REPLIES TO REVIEWERS

We would like to thank the reviewers for their constructive comments that helped improve our manuscript. Further we give our response to the comments of both reviewers point by point:

RC: Reviewer Comments, AC: Authors Comments and reply

R1 refers to Reviewer 1 and R2 to Reviewer 2

Replies to Reviewer 1

RC General Comments:

This manuscript presents a DEB-bioaccumulation model for microplastics. The model was calibrated and corroborated with field data available in the North Sea and Northern Ionian Sea, showing some skill in reproducing the (few) available observations. The topic is of interest to the readership of this Journal. The manuscript is very well written and clear. The model, the simulations and the analyses are robust and discussed thoroughly. I have a number of comments that I reported in the pdf version of the manuscript that I am attaching to this review. Here I will mention just two moderate concerns of mine regarding this work. 1) The authors used an ocean-colour chlorophyll product as input of the DEB model. However, this product might be biased in optically complex coastal waters, such as the Southern North Sea considered in this manuscript. The issue is relevant, because the authors pointed out the impact of the high chlorophyll concentration on the results they obtained in the North Sea. I recommend that the author discuss the reliability of the chlorophyll product they used. For example, they could compare the ocean colour product with in situ chlorophyll data from the ICES database (https://www.ices.dk/marine-data/data-portals/Pages/default.aspx), or with the NSBC climatology (https://icdc.cen.uni-hamburg.de/1/daten/ocean/knsc-hydrographic0/) 2) The authors should point out and discuss a bit more extensively some flaws in the results of their simulations and analysis (e.g. the overestimation of the observed MCs in Figure 6, and the mismatch between the regression results and the data at two sites in Figure 13). I appreciated that these flaws were clearly mentioned in the conclusions. I don’t think that these issues compromised the value of the work. More minor to moderate issues are mentioned in the attached pdf of the manuscript.

AC Reply

We would like to thank R1 for carefully reading the manuscript and the very useful comments. With regard to the two moderate concerns:

1) The satellite dataset that we used presented a much better spatial and temporal coverage in our study area, as compared to other available datasets, such as the regional CHL-a satellite product, recommended by the reviewer (CMEMS, OCEANCOLOUR_ATL_CHL_L3_REP_OBSERVATIONS_009_067). As demonstrated in several previous studies (i.e. Sara et al., 2011, 2012, Monaco & McQuaid, 2018) it is crucial to
include daily CHL-a data to force the DEB model, in order to properly simulate the daily fluctuations of the environmental forcing data and thus the MPs accumulation.

In order to assess the reliability of the chlorophyll product that we used, we have followed the reviewer’s recommendation and compared the ocean color product as an input of the DEB model, with in situ data available from the ICES database (https://www.ices.dk/marine-data/data-portals/Pages/default.aspx). Specifically, we used the in situ chlorophyll-a (CHL-a) data from the ICES database, that is derived from the surface layer (0-3m depth), and covers our study period of 2007-2011. A mean value, regarding depth, was computed, as representative of the upper surface layer. The in situ CHL-a data, which are included within our study area (Sec. 2.4, Southern North Sea, 51.08°-51.44° N, 2.19°-3.45° E) were averaged spatially in order to fit the “box model” of our study. Apparently some days were missing during this time period (2007-2011), while some days there were more than one measurement and then mean daily values were computed. The results from the comparison are presented also in Fig. 1 of the revised manuscript, along with the used satellite data (CMEMS-Globcolour, OCEANCOLOUR_ATL_CHL_L4_REP_OBSERVATIONS_009_098). The used satellite data are in quite good agreement with the available in situ data, presenting relatively small error (RMSE and correlation coefficient computed). The result of this comparison is shown also at the attached Fig.1 here.

The same comparison was conducted also with the regional CHL-a satellite product, suggested by the reviewer (CMEMS, OCEANCOLOUR_ATL_CHL_L3_REP_OBSERVATIONS_009_067), against the in situ data described above. We attached the Fig. 2 (only here) that shows the result of this comparison. The statistical analysis resulted in relatively higher RMSE and lower correlation coefficient, as compared to the one computed with the other (Globcolour) dataset.

Therefore, given also the better spatial and temporal coverage, we concluded that the satellite dataset that we used is optimal in our study area and time period.

2) Changes were made according to the reviewer’s comment at the corresponding parts throughout the manuscript. Specifically, we have pointed out and discussed more extensively some flaws in the results of our simulations and analysis, namely the overestimation of the observed MPs in Figure 6, and the mismatch between the regression results and the data at two sites in Figure 13 of the manuscript. These points are listed in detail below, based on the corresponding comments of R1 in the manuscript.

All the line numbers written in RC are the line numbers of the R1’s attached pdf manuscript, while the line numbers in AC correspond to the revised manuscript.

RC: line 15 replace “by” with “in”
AC: line 16 done.
RC: line 19 add “mussel”
AC: line 20 done.
RC: line 22 How long was the time window?
AC: line 24-25 the information was added “after 4 years and 1 year simulation”.
RC: line 35 "those": is it referred to MPs or sources?
AC: line 37 it is referred to MPs and was replaced with “those particles”.
RC: line 56 replace “from” with “at”
AC: line 58 done.
RC: line 194-195 please clarify what you mean with "represented", in this context
AC: line 196-197 the clarification was done by adding additional information in this sentence “assuming that all parameters referred to silt (or inedible particles) are applicable also to MPs particles”
RC: line 240 this implies that the last term in eq. 18 has the dimension of "particles" (C), rather than C/time. Please check the units
AC: line 242-252 The Reviewer is right. We reconstructed the Eq. 18 adding a parameter $k_f (d^{-1})$ in the third term that represents the post-ingestive selection mechanism utilized by the mussel to incorporate indigestible material (i.e. MPs) into faeces. This parameter is calibrated at a constant value for both study areas and illustrates the mussel’s mechanism to discriminate between particles in the gut based on physicochemical criteria according to Ward et al. (2019) review study. This parameter also compensates the units of the third term of Eq. 18 to C/time.
The whole paragraph was reconstructed to adjust the new information (line 242-254) of the revised manuscript.
RC: line 244 This should be defined just before eq. 18, to help the reader understanding the equation
AC: line 238-240 done.
RC: line 250 chlorophyll-a concentration includes large to pico functional groups of phytoplankton. Are all the groups suitable to feed mussels? If not, please clarify the limitation of this assumption in the context of your work.
AC: line 264-268 The information was added about the size limitations of the suspended matter that a mussel is able to filter.
RC: line 251 Has this result of Hatzonikolakis have general validity, or was it referred to a particular location/experiment/condition? Please discuss the extent to which this result can be reliable in the context of your application
AC: line 269-276 Although the result of Hatzonikolakis et al. (2017) was referred to a particular location/condition, the same result has been demonstrated also in other studies (i.e. Troost et al., 2010) referred to different locations and conditions. Considering that POC contributes to the mussel diet when the CHL-a concentration is low enough (Troost et al., 2010) and that our study areas
(North Sea is a more eutrophic environment and N. Ionian Sea refers to mussel farm and thus suitable conditions regarding CHL-a) do not present low Chl-a values, CHL-a can be assumed to be the dominant food source of mussels. The relevant information was added in these lines of the revised manuscript.

RC: line 258-260 Why did you not use the regional chlorophyll product available for the North West Shelf-Seas in the CMEMS catalogue? I think the regional product would have been preferable, because it takes account of the complexity of the optical waters in the coastal North Sea.

AC: line 287-296 This question has been answered in our first comment under the general R1’s comment. In this part we added the result of the comparison between the used satellite dataset and the available in situ data from the ICES database. The in situ data were also added in Fig. 1 of the revised manuscript. For more details about the comparison between the recommended regional chlorophyll product and the in situ data, the reader may see the first AC reply.

RC: line 266 Similar to my previous question: why did you not use the CMEMS product for the Mediterranean Sea?

AC: line 305-311 The chosen CHL-a dataset was found preferable, as compared with other available remote sensing datasets (i.e. CMEMS chlorophyll product for Mediterranean Sea), since it presented a better spatial and temporal coverage (Hourany et al., 2019, Garnesson et al., 2019). Unfortunately, available insitu data are very scarce in the study area and therefore an extended comparison between remote and in situ data could not be conducted. However, we have compared the used satellite data (Globcolour) and the proposed product (CMEMS for Mediterranean Sea) with very few available (unpublished) in situ data, which were obtained in the framework of WFD (Water Framework Directive data kindly provided by Georgia Assimakopoulou) (Fig. 3 attached here). The few available CHL-a data in our study area (N 39.49°-39.65°, E 20.09°-20.23°) - specifically “Station Igoumenitsa” (N 39.5°, E 20.2281°) and “Station Kalamas” (N 39.6°, E 20.1439°)- were sampled at 2 m depth, covering the time period of our study (November 2014 to December 2015). The sampling dates were the same in both stations (8-Dec-14, 8-Mar-15, 12-Dec-15), allowing us to compute the mean CHL-a dataset (final 3 values) representative of our “box model”. The comparison between the used satellite dataset, the proposed (regional) satellite product and the few in situ data showed the better temporal coverage and a slightly lower error for our satellite dataset (Globcolour) than the regional CMEMS product (Fig. 3 here).

RC line 272 Worth mentioning that PFT satellite products for your study regions were provided by Di Cicco et al., 2017 and Brewin et al 2017 and are now an operational product of CMEMS

AC line 315 the references were added.

RC line 281 Are you sure? Looking at your data I would say the peak is in Spring, isn't it? See also, e.g., Widdicombe et al. (2010), Journal of Plankton Research (although that paper refers to the English Channel)

AC line 322-325 the whole sentence has been clarified. We referred to the rivers discharge peaking at winter period (Van Beusekom et al., 2009), and not to the CHL-a concentration and/or productivity, which indeed peaks at spring season in the North Sea study area (shown also at Fig. 1 of the manuscript).
RC line 407 can you assume that the behavior of the model is close to linear within this range of parameter variation? In fact, SI is meaningful if the linear approximation is adequate.

AC line 450-451 The R1’s concern is justifiable, since we are aware of the general nonlinear effect of some parameters (i.e. temperature) on the DEB model; however within this range of parameter variation, we assumed that the model approximates the linear behavior. Our intention was to examine if the perturbed variables/parameters have an effect (or not) on the simulated MPs accumulation, in order to proceed with the development of the regression model, relating directly the environmental MPs concentration with the variables/parameters that had high effect on model’s result (CHL-a, temperature and the mussel’s weight and MPs load) (Eq. 20 of the manuscript). Nonetheless, the same method (sensitivity index, SI) has been also applied in other studies, which intended to examine the model’s sensitivity on specific variables/parameters regarding the mussel growth (Casas and Bacher, 2006, Rosland et al., 2009, Béjaoui-Omri et al., 2014, Hatzonikolakis et al., 2017).

RC line 430 the format could be better

AC line 477 the format was changed.

RC line 430 Please define C. Does it represent OBSERVATIONS of C in the environment?

AC line 480 done. C represents the simulated MPs accumulation in the mussel.

RC line 446 Seems to me that Fig 4 is referred to before Figure 3. Please revise the order of the figures

AC line 493 done.

RC line 447 can you provide an estimate of the error, please?

