez, E., Maranon, E., Moran, X., and Zubkov, M.: Vertical distribution of
  phytoplankton biomass, production and growth in the Atlantic subtropical gyres, Deep Sea
  Research Part I: Oceanographic Research Papers, 53 (10), 1616-1634, doi:
  10.1016/j.dsr.2006.07.008, 2006.
- Qian, S. B., Liu, D. Y., and Sun, J.: Marine phycology, Qingdao: China Ocean University Press, 1 5, 2005.
- Richardson, A. J., and Schoeman, D. S.: Climate impact on plankton ecosystems in the Northeast
   Atlantic, Science, 305 (5690), 1609-1612, doi: 10.1126/science.1100958, 2004.
- Richardson, K., and Bendtsen, J.: Vertical distribution of phytoplankton and primary production in
   relation to nutricline depth in the open ocean, Marine Ecology Progress Series, 620, 33-46, doi:
   10.3354/meps12960, 2019.
- Siokou-Frangou, I., Christaki, U., Mazzocchi, M. G., Montresor, M., Ribera, D. M., Vaqué, D., and
  Zingone, A.: Plankton in the open Mediterranean Sea: a review, Biogeosciences, 7 (5), 15431586, doi: 10.5194/bg-7-1543-2010, 2010.
- Smayda T J.: Harmful algal blooms: their ecophysiology and general relevance to phytoplankton
   blooms in the sea. Limnology and Oceanography, 42: 1137-1153, doi: 10.2307/2839007, 1997.
- Sohm, J. A., Webb, E. A., and Capone, D. G.: Emerging patterns of marine nitrogen fixation, Nature
   Reviews Microbiology, 9 (7), 499-508, doi: 10.1038/nrmicro2594, 2011.
- Stoecker, D. K.: Mixotrophy among dinoflagellates, Journal Eukaryot Microbiol, 46: 397-401, doi:
   10.1111/j.1550-7408.1999.tb04619.x, 1999.
- Sun, J.: Marine phytoplankton and biological carbon sink, Acta Ecologica Sinica, 31 (18), 53725378, 2011.
- Sun, J., and Liu, D. Y.: The Preliminary Notion on Nomenclature of Common Phytoplankton in
  China Sea Waters, Oceanologia Et Limnologia Sinica, 33 (3): 271-286, doi: 10.1088/10091963/11/5/313, 2002.
- Sun, J., Liu, D. Y., and Qian, S. B.: Planktonic diatoms in Ryukyu Islands and its adjacent waters species composition and abundance distribution in summer 1997, 2000.

- Tang, S. M., Cai, R. S., Guo, H. X., and Wang, L.: Response of phytoplankton ecology to climate
   change in offshore China, Applied Journal of Oceanography, 36 (4), 455-465, doi:
   10.3969/J.ISSN.2095-4972.2017.04.001, 2017.
- Tilman, D., Kiesling, R., Sterner, R., Kilham, S. S., and Johnson, F. A.: Green, bluegreen and diatom
  algae: taxonomic differences in competitive ability for phosphorus, silicon and nitrogen, Arch
  Hydrobiol, 106 (4): 473-485, doi: 10.1029/WR022i007p01162, 1986.
- Wang, F., Hu, D. X., Mu, M., Wang, Q., He, J. H., Zhu, J., and Liu, Z. Y.: Structure, Variations and
  Climatic Impacts of Ocean Circulation and the Warm Pool in the Tropical Pacific Ocean,
  Advances in Earth Sciences, 27 (6): 595-602, doi: CNKI:SUN:DXJZ.0.2012-06-000, 2012.
- Yan, X. H., Ho, C. R., Zheng, Q., and Klemas, V.: Temperature and size variabilities of the Western
   pacific warm pool, Science, 258: 1643-1645, doi: 10.1126/science.258.5088.1643, 1992.
- Yentsch, C. S.: Distribution of chlorophyll and phaeophytin in the open ocean, Deep-Sea Research,
  12, 653-666, doi: 10.1016/0011-7471(65)91864-4, 1965.
- Zehr, J. P.: Nitrogen fixation by marine cyanobacteria, Trends in Microbiology, 19 (4), 162-173, doi:
   10.1016/j.tim.2010.12.004, 2011.
- Zhang, Q.: Relationship between the precipitation in the rainy season in north China and the tropical
   western pacific warm pool and Kuroshio, Plateau Meteorology, 18 (4), 575-583, 1999.
- Zhao, X.: The characteristic of winter and spring atmospheric heat source over the East China Sea
   Kuroshio and western North Pacific subtropics and its impact on precipitation in Eastern China,
   NanJing University of Information Science & Technology, 2015.
- Zhu, J., Zheng, Q.N., Hu, J.Y., Lin, H.Y., Chen, D.W., Chen, Z.Z., Sun, Z.Y., Li, L.Y., Kong, H.:
  Classification and 3-D distribution of upper layer water masses in the northern South China
  Sea, Acta Oceanol. Sin., 38, 126–135, doi:10.1007/s13131-019-1418-2, 2019.