AC line 494 A standard deviation was estimated by tuning the model with various \( X_k \) values and comparing the model’s simulation with the available field data. The model’s result with the estimated value range (\( X_k = 8 \pm 1.5 \text{ mg m}^{-3} \)) was in agreement with the field data and within their standard deviation.

RC line 453 Was the French site a clean one?

AC line 499-505 The sentence was revised to better justify the high value of \( X_k \), regarding the food quality of the mussel’s diet. Apart from the fact that the French site presented lower CHL-a concentrations compared to our study area (North Sea), the DEB model applied at the French site, did not include inedible particles in the mussel’s food (so it was assumed clean of inedible particles). On the other hand, in our study the inedible particles (i.e. MPs) have been incorporated in the mussel’s diet through the modified relation of the functional response \( f \) (Eq. 5, Table 1 in the manuscript), which regulates the assimilation rate and thus the mussel’s growth (see also lines 202-208 in the manuscript). Consequently, the higher value of \( X_k \) in our study area, reflects the lower quality food and affects the half-saturation constant (\( X_k \)) according to Kooijman (2006).

RC line 464 This is not coherent with your comment of Figure 1 that CHL peaked in winter (which in fact I think was wrong)
AC See also the corresponding comment above. The comment was referred to the rivers discharge and not to the CHL-a peak. It was rephrased as mentioned, in lines 322-325 of the revised manuscript.

RC line 480 this reference comes after figure 4: please revise

AC line 532 done.

RC line 492 Please mention that the model overestimated the data range explicitly. It seems like the model reproduced a seasonal increase that was not observed.

AC line 547-548 R1’s comment was considered and discussed here and also throughout the whole manuscript at the corresponding parts, as recommended.

RC line 495 Is this explanation quantifiable?

AC line 551-554 This explanation was quantified, discussed and compared with our model’s result.

RC line 498-500 This sounds quite a speculation that goes too far. at the end of the day, you calibrated the model to fit the equations to the data.

AC 557-562 the whole sentence was revised, including the model’s overestimation in the North Sea simulation.

RC line 500 not really in agreement in the North Sea

AC 557-562 This was commented.

RC line 554 Please mention here in the text and discuss later in the manuscript that in the North Sea the range of variability of the data and model uncertainty do not really overlap significantly at the time of the observations.

AC line 613-615 R1’s comment was mentioned here and discussed in the corresponding parts of the manuscript (see also the specific parts discussed in the following comments).

RC line 594 please describe briefly the results shown in figure 13 (e.g. general overlapping of regressed and observed C, except in Hastings and Plymouth)

AC line 664-675 We described the results of Fig. 13 and discussed thoroughly about the possible explanation of the two exceptions (Hastings and Plymouth).

RC line 614-615 Please rephrase.

AC line 689-691 We rephrased the whole sentence.

RC line 623 Which approach? The seasonally variable approach? Please clarify this sentence

AC line 696-704 this was referred to our applied approach with the daily CHL-a fluctuations. In fact, it was further clarified by adding a sentence describing the evolution step that we made, by applying a daily variable approach.

RC line 627 Please remind the reader which figures presented these results.
AC line 706 done.

RC line 630 Please mention and discuss the model overestimation of MP in Figure 6.

AC line 706-712 We would like to thank R1 for the comment, as it allowed us to interpret better our results and compare further the observed field data with the model’s result, regarding not only the MPs accumulation in the mussel but also the MPs elimination after 24 hours of depuration. The comment was mentioned and discussed thoroughly.

RC line 641-643 Not clear to me why this sentence is relevant here. Please clarify.

AC line 726-729 R1 is right. We also found this sentence irrelevant here, so it was deleted. The information of this sentence has been communicated to the reader earlier in the relevant part (Sec. 3.3 of the manuscript).

RC line 646-647 This sounds an over-stretched statement. please provide more evidence

AC line 731-734 We rephrased it and provided more evidence, as recommended.

RC line 662 Am I wrong, or Cenv is the dependent variable and C the predictor, in eq. 20?

AC line 751 R1 is right. This was a misprint and was corrected.

RC line 663 But in Hastings and Plymouth, why?

AC line 757-759 This was clearly stated and an explanation was suggested. The specific issue was discussed more in lines 664-675, as mentioned in a previous comment reply.

RC line 670-674 quite long sentence, please consider to split it in 2.

AC line 766-771 done.

RC line 677 Please specify what data you are talking about. MP in mussels, right?

AC line 773 This was done. Yes, we were referred to MPs in mussels.

RC line 686 overstatement. In figure 6 the data are clearly overestimated

AC line 782-784 The sentence was rephrased, including the overestimation statement.

RC line 687 replace “to be close to reality” with “to represent the natural variability”

AC line 785 done.

RC line 693 Here I do not understand: Cenv is highly variable because it is naturally variable, or because there are huge observational errors? please clarify

AC line 790-795 We clarified this by adding a sentence. We thank R1 for the comment.

RC line 729 Is this info relevant?

AC line 833 The information was deleted as we also found it irrelevant.

RC line 745 approximated? considered indirectly?
AC line 851 We replaced it with “considered indirectly”.

RC line 747-752 the logical flow of these last sentences was rather unclear to me. Please consider rephrasing this part.

AC line 851-859 We rephrased these sentences.

RC line 756-757 This is a strong critic, which I appreciate and don't consider a crucial flaw. However, such critic was not as much evident in the presentation and discussion of the results (e.g. Figure 6?). Please report the model flaw also in the presentation and discussion of the results.

AC line 864-865 This sentence was rephrased. The model flaws were reported and discussed extensively at the corresponding parts throughout the whole manuscript (i.e. presentation and discussion of the results, see also above comments reply).

RC line 772-774 I found that the sentence included redundant information, thus I suggested some changes. Please feel free to reject my suggestions

AC line 881-883 We rephrased the sentence, adjusting some changes.

RC page 40 (Figures) Can't you plot just the bottom x-axis, in black? (the upper one is redundant)

AC page 43 Fig. 8 done.

Replies to Reviewer 2

RC General Comments:

In the current manuscript, a Dynamic Energy Budget model is developed aiming to simulate the uptake and excretion rate of microplastics, by two species of mussels at two different regions (North Sea and N. Ionian Sea). The authors claim that the biophysical regime (in this case chlorophyll and sea surface temperature) influences the accumulation rates in filter feeders.

Overall, I think that the paper is well-written, without any major issues or inaccuracies. I appreciate the clear figures that allow following the manuscript. The literature is well cited and extensive. I truly enjoyed reading it; the authors have put substantial efforts in preparing their manuscript. I am, thus, recommending a few minor comments/ suggestions for their consideration. My only semi-major comment, which does not impact the overall research output, is about the overestimation of the satellite derived chlorophyll concentrations at the southern North Sea. Please see the specific comment in the next section.

AC Reply

We would like to thank R2 for his time and considerations and for carefully reading the manuscript and providing useful comments. With regard to the semi-major comment:
Since this was one of the two moderate concerns of R1, we have already examined and discussed the reliability of the satellite derived chlorophyll concentrations that were used at the southern North Sea. We compared the used satellite data (CMEMS-Globcolour), as well as, the regional satellite product (CMEMS product for the North West Shelf-Seas) with the available in situ data (ICES database) (see Fig. 1 and Fig.2 attached here). We concluded that the satellite data used, have a better spatial and temporal coverage and relatively lower error as compared to the regional CHL-a product. For more details, the reader may also see the first reply in R1’s comment.

All the line numbers written in RC are the line numbers referred to the original manuscript (written in R2’s review), while the line numbers in AC correspond to the revised manuscript.

Abstract

RC line 7: MavroLithari should read Mavrolithari or Mavro Lithari.

AC line 7: it should read Mavro Lithari, so we corrected it.

RC line 29-30: “… with MPs accumulation in mussel’s soft tissue, temperature and chlorophyll-a.”. The sentence does not flow well. Pease revise.

AC line 31-32 R2 is right, so we rephrased the sentence.

Introduction

RC: It flows very well, with informative and well-referenced text. The novelty of the current study is clear.

Line 95 (and 97): DEB has been abbreviated in the abstract, please check the journal rules (if abbreviated in the abstract, is there a need to be abbreviated in the text too?). Once you find out the rule, please apply to the species M. edulis and M. galloprovincialis (lines 96/97).

AC We would like to thank R2.

According to the journal rules, abbreviations need to be defined in the abstract and then again at the first instance in the rest of the text. We followed this rule and abbreviated the DEB in the abstract and then at the first instance (line 97). The same was applied also for the species (abstract and line 99-100).

Materials and Methods

RC line 112: The North Sea is a marginal sea rather a semi-enclosed environment (clear openings and influence from the Atlantic from both sides).

AC line 116 The suggestion was done. “semi-enclosed” was replaced with “marginal”.

RC line 180: DEB is abbreviated again – please revise.
AC line 184 R2 is right, this was corrected.

RC line 250: chlorophyll-a concentrations (CHL-a, an index of phytoplankton biomass).

AC line 266: done.

RC line 258: in the future, please consider using the OC-CCI Chl-a product, which is a better product for coastal regions. Also available in Copernicus.

AC line 289-298 We have already discussed the reliability of the satellite product that we used in our study area for the specific time period above (in our first reply to R1 and R2 comments) and concluded that the used satellite CHL-a data were suitable for the specific place and time period. However, we thank R2 for the kind recommendation and we will consider this in a future relevant study.

RC line 265: it should read (i.e. SeaWiFS, MERIS, MODIS, VIIRS and OLCI-a). All these are abbreviations and not just names.

AC line 301 R2 is right and we corrected it.

RC: Please provide a simple reasoning why two different products of Chl-a were used for the two different study areas.

AC: This question has been answered also earlier in R1’s comment (“why did you not use the CMEMS product for the Mediterranean Sea?”). We provide the same answer here for the reader’s convenience:

“AC: line 305-311 The answer was included to the added part of the revised manuscript. Moreover, to justify our reply to R1’s question we compared the used satellite data (Globcolour) and the proposed product (CMEMS for Mediterranean Sea) with very few available in situ data, which were obtained in the framework of WFD (Water Framework Directive data kindly provided by Georgia Assimakopoulou). Unfortunately, there are not enough available in situ data for the specific area (Fig. 3 attached here). The available few CHL-a data are included in our study area (N 39.49°-39.65°, E 20.09°-20.23°) - specifically “Station Igoumenitsa” (N 39.5°, E 20.2281°) and “Station Kalamas” (N 39.6°, E 20.1439°) - and sampled at 2 m depth, covering the time period of our study (November 2014 to December 2015). The sampling dates were the same in both stations (8-Dec-14, 8-Mar-15, 12-Dec-15), allowing us to compute the mean CHL-a dataset (final 3 values) representative of our “box model”. The comparison between our used satellite data, the proposed satellite product and the few in situ data showed a better temporal coverage and a slightly lower error for our satellite dataset (Globcolour) than the proposed CMEMS product (Fig. 4 here)”

RC line 275: _0.88 mg chl-a m^-3 = should read _0.88 mg m^-3. Please change throughout.

AC This was changed throughout the manuscript.

RC line 282: sea surface temperature has been already abbreviated, please use the abbreviation (i.e. SST)

AC line 327 done.
RC line 325: “...Pouvreau et al., 2006). In order to...” insert a space after the punctuation mark (full stop).

AC line 370 done.

RC line 278 and Figure 1: it is a bit worrying to observe such high chlorophyll values in the coastal North Sea region. In reality, this environment is eutrophic (no doubt) and certainly very high concentrations are expected. However, your North Sea region belongs to CASE II waters, where algorithms tend to overestimate chlorophyll concentrations. In optically-complex Case II waters, Chl-a can not readily be distinguished from particulate matter and/or yellow substances (dissolved organic matter) and so global chlorophyll algorithms are less reliable. This has to be communicated to the readers.

Please add a few sentences to acknowledge the issue. To facilitate your revision, you will find this reference very useful:


AC line 289-298 We justified the satellite product that we used in previous comments and added a few sentences to acknowledge the issue as requested. Also the above suggested reference was added.

RESULTS

RC line 448: please use the same units (they are the same after all mg m-3 = g l-1)

AC line 497 this was corrected.

END.

**Figures in Replies to Reviewers document**
Fig. 1. Comparison between the satellite data (CMEMS-Globcolour) with the available in situ data (ICES database).

Fig. 2. Comparison between the satellite data (regional CMEMS) with the available in situ data (ICES database).
Fig. 3. Comparison between the Globcolour satellite data, the CMEMS satellite data for the Mediterranean Sea and the (few) available in situ data.
Modelling mussel (*Mytilus spp.*) microplastic accumulation.

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Abstract: Microplastics (MPs) are a contaminant of growing concern due to their widespread distribution and interactions with marine species, such as filter feeders. To investigate the MPs accumulation in wild and cultured mussels, a Dynamic Energy Budget (DEB) model was developed and validated with the available field data of *Mytilus edulis* (*M. edulis*, wild) from the North Sea and *Mytilus galloprovincialis* (*M. galloprovincialis*, cultured) from the Northern Ionian Sea. Towards a generic DEB model, the site-specific model parameter, half saturation coefficient (*Xk*) was applied as a power function of food density for the cultured mussel, while for the wild mussel it was calibrated to a constant value. The DEB-accumulation model simulated the uptake and excretion rate of MPs, taking into account of environmental characteristics (temperature and chlorophyll-a). An accumulation of MPs equal to 0.53⁶⁴ particles individual⁻¹ (fresh tissue mass 1.9 g) and 0.91 particles individual⁻¹ (fresh tissue mass 3.3⁴ g) was simulated for the wild and cultured mussel after 4 years and 1 year respectively, in agreement with the field data. The inverse experiments investigating the depuration time of the wild and cultured mussel in a clean from MPs environment showed a 90% removal of MPs load after 2.5³ and 1²⁴ days, respectively. Furthermore, sensitivity tests on model parameters and forcing functions highlighted that besides MPs concentration, the accumulation is highly depended on temperature and chlorophyll-a of the surrounding environment. For this reason, an empirical equation was found, directly relating directly the environmental concentration of MPs in seawater, with the seawater’s temperature, chlorophyll-a and the MPs accumulation in mussel’s soft tissue MPs load, temperature and chlorophyll-a.
1. Introduction

Microplastic particles (MPs) are synthetic organic polymers with size below 5 mm (Arthur et al., 2009) that originate from a variety of sources including—mainly—those particles that are manufactured for particular household or industrial activities, such as facial scrubs, toothpastes and resin pellets used in the plastic industry (primary MPs), and those formed from the fragmentation of larger plastic items (secondary MPs) (GESAMP, 2015). Eriksen et al. (2014) estimated that more than 5 trillion microplastic particles, weighing over 250,000 tons, float in the oceans. Due to their composition, density and shape, MPs are highly persistent in the environment and are, therefore, accumulating in different marine compartments at increasing rates: surface and deeper layers in the water column, as well as at the seafloor and within the sediments (Moore et al., 2001, Lattin et al., 2004, Thompson, 2004, Lusher, 2015). Since the majority of MPs entering the marine environment, originate from the land (i.e. land-fills, littering of beaches and coastal areas, rivers, floodwaters, untreated municipal sewerage, industrial emissions), the threat of MPs pollution in the coastal zone puts considerable pressure on the coastal ecosystems (Cole et al., 2011, Andrady, 2011). In recent years, initiatives under various projects (i.e. CLAIM, DeFishGear) target at evaluating the threat and impact of marine litter pollution; the European framework of JERICO-RI focuses on a sustainable research infrastructure in the coastal area to support the monitoring, science and management of coastal marine areas (http://www.jerico-ri.eu/). In the framework of JERICO-NEXT, a recent study addressed the environmental threats and gaps within monitoring programmes in European coastal waters, including the marine litter (i.e. MPs) as one of the most commonly identified threat to the marine environment and highlighted the need for improved monitoring of the MPs distribution and their impacts in European coastal environments (Painting et al., 2019).

Numerous studies have revealed that MPs are ingested either directly or through lower trophic prey by animals at from all levels of the food web; from zooplankton (Cole et al., 2013), small pelagic fishes and mussels (Digka et al., 2018a) to mesopelagic fishes (Wieczorek et al., 2018) and large predators like tuna and swordfish (Romeo et al., 2015). Microplastic ingestion by marine animals can potentially affect animal health and raises toxicity concerns, since plastics can facilitate the transfer of chemical additives and/or hydrophobic organic contaminants to biota (Mato et al., 2001, Rios et al., 2007, Teuten et al., 2007, 2009, Hirai et al., 2011). Human, as a top predator, is also contaminated by MPs (Schwabl et al., 2019). Mussel and small fishes that are commonly consumed whole, without removing digestive tracts, where MPs are concentrated, are among the most likely pathways for MPs to embed in the human diet (Smith et al., 2018). Especially regarding marine organisms (i.e. mussels), it is notable that the levels of their contamination has been added
to the European database (www.ecsafeseafooddbase.eu) as an environmental variable of growing concern, reflecting the health status (Marine Strategy Framework Directive (MSFD) Descriptor 10 – Marine Litter (Decision 2017/848/EU)) (De Witte et al., 2014, Vandermeersch et al., 2015, Digka et al., 2018a). Today, a series of studies have denoted the presence of MPs in mussels’ tissue intended for human consumption (Van Cauwenberghe and Janssen, 2014, Mathalon and Hill, 2014, Li et al., 2016, 2018, Hantoro et al., 2019). For instance, in a recent study, Li et al. (2018) sampled mussels from coastal waters and supermarkets in the U.K and estimated that a plate of 100g mussels contains 70 MPs that will be ingested by the consumer. The presence of MPs in mussels has also been demonstrated during laboratory trials in their faeces, intestinal tract (Von Moos et al., 2012, Van Cauwenberghe et al., 2015, Wegner et al., 2012, Khan and Prezant, 2018), as well as in their circulatory system (Browne et al., 2008). Other laboratory studies showed several effects of microplastic ingestion in laboratory exposed mussels, including histological changes, inflammatory responses, immunological alterations, lysosomal membrane destabilization, reduced filtering activity, neurotoxic effects, oxidative stress effects, increase in hemocyte mortality, dysplasia, genotoxicity and transcriptional responses (reviewed by Li et al., 2019). However, the tested concentrations of MPs in laboratory experiments are frequently unrealistic, being several orders of magnitude higher (2 to 7 orders of magnitude) than the observed seawater concentrations (Van Cauwenberge et al., 2015, Lenz et al. 2016).

Mussels, through their extensive filtering activity, feed on planktonic organisms that have similar size with MPs (Browne et al., 2007) and considering also their inability to select particles with high energy value (i.e. phytoplankton) during filtration (Vahl, 1972, Saraiva et al., 2011a), they are directly exposed to MPs’ contamination. Recent studies suggest a positive linear correlation between MPs concentration in mussels and surrounding waters (Capolupo et al., 2018, Qu et al. 2018, Li et al. 2019). The filtering activity of mussels, which directly affects the resulting MPs accumulation, is a complicated process that is controlled by other factors (food availability, temperature, tides etc.).

The purpose of the present work is to study the accumulation of MPs in the mussels and reveal relations between the accumulated concentrations in mussels’ soft parts and environmental features. In this context, an accumulation model was developed based on Dynamic Energy Budget theory (DEB, Kooijman , 2000) and applied in two different regions, in two different modes of life (wild and cultivated): in the North Sea (Mytilus- eEdulis (M. edulis), wild) and in the Northern Ionian Sea (Mytilus- gGalloprovincialis (M. galloprovincialis), cultivated). DEB theory provides all the necessary detail to model the feeding processes and aspects of the mussel metabolism, taking into account the impact of the environmental variability on the simulated individual. Apart from modelling the growth of bivalves (Rosland et al., 2009, Sara et al., 2012, Thomas et al., 2011,
Saraiva et al., 2012, Hatzonikolakis et al., 2017, Monaco & McQuaid, 2018), DEB models have been used to study other processes as well, such as bioaccumulation of PCBs (Polychlorinated Biphenyls) and POPs (Persistent Organic Compounds) (Zaldivar, 2008), trace metals (Casas and Bacher, 2006) and the impact of climate change on individual’s physiology (Sara et al., 2014). However, to our knowledge this is the first time that a DEB-based model is used to assess the uptake and excretion rates of MPs in mussels.

2. Materials and Methods

2.1 Study areas and field data

The North Sea is a marginal large semi-enclosed sea on the continental shelf of north-west Europe with a total surface area of 850,000 km² and is bounded by the coastlines of 9 countries. The sea is shallow (mean depth 90 m), getting deeper towards the north (up to 725 meters) and the semi-diurnal tide (tidal range 0-5 m) is the dominant feature of the region (Otto et al., 1990). Major rivers, such as Rhine, Elbe, Weser, Ems and Thames discharge into the southern part of the sea (Lacroix et al., 2004), making this area a productive ecosystem. In this study, the area is limited along the French, Belgian and Dutch North Sea coast (N 50.98°-51.46°, W 1.75°-3.54°). This is located close to harbors, where shipping, industrial and agricultural activity is high, putting considerable pressure on the ecological systems of the region (Van Cauwenberghe et al., 2015).

The MPs concentration in mussels’ tissue and seawater that were used to validate and force the model respectively at its North Sea implementation were derived from Van Cauwenberghe et al. (2015). Van Cauwenberghe et al. (2015) examined the presence of MPs in wild mussels (M. edulis), and thus collected both biota and water at 6 sampling stations along the French, Belgian and Dutch North Sea coast in late summer of 2011. M. edulis (mean shell length: 4 ± 0.5 cm and wet weight (w.w.): 2 ± 0.7 g) and water samples were randomly collected on the local breakwaters, in order to assess the MPs concentration in the organisms and their habitat. MPs were present in all analyzed samples, both organisms and water. Seawater samples (N=12) had MPs (<1mm) on average 0.4 ± 0.3 particles L⁻¹ (range: 0.0 – 0.8 particles L⁻¹) and M. edulis contained on average 0.2 ± 0.3 particles g⁻¹w.w. (or 0.4 ± 0.3 particles individual⁻¹) (Van Cauwenberghe et al., 2015). The size range of MPs found within the mussels was 20-90 μm (size <1 mm).
The Northern Ionian Sea is located in the transition zone between the Adriatic and Ionian Sea. The long and complex coastline, presents a high diversity of hydrodynamic and sedimentary features. Rivers discharging into the Northern Ionian Sea include Kalamas/Thyamis (Greece) and Butrinto (Albania) (Skoulikidis et al., 2009), making the area suitable for aquaculture. Small farming sites and shellfish grounds are operating in Thesprotia (northwestern Ionian Sea) (Theodorou et al., 2011). The main source of marine litter inputs in the area originates from anthropogenic activities that mainly include shoreline tourism and recreational activities, poor wastewater management, agricultural practices, fisheries, aquacultures and shipping (Vlachogianni et al., 2017; Digka et al., 2018a). According to Politikos et al. (2020), the area around the Corfu island (Northern Ionian Sea) is characterized as a retention area of litter particles probably due to the prevailing weak coastal circulation. Furthermore, a northward current on the east Ionian Sea facilitates the transfer of litter particles towards the Adriatic Sea, which has been characterized as a hotspot of marine litter and one of the most affected areas in the Mediterranean Sea (Pasquini et al., 2016, Vlachogianni et al., 2017, Liubartseva et al., 2018, Politikos et al., 2020).

The field data used to validate the model output in the N. Ionian Sea were obtained from Digka et al. (2018b, 2018a). In the framework of the “DeFishGear” project, mussels (M. galloprovincialis) were collected by hand from a long line type mussel culture farm in Thesprotia (N 39.606567° E 20.149421°), in summer 2015 (end of June-July) at a sampling depth up to 3 m (Digka et al., 2018a). The average MPs accumulation was calculated from a total population of 40 mussels originated from the farm, with 18 of them found contaminated with MPs (46.25%). The average load of MPs (size <1 mm) per mussel (mean shell length 5.0 ± 0.3 cm) was 0.9 ± 0.2 particles individual⁻¹ and the size of MPs found in the mussel’s tissue ranged from 55 to 620 μm. Both clean and contaminated mussels were included in the calculated mean value in order to represent the mean state of the contamination level for the individual inhabiting the study area.

The seawater concentration of MPs for the N. Ionian Sea implementation was obtained from Digka et al. (2018b) and the DeFishGear project results (http://www.defishgear.net/project/main-lines-of-activities). In total, 12 manta net tows were conducted in the region, collecting a total number of n1=2,027 particles on October 2014 and n2=1,332 on April 2015, leading to an average of 280 particles per tow with size <1 mm and >330 μm (Digka et al., 2018b). In order to estimate the mean MPs concentration in the region, expressed as particles per volume, the dimensions of the manta net (W 60 cm H 24 cm, rectangular frame opening, mesh size 330 μm) and the sampling distance of each tow (~2 km) were used by multiplying the sample surface of the net by the trawled distance in meters (Maes et al., 2017), which resulted in a mean MPs concentration of 1.17 particles m⁻³ (233,333 particles km⁻²). Moreover, in the wider region of the Adriatic Sea, Zeri et al. (2018) found a mean density of 315,009 ± 568,578 particles km⁻² (1.58 ± 2.84 particles m⁻³), out of which
34% sized <1 mm. A relatively high value of standard deviation (one order of magnitude higher than the mean value) is adopted (0.0012 ± 0.024 particles L$^{-1}$), considering that the mussel farm is established in an enclosed gulf and close to the coast, since, according to Zeri et al. (2018), the abundance of MPs is one order of magnitude higher in inshore (<4 km) compared to offshore waters (>4 km). Furthermore, it may be assumed that the adopted range (standard deviation is also multiplied by a factor of 2) includes also the smaller particles sized between 50 μm and < 330 μm, which have been found in mussel’s tissue (Digka et al., 2018a), but were overlooked during the seawater sampling due to the manta net’s mesh size (> 330 μm). According to Enders et al. (2015) the relative abundance of small particles (50-300 μm) compared to particles larger than 300 μm is approximately 50%.

2.2 DEB model description

In the present study, a Dynamic Energy Budget (DEB, Kooijman, 2000, 2010) model is used as basis to simulate the accumulation of MPs by mussels. In DEB theory (Kooijman, 2000), the energy assimilated through food by the simulated individual is stored in a reserve compartment from where a fixed energy fraction $\kappa$ is allocated for growth and somatic maintenance, with a priority for maintenance. The remaining energy ($1 - \kappa$) is spent on maturity maintenance and reproduction. The individual’s condition is defined by the dynamics of three state variables: energy reserves E (joules), structural volume V (cm$^3$) and energy allocated to reproduction R (joules). The energy flow through the organism is controlled by the fluctuations of the available food density and temperature characterizing the surrounding environment.

The DEB model implemented here is an extended version of the model described in Hatzonikolakis et al. (2017), where the growth of the Mediterranean mussel is simulated by taking into account only the assimilation rate of the individual. Since the present study focuses on simulating the MPs accumulation, it is crucial to include a detailed representation of the mussel’s feeding mechanism. In this context, the DEB model was extended by including the clearance ($C_r$), filtration ($\dot{p}_{XIF}$) and ingestion ($\dot{p}_{XII}$) rates of the mussel, following Saraiva et al. (2011a), assuming that all parameters referred to silt (or inedible particles) are applicable also to MPs particles represented by the silt variable. In this approach, a pre-ingestive selection occurs between filtration and ingestion, returning the rejected material in the water through pseudofaeces ($J_{pfi}$). Consequently, energy is assimilated through food while the non-assimilated particles are excreted through the faeces production ($J_f$). The model’s equations, variables and parameters are shown in Table 1, 2 and 3 respectively. The scaled functional response $f$ (Eq. 5, Table 1), which regulates the
assimilation rate, is modified following Kooijman (2006) to include an inorganic term representing the non-digestible matter i.e. microplastics: \( f = \frac{X}{(X + K_y)} \) and \( K_y = X_K \cdot (1 + Y/Y_K) \) where \( Y \) and \( Y_K \) are the concentration of MPs, converted from particles \( L^{-1} \) to \( g \text{ m}^{-3} \) (Everaert et al., 2018) and the half saturation coefficient of inorganic particles here represented by MPs (\( g \text{ m}^{-3} \)), respectively. Thus, the assimilation rate that is regulated by \( f \) is decreasing when the concentration of MPs is increased. The same approach is followed by other authors who considered inedible particles in the mussel’s diet (Ren, 2009, Troost et al., 2010). During the filtration process the same clearance rate for all particles is used (\( \hat{C}_r \)), representing the same searching rate for food that depends on the organism maximum capacity (\( \hat{C}_{rm} \)) and environmental particle concentrations (Vahl, 1972, Widdows et al., 1979, Cucci et al., 1989). During the ingestion process the mussel is able to selectively ingest food particles and reject inedible material, in order to increase the organic content of the ingested material ((Kiørboe & Møhlenberg, 1981, Jørgensen et al., 1990, Prins et al., 1991, Maire et al., 2007, Ren, 2009, Saraiva et al., 2011a). This selection is reflected by the different binding probabilities adopted for each type of particle (\( \rho_l \) for algae particles and \( \rho_i \) for inorganic particles i.e. MPs, see Eq. 14 and table 3). The equations representing the feeding processes handle each type of particle separately, while there is interference between the simultaneous handling of different particle types (Eq. 12-14, Table 1) (Saraiva et al., 2011a). Finally, during the assimilation process, suspended matter (i.e. MPs) that the mussel is not able to assimilate due to its different chemical composition from the reserve compartment (Saraiva et al., 2011a) or incipient saturation at high algal concentrations (Riisgard et al., 2011) results in the faeces production (Eq. 16, Table 1).

### 2.3 Microplastics accumulation sub-model

With the DEB model as a basis, a sub-model describing the microplastics (MPs) accumulation by the mussel was developed, assuming that the presence of MPs in the ambient water does not cause a significant adverse effect on the organisms’ overall energy budget, in accordance with laboratory experiments, conducted in mussel species (Van Cauwenbergh et al., 2015: *Mytilus edulis*, Santana et al., 2018: mussel *Perna perna*). Additionally, it was assumed that the mussel filters MPs present in the water, without the ability of selecting between the high energetic valued particles and the MPs during the filtration process (Van Cauwenbergh et al., 2015, Von Moos et al., 2012, Browne et al., 2008, Digka et al., 2018a among others). The uptake of MPs from the environment is taken into account through the process of clearance/filtration rate, while the excretion of the contaminant is derived from two processes: (i) pseudofaeces production and (ii) faeces production. The resulting MPs accumulation is influenced by external environmental factors.
(MPs concentration, food availability, temperature) and internal biological processes (clearance, filtration, ingestion, growth). All these are described by the following differential equation describes the change of the individual MPs accumulation \( C \), particles individual\(^{-1}\), taking into account the processes mentioned above:

\[
\frac{dC}{dt} = C_{\text{env}} \cdot \dot{C}_R - \dot{j}_{pf2} - k_f \cdot \frac{j_f}{p_{X11}} \cdot C
\]

\text{(Eq. 18)}

where \( \dot{C}_R \) is the clearance rate for water (L h\(^{-1}\)), containing a concentration of MPs \( C_{\text{env}} \) (particles L\(^{-1}\)). The terms of \( \dot{j}_{pf2} \) and \( \frac{j_f}{p_{X11}} \) represent the elimination rate of MPs through pseudofaeces (particles h\(^{-1}\)) and the non-dimensional rate of faeces production with respect to the ingestion rate, respectively (see Table 1, Eq. 15-16). The parameter \( k_f \) represents the post-ingestive selection mechanism utilized by the mussel to incorporate indigestible material (i.e. MPs) into faeces and was calibrated using the available field data of mussel’s MPs accumulation from both study areas (Table 3). Mussel is able to discriminate among particles in the gut based on size, density and chemical properties of the particles (i.e. between microalgae and inorganic material) and thus to eliminate them through faeces (Ward et al., 2019a and references therein). In this context, the pseudofaeces production incorporates the rejected MPs prior to the ingestion, while the faeces production includes MPs that are rejected along with the food particles that are not assimilated by the mussel.

The model’s time step has been set to one hour in order to capture the dynamics of the rapidly changing processes, such as feeding and excretion.

The accumulation of MPs in the individual is represented by the state variable \( C \) (particles individual\(^{-1}\)) which is computed at every model time step. This has been set to one hour, in order to properly resolve the dynamics of the rapidly changing processes, such as feeding and excretion.

### 2.4 Environmental drivers

Besides MPs concentration in the seawater, the DEB model is forced by sea surface temperature (SST) and food availability, represented defined by chlorophyll-a concentrations (CHL-a, an index of phytoplankton biomass). *M. edulis* has been demonstrated to filter suspended particles greater than 1\( \mu \)m, a size class that includes all of the phytoplankton, zooplankton and much of the detritus (Vahl, 1972, Mohlenberg and Riisgård, 1978, Saraiva et al., 2011a, Strohmeier et al., 2012), including even aggregated picoplankton-size particles (i.e. marine snow) (Kach and Ward, 2008, Ward and Kach, 2009). CHL-a has been considered the most reliable food quantifier for the calculation of DEB shellfish parameters (Pouvreau et al., 2006, Sara et al., 2012).
Hatzonikolakis et al. (2017 and references therein) have tested the performance of the model, considering also particulate organic carbon (POC) in the mussel’s diet, which, however, did not have an important impact on the model’s skill against field data in the Mediterranean Sea study areas. This outcome agrees with Troost et al. (2010) demonstration that POC contributes to the mussel’s diet when CHL-a concentrations are low at the southwest of Netherlands. Thus, in the present study, only CHL-a is considered as the available food source for mussels originated from the Southern North Sea and the Northern Ionian Sea. Thus, only CHL-a is considered as the available food source. For both study areas SST and CHL-a are derived from daily satellite data, a method also used by other authors (i.e. Thomas et al., 2011, Monaco & McQuaid, 2018).

In the North Sea, SST data were obtained from daily satellite images provided by Copernicus Marine Environmental Monitoring Service (CMEMS) at 0.04 degree spatial resolution and CHL-a data obtained from the Globcolour daily multi-sensor product provided by CMEMS-Globcolour database at 1 km spatial resolution based on the OC5 algorithm of Gohin et al. (2002) (http://marine.copernicus.eu/, generated using CMEMS Products, production center ACRI-ST). The environmental forcing data (SST, CHL-a) were averaged over the study area (51.08°-51.44° N, 2.19°-3.45° E), covering the period 2007-2011 (5 years), in order to realistically simulate the wild mussel’s growth harvested in late summer 2011 (Van Cauwenberghe et al., 2015). It is notable that the study area of the North Sea belongs to CASE II waters (coastal region), where algorithms tend to overestimate CHL-a concentrations. In optically-complex Case II waters, CHL-a cannot readily be distinguished from particulate matter and/or yellow substances (dissolved organic matter) and so global chlorophyll algorithms are less reliable (IOCCG, 2000). However, the CHL-a dataset that was used was found in good agreement with available in situ data from ICES database (https://www.ices.dk/data/Pages/default.aspx) for the specific study area and time period, showing a relatively smaller bias and better time-space coverage, as compared with other tested remote sensing datasets (not shown) (i.e. regional chlorophyll product available for the North West Shelf-Seas in the CMEMS catalogue, https://resources.marine.copernicus.eu/).

In the North Ionian Sea, daily satellite SST data were also obtained from the CMEMS database for the Mediterranean Sea with 0.04 degree spatial resolution, while CHL-a daily data were derived from the Globcolour multi-sensor (i.e. SeaWiFS, MERIS, MODIS, VIIRS and OLCI-a) merged product (http://globcolour.info) at 1 km spatial resolution based on the OC5 algorithm suitable for coastal regions (Gohin et al., 2002) while CHL-a data were derived from the merged product of many satellites (i.e. SeaWiFs, Meris, Modis, Viirs and Olci-a) provided by Globcolour web interface (http://globcolour.info) at a daily temporal resolution and 1 km spatial resolution. The forcing data were averaged over the study area (N 39.49°-39.65°, E 20.09°-20.23°) covering the
period 2014-2015 (2 years), when the cultured mussel is ready for the market. The chosen CHL-a dataset was found preferable, as compared with other available remote sensing datasets (i.e. CMEMS chlorophyll product for Mediterranean Sea), since it presented a better spatial and temporal coverage (Hourany et al., 2019, Garnesson et al., 2019) and a slightly lower error, as compared with the very few available in situ data in the study area (not shown). Unfortunately, these were very scarce and therefore an extended comparison between remote and in situ data could not be conducted. The satellite-derived CHL-a data were estimated based on the OC5 algorithm of Gohin et al. (2002) in both study areas, which is regarded as suitable for coastal waters. Satellite data have facilitated large scale ecological studies by providing maps of phytoplankton functional types and sea surface temperature (Raitos et al., 2005, 2008, 2012, 2014, Palacz et al., 2013, Di Cicco et al., 2017, Brewin et al., 2017). The daily environmental forcing data are shown in Fig. 1 and Fig. 2 for the North Sea and the N. Ionian Sea, respectively. The two coastal environments present some important differences regarding both CHL-a and SST. Specifically, in the N. Ionian Sea, CHL-a is relatively low (annual mean ~ 0.88 mg chl-a m$^{-3}$) and peaks during winter (maximum ~2.64 mg chl-a m$^{-3}$ at December 2014), while in the North Sea CHL-a is about four times higher (annual mean 4.25 mg chl-a m$^{-3}$), peaking in April every year (maximum range 29.44-33.38 mg chl-a m$^{-3}$), as soon as light availability reaches a critical level (Van Beusekom et al., 2009). The higher productivity during spring season in the North Sea is related with the nutrient inputs from the English Channel, the North Atlantic and particularly the river discharge of nutrient-rich waters along the Belgian-French-Dutch coastline, which peaks earlier, during winter period (Van Beusekom et al., 2009). The SST sea surface temperature peaks during August in both areas (Fig. 1 and Fig. 2), but is significantly higher in the N. Ionian Sea (maximum 28.8°C), as compared to the North Sea (maximum 18-19.3°C).

The environmental concentration of MPs, $C_{env}$ (particles L$^{-1}$) was obtained also at a daily time step as randomly generated values of the Gaussian distribution that is determined by the mean value and standard deviation of the observed field data (0.4 ± 0.3 particles L$^{-1}$, North Sea, Van Cauwenbergh et al., 2015, 0.0012 ± 0.024 particles L$^{-1}$, N. Ionian Sea, Digka et al., 2018a). Considering that these values originate from surface waters and that mussels live in the near surface layer (0-5 m), $C_{env}$ is estimated as a mean value of the upper layer with the methods described by Kooi et al. (2016), who studied the vertical distribution of MPs, considering an exponential decrease with depth. Specifically, in the N. Ionian Sea, mussels were collected from a depth up to 3 m (Digka et al., 2018a), while in the North Sea (Van Cauwenbergh et al., 2015), there is no information and thus a maximum depth of 5 m is adopted.

In the North Sea simulation, the effect of tides is taken into account by considering that the mussel originated from the intertidal zone, is submerged 12 hours during the day (Van
In the N. Ionian Sea simulation, tides are not considered, given the very small tide amplitude (few centimeters) in the Mediterranean (i.e. Sara et al., 2011; Hatzonikolakis et al., 2017) and thus the cultured mussel is assumed permanently submerged. *In situ* hourly tide data (2007-2011) from the coastal zone of the region (Dunkerque station N 51.04820°, E 2.36650°) obtained from Coriolis and Copernicus data provider (http://marine.copernicus.eu, http://www.coriolis.eu.org), showed that mussels experience alternating periods of aerial exposure and submergence at approximately every 6 hours (2 high and 2 low tides). During aerial exposure the model suspends the feeding processes (Sara et al., 2011) and simulates metabolic depression (Monaco & McQuaid, 2018) where, the Arrhenius thermal sensitivity equation (Eq. 9) is corrected by a metabolic depression constant ($M_d = 0.15$), a value representative for *M. galloprovincialis* and here applied also for *M. edulis*. In the present study, the mussel’s body temperature change during low tide is ignored, inducing a model error. The mussel’s body temperature (i.e. surrounding water temperature for submerged mussels) during air exposure depends on many factors, such as solar radiation, air’s temperature, wind speed and wave height, according to studies investigating the temperature effect on intertidal mussels (Kearney et al., 2010, Sara et al., 2011). However, the present study aims to primarily examine the MPs accumulation and thus the intertidal mussel’s body temperature was not thoroughly examined. Nonetheless, the time that the mussel is able to filter, ingest and excrete the suspended matter (i.e. food and MPs particles) and the effect on the mussel’s growth through the modified relation of $k(T)$ are included, since the assimilation process occurs whether the mussel is submerged or not (Kearney et al., 2010).

### 2.5 Parameter values

Most of the DEB model parameters were obtained from Van der Veer et al. (2006) and are referred to the blue mussel *Mytilus edulis* in the northeast Atlantic (see Table 3 for the exceptions). This assumption has also been adopted in previous studies which showed that this parameter set for *M. edulis* applies also for *M. galloprovincialis* (i.e. Casas and Bacher, 2006, Hatzonikolakis et al., 2017). The half saturation coefficient $X_k$ represents the density of food at which the food uptake rate reaches half of its maximum value and should be treated as a site–specific parameter (Troost et al., 2010, Pouvreau et al., 2006). In order to estimate the value of $X_k$, a different approach was followed for each study area.

For the North Sea simulation, $X_k$ was tuned so that the simulated individual has the recorded size at the corresponding estimated age (Van Cauwenberghe et al., 2015) growing with the representative growth rates of wild *M. edulis* at the region (Saraiva et al., 2012, Sukhotin et al., 2007). For the N. Ionian Sea simulation, an alternative method was adopted, aiming to generalize
the DEB model to overcome the problem of site-specific parameterization. The DEB model was tuned against literature field data for cultured mussels originated from different areas in the Mediterranean and Black Seas, where the average CHL-a concentration ranged between 1.0 and 5.0 mg chl-a m\(^{-3}\), and one \(X_k\) value was found for each area. The four areas used, their characteristics and the corresponding value of \(X_k\) adopted, are shown in Table 4. These values of \(X_k\) are related to the prevailing CHL-a concentration of each area ([CHL-a]) through three different functions: linear: \(f(x) = a \cdot [\text{CHL} - a] + b\) exponential: \(f(x) = a \cdot \exp(b \cdot [\text{CHL} - a])\) and power: \(f(x) = a \cdot [\text{CHL} - a]^b + c\). The curve fitting app of Matlab (Matlab R2015a) was used for the determination of \(a, b\) and \(c\) of each function taking into account the 95\% confidence level. The score of each function regarding the somatic/mussel growth simulation in all four regions is tested through target diagrams (Jolliff et al., 2009) by computing the bias and unbiased root-mean-square-deviation (RMSD) between field and simulated data of all 4 regions and the function with the best score is adopted. A similar approach was followed by Alunno-Bruscia et al. (2011) for the oyster *Crassostrea gigas* in six Atlantic ecosystems who expressed the \(X_k\) as a linear function of food density (e.g. phytoplankton). Unfortunately, the approach described for the N. Ionian Sea simulation could not be applied in the North Sea, as the limited amount of growth data from the literature for wild *M. edulis* in similar environments did not permit a statistically significant fit of a similar function (\(X_k = f(chl - a)\)).

### 2.6 Simulation of reproduction-Initialization of the model

The reproductive buffer (R) is assumed to be completely emptied at spawning (\(R = 0\)) (Sprung, 1983, Van Haren et al., 1994). In order to simulate mussel’s spawning, the gonado-somatic index (GSI) defined as gonad dry mass over total dry flesh mass was computed at every model’s time step (Eq. 17 Table 1; the water content of the fresh tissue mass was assumed 80\% according to Thomas et al. (2011)). Spawning was induced by a critical value of GSI (GSI\(_{th}\), Table 3) and a minimum temperature threshold (\(T_{th}\)) at each study area, obtained from the literature. In the North Sea implementation, \(T_{th}\) was set at 9.6 °C (Saraiva et al., 2012), while in the N. Ionian Sea, at 15 °C (Honkoop and Van der Meer, 1998). This kind of formulation for the spawning event in bivalves has been used in previous studies (i.e. Pouvreau et al., 2006, Troost et al., 2010, Thomas et al., 2011, Monaco & McQuaid, 2018). The simulated abrupt losses of the mussel's tissue mass correspond to spawning events and the model’s prediction was compared with the available literature data regarding the spawning period in each study area. Theodorou et al. (2011) demonstrated that the spawning events occur during winter for *M. galloprovincialis* in the mussel
farms of Greece, while in the North Sea the spawning period for *M. edulis* is extended from the end of April until the end of June (Sprung, 1983, Cardoso et al., 2007).

In both areas, the model was initialized so that the simulated individual is in the juvenile phase (*V* < *V*<sub>p</sub>; Table 3) and the reproductive buffer can be considered to be empty (*R* = 0) (Thomas et al., 2011). As stated by Jacobs et al. (2015) amongst others, juvenile mussels (*M. edulis*) range between 1.5-25 mm in size. Specifically, in the North Sea the settlement of mussel larvae (*M. edulis*) takes place in June and the juveniles grow to a maximum size of 25 mm within 4 months (Jacobs et al., 2014). In the N. Ionian Sea, the operating mussel farms follow the life cycle of *M. galloprovincialis*, starting the operational cycle each year by dropping seed collectors from late November until March and the juvenile mussels grow up to 6-6.5cm after approximately one year according to the information obtained from the local farms in the region and Theodorou et al. (2011). The initial fresh tissue mass was distributed between the structural volume (*V*) and reserves energy (*E*). Energy allocated to those two compartments was firstly constrained by the initial length (*L*) and then energy allocated to *V* was in Eq.10 (Table 1). The initial value of *E* was set so that the simulated individual has an initial weight that corresponds to the juvenile phase (*V* < *V*<sub>p</sub>) (Table 5). Finally, for both model implementations, the initial accumulation of MPs in the mussel’s tissue (*C*) was set to zero.

### 2.7 Simulation Runs

The DEB-accumulation model simulates at an hourly basis the growth and MPs accumulation of the wild mussel from the North Sea and the cultured mussel from the N. Ionian Sea. Initially, a model run is performed at each study area during the periods July 2007 to August 2011 (4 years) for the North Sea simulation and late November 2014 to January 2016 (~ 1 year) for the N. Ionian Sea simulation. Additionally, the inverse simulations were performed in order to evaluate the depuration phase of both cultured and wild mussel, by setting the environmental MPs concentration equal to zero (*C*<sub>env</sub>=0), after a period of 1 year simulation at the N. Ionian Sea, when the cultured mussel has the appropriate size for market, and after 4 years at the North Sea, when literature field data are available (Van Cauwenberghe et al., 2015). In this simulation, the mussel’s gut clearance is achieved by the excretion of MPs through faeces (3<sup>rd</sup> term of Eq. 18), and thus it is necessary to maintain the existence of food in the mussel’s environment in order to ensure that the feeding-excretion processes will occur.

Furthermore, to examine the model’s uncertainty related to the environmental MPs concentration, a series of 15 and 13 simulations were performed in the North Sea and N. Ionian Sea.
respectively, adopting different constant values of $C_{env}$ within the observed range of each area.

Finally, the effect of the environmental forcing data and some model’s parameters on the resulting MPs accumulation by both mussels was explored through sensitivity experiments. These were used to derive a new function that predicts the level of MPs pollution in the environment.

### 2.8 Sensitivity tests and Regression analysis

The effect of the environmental data (CHL-a, temperature, $C_{env}$) and two parameters representative of mussel’s growth ($X_k$, $Y_k$) on the MPs accumulation by the mussel for each study area was examined through sensitivity experiments with the DEB-accumulation model. Each variable (CHL-a, T, $C_{env}$) and parameter ($X_k$, $Y_k$) was perturbed by ±10% to examine their effect on the simulated MPs accumulation, and the results of each run were analyzed using the sensitivity index ($SI$) which calculates the percentage change of the mussel’s MPs accumulation $SI = \frac{1}{n} \sum_{t=1}^{n} \left( \frac{C_{t}^1 - C_{t}^0}{C_{t}^0} \right) \cdot 100 \%$, where $n$ is the simulated time steps, $C_{t}^0$ is the MPs accumulation predicted with the standard simulation at time $t$ and $C_{t}^1$ is the MPs accumulation with a perturbed variable/parameter at time $t$; for details see Bacher and Gangnery, (2006). The same method has been also applied to other studies, which examined the model’s sensitivity on specific variables/parameters regarding the mussel growth (Casas and Bacher, 2006, Rosland et al., 2009, Béjaoui-Omri et al., 2014, Hatzonikolakis et al., 2017). In order to also examine the effect of tides, in the North Sea implementation, the sensitivity experiments were conducted twice: the first time assuming that the mussel is permanently submerged and the second time assuming that the mussel is periodically exposed to the air.

Preliminary sensitivity experiments showed that the MPs accumulation is highly depended on the prevailing conditions regarding the CHL-a, temperature and $C_{env}$ and the mussel’s growth that is regulated by the half saturation coefficient ($X_k$). Therefore an attempt was made using the model’s output to describe the MPs accumulation as a function of these variables through a custom regression model:

$$y = b_1 \cdot W + b_2 \cdot \exp \left( \frac{1}{T} \right) + b_3 \cdot \frac{1}{[CHL-a]} + b_4 \cdot C_{env}$$  \hspace{1cm} (Eq. 19)

where $y$ (particles/individual) is the response variable and represent the predicted MPs accumulation by the mussel; $W$ (g) the mussel’s fresh tissue mass, $T$ (K) the sea surface temperature, CHL-a and $C_{env}$ are the concentrations of chlorophyll-a and MPs in the water respectively, which are the predictor variables. The values of coefficients $b_1$, $b_2$, $b_3$, and $b_4$ are
calculated using the nonlinear regression function (nlinfit, Matlab R2015a) which attempts to find values of the parameters $b$ that minimize the least squared differences between the model’s MPs accumulation output $C$ and the predictions of the regression model $y = f(W, T, [chl~a], Cenv, b)$.

The ultimate aim of this analysis, once coefficients are determined, is to use Eq. 19 to obtain the environmental MPs concentration:

$$C_{env} = \frac{1}{b_4} \left( C - b_1 * W - b_2 * \exp \left( \frac{1}{T} \right) - b_3 * \frac{1}{[CHL-a]} \right)$$

(Eq. 20)

which could be a very useful tool to predict the MPs concentration in the environment, when all involved variables are known (mussel size, mussel’s accumulated MPs ($C$), wet weight ($W$), temperature ($T$) and CHL-a), using the mussel as a potential bioindicator (Li et al., 2016, Li et al., 2019). The score of this custom model was tested by applying Eq. 20 in our study areas and 6 more areas around the U.K., where information of mussel’s wet weight and both mussels’ and environment’s MPs load is available (Li et al., 2018). CHL-a and temperature, which were not included in Li et al. (2018), were obtained from daily satellite images (same source as in the North Sea, see 2.4 section), covering the period that the mussels were harvested (Li et al., 2018).

### 3. Results

#### 3.1 Growth simulations

The growth simulations of *M. edulis* and *M. galloprovincialis* for the North Sea and the N. Ionian Sea are shown in Fig. 34 and Fig. 45 respectively. In the North Sea implementation, $X_k$ was tuned to a constant value: $X_k=8 \text{ mg~chl-a~m}^{-3} (\pm 1.5 \text{ mg~m}^{-3})$. The fitted value was slightly higher, as compared to the one ($X_i=3.88 \text{ mg~m}^{-3} \text{ μg~chl-a~l}^{-1}$) used by Casas and Bacher (2006) in productive areas of the French Mediterranean shoreline (average CHL-a concentration $1.45 \text{ mg~m}^{-3} \text{ μg~chl-a~l}^{-1}$ maximum peak at $20 \text{ mg~m}^{-3} \text{ μg~chl-a~l}^{-1}$), as a consequence of given the even higher productivity in the North Sea (average CHL-a concentration $4.25 \text{ mg~m}^{-3} \text{ μg~chl-a~l}^{-1}$; maximum peak at ~$33.40 \text{ mg~m}^{-3} \text{ μg~chl-a~l}^{-1}$). The high value of $X_k$ -could also be explained by the presence of silt and other inedible particles (i.e. MPs) which led to lower quality food in the mussel’s diet compared with an assumed “clean of inedible particles” site environment (Kooijman, 2006, Ren, 2009). In the present study the inedible particles (i.e. MPs) have been incorporated in the mussel’s diet through
the modified relation of the functional response $f$ (Eq. 5, Table 1), which regulates the assimilation rate and thus the mussel’s growth. However, the DEB model applied at the French site, did not account inedible particles in the mussel’s food. Furthermore, it has been reported that wild mussels grow considerably slower than farmed mussels (~1.7 times) (Sukhotin and Kulakowski, 1992) and thus, a higher value of $X_k$ promotes a lower mussel growth, which is the case of the North Sea mussel. The simulated mussel shell length after 4 years, in August, is 4.35 cm and the fresh tissue mass is 1.87 gr, in agreement with Van Cauwenbergh et al. (2015) and other studies conducted on wild mussels (Sukhotin et al., 2007, Saraiva et al., 2012, MarLIN, 2016). In particular, Saraiva et al. (2012) found that after 16 years of simulation, the wild mussel of the Wadden Sea (North Sea) is 7 cm long, while according to Bayne and Worral (1980) a mussel with shell length 4 cm corresponds to the age of 4 years, in agreement with the current study. The simulated growth presents a strong seasonal pattern, being higher during spring and summer season, as compared to autumn and winter, which is consistent with the seasonal cycle of temperature and CHL-a concentration, for a typical year in the region (Fig. 1). The increase of food availability and temperature during spring (April) results in high mussel growth for a 4-month period, while the decrease of CHL-a from summer until the end of the year, in conjunction with the temperature decrease in autumn, result in a lower mussel growth. Spawning events that occurred each year in late between the end of April - early and beginning of May (30 April-2 May) each year, are responsible for the sharp decline in mussel’s fresh tissue mass, shown in Fig. 4 (Handa et al., 2011; Zaldivar, 2008) and in agreement with the literature (Sprung, 1983, Cardoso et al., 2007, Saraiva et al., 2012). The predicted weight loss due to spawning was around 7% at the first year of simulation, while the second, third and fourth year the percentage of weight loss increased gradually to 8.3%, 12.6% and 14.4% respectively. Bayne and Worral (1980) demonstrated that the weight losses on spawning for individuals of 1 g weight vary between 2.1% and 39.8%, presenting a weight-specific increase with size.

In the N. Ionian Sea implementation, $X_k$ is applied as a function of CHL-a concentration through the method described in section 2.5. The target diagram showing the performance of each tested function (linear: $f(x) = a \times [CHL - a] + b$, where $a = 0.959$ and $b = -1.420$; exponential: $f(x) = a \times \exp(b \times [CHL - a])$ where $a = 0.2$ and $b = 0.567$; power: $f(x) = a \times [CHL - a]^b + c$ where $a = 0.01$, $b = 3.529$ and $c = 0.480$) is shown in Figure 5. The linear and power function of $X_k$ present a good skill, with the power function leading to the most successful simulation of the cultured mussel’s growth in all four areas (diagram marks for mussel length and fresh tissue mass are closer to the target’s center). The power function applied in the N. Ionian Sea, resulted in mussel’s shell length 5.8 cm and fresh tissue mass 5.92 gr after one year simulation, in agreement with Theodorou et al. (2011). The spawning event occurred at the
beginning of December (Theodorou et al., 2011) and was illustrated by a 12.6% tissue mass decline.

3.2 Microplastics accumulation and depuration phase

The hourly simulated MPs accumulation by the mussel in the North Sea and N. Ionian Sea are shown in Fig. 6 and Fig. 7 respectively. Calibration of the parameter $k_f$ ($1.2 \text{ d}^{-1}$) led to a model which was well fitted to the observed MPs accumulation in the mussel of both study areas. In the North Sea, a 4-year-old wild mussel ($L=4.35 \text{ cm}, W=1.87 \text{ g}$) contains 0.5364 particles individual$^{-1}$ in August within the range value found by Van Cauwenberghe et al. (2015) (0.4 ± 0.3 particles individual$^{-1}$), although the model overestimated the data range reproducing a seasonal increase that was not observed. This is most likely due to the fact that Van Cauwenberghe et al. (2015) allowed a 24 h clearance period, before analyzing the mussels' tissue for MPs, possibly resulting in slightly lower MPs accumulation than the model’s prediction. The MPs egested through faeces by the 4 year old mussel after 24 h were 0.2 ± 0.2 particles individual$^{-1}$ (Van Cauwenberghe et al., 2015), which agree also with model’s output (0.3 particles individual$^{-1}$, Fig. 8) regarding the depuration phase and could compensate for the observed difference in mussel’s MPs load between the simulated and field data. In the N. Ionian Sea, the simulated MPs accumulation by the cultured mussel with $L=4.858 \text{ cm}$ and $W=3.343 \text{ g}$ was 0.91 particles individual$^{-1}$ in the end of JuneJuly, in agreement with field observations obtained from Digka et al. (2018a) (0.9 ± 0.2 particles individual$^{-1}$). Overall, the developed model simulated the MPs accumulation by both mussels in the two different areas, using the same parameter set (see Table 3 for the exceptions), under the assumption that parameters referred to silt particles (i.e. inedible particles) may be used to describe also the MPs accumulation. Both simulations were in good agreement with the available field data, with a small deviation for the North Sea. In both regions, the model computed MPs accumulation, assuming that the mussel treats MPs as silt particles (i.e. inedible particles) and is in agreement with the available field data. This may lead to the assumption suggests that mussels—probably present a common behavior against all inedible particles. In model’s results, based on the uptake and excretion rates of MPs by the mussels in both study areas, the majority of MPs are rejected through pseudofaeces and fewer through faeces production (not shown). This is in agreement with Woods et al. (2018) who found that most microplastic fibers (71%) were quickly rejected as pseudofaeces and < 1% excreted in faeces.

The small-scale (daily) fluctuations of MPs in the mussel (wild and cultivated) reflect the adopted random variability of the environmental MPs concentration $C_{env}$ and the daily fluctuations...
of the environmental forcing (CHL-a, temperature). The large-scale (seasonal) variability follows mainly the variability of the clearance rate. The seasonal variability of the CHL-a concentration and temperature greatly determines the variability of the clearance rate and hence the variability of MPs in the individual. Moreover, the model predicts that mussel’s energy needs are increased as it grows and therefore the clearance rate is increased, resulting in higher MPs accumulation.

The simulated time needed to clean the mussel’s gut from the MPs load for both areas is shown in Fig. 8. In both areas, the cleaning follows an exponential decay, in agreement with laboratory experiments by Woods et al. (2018). In particular, the model predicts a 90% mussel’s cleaning after 284±30 hours (~1.24 days) and 566±3 hours (~2.53 days) for the N. Ionian Sea and North Sea respectively. The cleaning process is more rapid in the North Sea simulation, which can be attributed to the higher CHL-a concentration found in this area, leading to increased production of faeces by the mussel and hence faster excretion of the accumulated MPs. In the N. Ionian Sea, on the other hand, the rate of the mussel’s cleaning is slower, due to the limited food availability.

3.3 Model’s uncertainty regarding the environmental microplastics concentration

The MPs concentration in the environment presents a strong variability in both temporal and spatial scales. To examine the model’s uncertainty related to the environmental MPs concentration ($C_{env}$), a series of 15 and 13 simulations were performed in the North Sea and N. Ionian Sea respectively, adopting different values of $C_{env}$ within the observed range of each area. In the North Sea, the adopted $C_{env}$ ranged between 0.1 and 0.8 particles L$^{-1}$ with a step of 0.05 (15 runs), while in the N. Ionian Sea $C_{env}$ ranged between 0.0012 and 0.0252 particles L$^{-1}$ with a step of 0.002 (13 runs). The mean seasonal values and standard deviation of the 15 simulations in the North Sea and the mean monthly values and standard deviation of the 13 simulations in the N. Ionian Sea were computed and plotted in Fig. 9 and Fig. 10, respectively. Each error bar represents the uncertainty of the simulated accumulation at the specific time, related to the environmental MPs concentration.

In both case studies, the uncertainty of the model appears to increase as the MPs accumulation is increased. As the mussel grows in the North Sea, the mean value and standard deviation of MPs accumulation is increased during the same season every year, illustrating the effect of the mussel’s weight. Moreover, the seasonal variability of the MPs accumulation appears to be related with should be caused by the seasonality of CHL-a concentration. This is apparent during each year’s spring: when CHL-a concentration peaks at its maximum value (~30 mg m$^{-3}$; see Fig. 1), the filtration rate is decreased (Riisgard et al., 2003, 2011), leading to lower MPs accumulation by the mussel and thus lower model’s uncertainty. In the N. Ionian Sea, the effect of the mussel’s weight is
more apparent in the early months (~ 6 months), resulting on higher MPs accumulation and model uncertainty as the mussel grows. Afterwards, the seasonality of both CHL-a concentration and temperature plays the major role. During summer, when the CHL-a concentration is progressively decreased, reaching minimum values (~0.7 mg/m³) and temperature is increased (>20° C), the filtration rate is significantly decreased or stopped, resulting in lower MPs accumulation and lower model’s uncertainty. This is in line with studies reporting that the mussel suspends the filtering activity and thus closes its valves until better conditions occur (Pascoe et al., 2009, Rissgard et al., 2011). Overall, the available field data lie within the model’s uncertainty, apart from the North Sea case, where the range of field data variability and model uncertainty do not overlap significantly at the time of the observations for both study areas.

Moreover, to evaluate the scenario adopted with the set-up of the previous experiments (random $C_{env}$ at a daily time step) 3 additional model runs are performed in each study area, adopting each time different stochastic sequences of daily random $C_{env}$ values within the observed range, which is considered to reflect the high spatial and temporal variability of the environmental MPs concentration. The mean value and standard deviation of these “stochastic” runs lie most of the time within the standard deviation of the overall model’s uncertainty in both case study areas (Fig. 9 and Fig. 10).

3.4 Sensitivity and Regression analysis results

The results of the sensitivity experiments regarding the MPs accumulation by the mussels are shown in Fig. 11 and 12 for the North Sea and N. Ionian Sea respectively. The comparison between the intertidal and subtidal mussel of the North Sea revealed that both +10% and -10% perturbation of CHL-a and $X_k$ have a slightly lower effect on the MPs accumulation by the intertidal mussel which is probably attributed to the intermittent feeding periods experienced by the individual due to the tide effect. As far as the temperature effect, both +10% and -10% perturbed value led to higher sensitivity on the MPs accumulation by the intertidal mussel, due to the adopted modified temperature relation during low tide. Especially, if the mussel’s body temperature change during air exposure would be considered, the perturbed temperature will probably affect even more the MPs accumulation on the intertidal than the subtidal mussel. The sensitivity effect of the $C_{env}$ is slightly higher and lower on the MPs accumulation by the intertidal mussel when perturbed either +10% or and -10% is almost the same for the intertidal and subtidal mussel respectively, however the difference of the sensitivity index (%) between the two mussels (intertidal vs. subtidal) is small,
indicating that the environmental MPs concentration affects similarly both mussels, regardless the continuous or intermittent feeding-excretion process.

The comparison between the mussel sensitivity indexes in the N. Ionian and the North Sea (in conditions of submergence) study areas reveals some important differences. Generally, most of the perturbed (either +10% or -10%) variables and parameters (i.e. CHL-a, temperature, \(X_k\)) present higher sensitivity on the MPs accumulation by the mussel from the N. Ionian Sea. This is attributed to the prevailing environmental conditions and specifically the lower food availability (CHL-a) and the higher temperature range in the N. Ionian Sea compared to the North Sea, which greatly determine the feeding processes, the mussel’s growth and hence the MPs accumulation. The perturbed \(C_{env}\) in both study areas appears to affect similarly the MPs accumulation on both mussels (\(\approx 10\%\)), with the small difference (\(<2\%) probably attributed to the higher abundance of seawater’s MPs present in the North Sea compared to the N. Ionian Sea. Finally, the half saturation coefficient for the inorganic particles (\(Y_k\)) has no effect on the MPs accumulation of both North Sea and N. Ionian Sea mussels, indicating that the amount of inedible particles (i.e. MPs) is relatively low in both areas and thus the \(Y_k\) does not affect the way that the organic particles are being ingested (Kooijman, 2006). According to Ren (2009), when the inorganic matter is low, the K(y) (Eq. 5; Table 1) is approximately equal to \(X_k\) and then \(Y_k\) is the least sensitive parameter for the ingestion rate and thus growth.

The DEB-accumulation model output was used to determine the coefficients in Eq. 19 by the nonlinear regression analysis: \(b_1= 0.1909 \ (\pm \ 0.0006)\), \(b_2= 0.0412 \ (\pm \ 0.0019)\), \(b_3= 0.1315 \ (\pm \ 0.0021)\) and \(b_4=1.1060 \ (\pm \ 0.0253)\). The accurate estimation of the confidence intervals for the estimated coefficients \((b_1, b_2, b_3, b_4)\) is indicated by the low confidence intervals, while the mean squared error of the regression model appears also sufficiently small are small enough which indicates an accurate estimation of them and the mean squared error of the regression model is small enough (MSE=0.0523). Subsequently, as shown in Figure 13, Equation 20 may be used to predict the MPs concentration of the environment where mussels live, being in most cases, the predicted MPs concentration is found within the standard deviation of the field data. Two exceptions are shown in Hastings-A and Plymouth areas. The reasons behind these discrepancies may be related to the environmental conditions prevailing in each area at the sampling time. For example, Eq. 20 does not take into account the impact of tides that may affected the mussel’s MPs load (C) and the lack of information on the exact sampling date led to using a mean SST and CHL-a value representative of the given sampling time period (Li et al., 2018). Although, Eq. 20 does not account for the tide effect, however, the sensitivity analysis (Fig. 11) showed that the effect of \(C_{env}\) on the mussel’s MPs accumulation was the same for both intertidal and subtidal mussel in the North Sea. This result may also apply at the two exceptions areas, leading to the assumption that the
discrepancies are due to the lack of the ambient temperature and CHL-a information during the sampling date. In any case, however, this is just a first rough demonstration of the method and should be implemented in more environments in order to be further validated.

4. Discussion

A DEB-accumulation model was developed and validated with the only available data from the North Sea and the N. Ionian Sea, to study the MPs accumulation by wild M. edulis and cultured M. galloprovincialis, as they grown in different, representative environments. Although the study is limited by scarce validation data, it should be noted the MPs accumulation model parameter set, except one tuning parameter (k_i), was extracted from the literature (Table 3), assuming it is notable that the accumulation submodel’s parameters are extracted from literature (Table 3) illustrating that mussels adopt a common defensive mechanism against inedible particles (i.e. silt, MPs). Thus, the theoretical background constructed by Saraiva et al., (2011a) (based on Kooyman, 2010) regarding the feeding and excretion processes of the mussel remains unspoiled.

Through the strong theoretical background of DEB theory, this study highlights that the accumulation of MPs by the mussel is highly depended on the prevailing environmental conditions which control the amount of MPs that the mussel filtrates and excretes.

Towards a generic DEB model: Beginning with the generalization of the DEB model regarding the site-specific parameter X_k in the N. Ionian Sea simulation, the applied function of the half saturation coefficient \( f(x) = a \cdot [CHL - a]^b + c \) successfully captures the physiological responses and thus the growth rate of the cultured mussel at the N. Ionian Sea implementation. In the current study, a demonstration of this method led to a robust and generic DEB growth model able to simulate robust enough with a sufficiently generic nature for the simulation of the mussel growth in representative mussel habitats of the Mediterranean Sea, covering a range of productivity and sea surface temperature. This approach supports and takes a step further of Bourles et al. (2008) suggestion about suggested for oyster growth (Crassostrea gigas) that a seasonally varied half saturation coefficient, demonstrating an improvement could improve the accuracy of the food quantifier. The applied function of X_k considers the daily CHL-a fluctuations, and thus the seasonal variation of the seawater composition, because seawater composition is closely related to the season. As more field data becomes available from various environments, the applied such an approach could result to more generic formulations for the site-
specific parameter \( X_k \), so that the model could be applied in several areas of interest, where field growth data are absent and/or to simulate the potential mussel growth in the 2D space.

The simulation of MPs accumulation by the mussels, using the DEB-accumulation model, is in good agreement with the available field data (Fig. 3 and Fig. 4). The simulated values lie within the observed field data range (mean ±SD), although the seasonal increase reproduced by the model at the North Sea implementation did not exactly overlap with the field data at the time of observations. This could be attributed to the clearance period (24 h) that allowed mussels to excrete MPs through faeces (0.2 ± 0.2 particles individual\(^{-1}\)) before the mussel’s tissue analysis (Van Cauwenberghe et al., 2015). The measured loss of mussel’s MPs is in agreement with the model’s result on the depuration experiment after 24 h. The MPs accumulation by the cultivated mussel (fresh tissue mass 3.3342 g) originated from the N. Ionian Sea with mean \( C_{env}= 0.0012 ± 0.024 \) particles L\(^{-1}\), is 0.91 particles individual\(^{-1}\) and by the wild mussel (fresh tissue mass 1.87 g) from the North Sea with mean \( C_{env}= 0.4 ± 0.3 \) particles L\(^{-1}\) is 0.5364 particles individual\(^{-1}\). If these concentrations are expressed per gram of wet tissue of mussels, the cultivated mussel contamination (0.27 particles g\(^{-1}\)w.w.) is comparable with the wild mussel (0.2834 particles g\(^{-1}\)w.w.), despite the much lower environmental MPs concentration (\( C_{env} \)) in the N. Ionian Sea than the North Sea. This comparison aims to highlight the significant impact of the prevailing environmental conditions (CHL-a and temperature) on the MPs accumulation by the mussels, although they originate from different areas and lived different time period. The generally high abundance of CHL-a in the North Sea simulation, contributes to a reduction of the filtering activity and hence of the MPs accumulation. The threshold algal concentration for reduction of the mussel’s filtration rate (incipient saturation) has been found to lie between 6.3 and 10.0 mg m\(^{-3}\)μg chla L\(^{-1}\) (Riisgard et al., 2011), which is a range comparable to the CHL-a concentrations in the North Sea case. Furthermore, in the N. Ionian Sea simulation, the filtration, ingestion, pseudofaeces and faeces production rates are decreased during the summer season when the CHL-a and temperature has downward and upward trend respectively, gradually leading to a decline of the mussel’s MPs accumulation. Van Cauwenberghe and Janssen (2014) found that cultivated \( M. edulis \) from the North Sea contained on average 0.36 ± 0.07 particles g\(^{-1}\)w.w., a slightly higher/similar value with that found in the present study for the wild mussel of the North Sea (0.2834 particles g\(^{-1}\)w.w.). This could be attributed to mussel farms acting as a potential source of MPs contamination for the mussels due to plastic materials (i.e. plastic sock nets and polypropylene long lines) used during cultivation (Mathalon and Hill, 2014, Santana et al., 2018), probably highlights the small contribution of mussel farms as a source of MPs pollution (Santana et al., 2018). Moreover, the intertidal wild mussel (present study) is assumed to filter and excrete MPs half of the time in comparison with the submerged cultured mussel in the North Sea, resulting though in similar
accumulation level. The model also predicts the time needed for the 90% gut clearance of both cultured (N. Ionian Sea) and wild (North Sea) mussel to be almost 284330 hours and 5663 hours (equivalent to 124 and 2.53 days) respectively, when MPs contamination is removed from their habitat. This is in line with a series of studies which demonstrated that the depuration time varies between 6-72 hours and can last up to 40 days depending on several factors such as species, environmental conditions (Bayne et al., 1987), size and type of MPs (Browne et al., 2008, Ward and Kach, 2009, Woods et al., 2018, Birnstiel et al., 2019).

The strong dependence of food (CHL-a), temperature and seawater’s MPs concentration on the MPs accumulation by the mussel, regarding its wet weight, is demonstrated through sensitivity experiments that were used to derive a rather simple nonlinear regression model (Eq. 19). The comparison of the regression model’s with the DEB model’s output resulted in a quite accurate estimation of the coefficients, which in turn sparked the idea of a ‘new’ relationship (Eq. 20) that could potentially predict the MPs concentration in the environment ($C_{env}$) when certain conditions are known (CHL-a, T, $C_{env}$, W). The latter equation was applied in 8 areas in total (2 from the present study areas and 6 from Li et al. (2018)), with relatively good results since there is general overlapping of regressed and observed MPs concentration in the environment ($C_{env}$), except for Hastings-A and Plymouth areas, probably due to missing information on the environmental conditions (CHL-a, SST) during the sampling the predicted value is within the observed range of field data in most regions, suggesting that making the mussels can be used as potential bioindicators.

Mussels have been previously proposed as bioindicators for marine microplastic pollution (<1 mm), although the efficient gut clearance and selective feeding behavior limit their quantitative ability (Lusher et al., 2017, Brate et al., 2018, Beyer et al., 2017, Fossi et al., 2018, Li et al., 2019). The very recent study by Ward et al. (2019b) demonstrated that bivalves are poor bioindicators of MPs pollution due to the particle selection during feeding and excretion processes that is based on the physical characteristics of the MPs. Considering that the MPs accumulation is site-dependent and that sampling of mussels is usually easier than seawater (Karlsson et al., 2017, Brate et al., 2018), models like the one described in Eq. 20 that besides the MPs accumulation take into account also characteristics of the environment, that which are crucial for the way that mussels accumulate MPs. This method possibly could be possibly used at global level and allow comparisons between various environments. However, the method described should be validated in more environments with more frequent field data to be able to provide secure results.

In addition to Despite the scarce validation data regarding the MPs accumulation in mussels, in this study has some more there are some other limitations. First of all, the data regarding the concentration of MPs in the mussels’ environment are also scarce; since MPs is a relatively recent subject of study, the existing knowledge of the spatial and temporal distribution is still quite limited.
To overcome the lack of environmental MPs time series, a function of randomly generated values within the observed range of each area was applied and its uncertainty was examined through an ensemble forecasting. Specifically, the model’s uncertainty due to the environmental MPs concentration ($C_{env}$) was tested by performing a series of model runs forced by an envelope of representative values of $C_{env}$ and $T$. The results (section 3.3) showed that the adopted stochastic scenario simulated quite satisfactorily realistically the MPs accumulation by the mussels, lying within the observed field range, although a slight overestimation was found in the North Sea and in agreement with the available field data. The approach used is assumed to represent the natural variability be close to reality since it has been reported that MPs quantification in the water is rather a complicated procedure due to the influence of many factors such as tides, wind, wave action, ocean currents, river inputs and hydrodynamic features resulting to high spatially and temporally variability of MPs distribution even in very small scales (Messinetti et al., 2018, Goldstein et al., 2013). In addition, the nature of the variable $C_{env}$ makes it difficult to estimate, presenting large observational errors, not only due to the intense physical variation but also due to different sampling and analysis techniques that were used. In a future work the DEB-accumulation model could be coupled with a high-resolution MPs distribution model (Kalaroni et al., 2019), being extensively validated against field data that will have been collected and processed according to a common scientifically defined protocol, to overcome this limitation. Moreover, the approach followed in calculating the value of MPs concentration in the near surface layer (0-5m depth) (Kooi et al., 2016), resulted in a representative value of the upper ocean layer. In depth knowledge of the MPs distribution, both horizontally and vertically, is essential to understand and mitigate their impact not only on the various marine compartments but also on the organisms inhabiting those compartments (Van Sebille et al., 2015, Kooi et al., 2016). For that reason, it is important to enhance the monitoring activity especially in the vulnerable coastal environments, adopting integrated cross-disciplinary approaches and monitoring of biological, physical and chemical parameters which provide information on the ecosystem function, in order to improve the assessment of emerging pollutants (i.e. MPs) and their impacts on biota (objective of JERICO-RI framework).

Our assumption that the mussel has the same filtration rate for all particles independently of their chemical composition, size and shape, is a simplification and an open contradictory theme of discussion (see Saraiva et al., 2011a for details). However, in our model application through the model, a pre-ingestive particle selection by the mussel is implied based on the organic-inorganic content of the suspended matter illustrating the different binding probabilities applied for algal and MPs particles during the ingestion process. Through an investigation of wild
mussel’s faeces and pseudofaeces production in laboratory conditions, Zhao et al. (2018) found that the length of MPs was significantly longer in pseudofaeces than in the digestive gland and faeces. Furthermore, Van Cauwenberghe et al. (2015) demonstrated that mussel’s faeces contained larger MPs (15–500 μm) compared to the mussel’s tissue (20–90 μm). Apparently, smaller sized MPs seem to be dominant within the mussels in comparison with the size of the MPs in the ambient environment (Li et al., 2018, Qu et al., 2018, Digka et al., 2018b), implying that the mussel is more prone to ingest and retain smaller sized MPs. As an example, Digka et al. (2018b) confirmed that the smaller MPs (<1 mm) occupy the 62.3%, 96.9% and 100% of the total MPs in seawater, sediments and mussels from the N. Ionian Sea respectively. In a future work this selection pattern regarding size, could be simulated by suitable preference weights among different MPs sizes. This will improve the knowledge of the feeding and excretion mechanisms used by the mussels against MPs pollution and the assessment of the ecological footprint (Rist et al., 2019).

Our use of the tide data led to some model bias. Furthermore, the tide data as considered in the present study impose model's bias, since the model does not take into account of the mussel’s body temperature change when this is exposed to air was not taken into consideration. Assessing the mussel’s body temperature requires demand extended experiments in field conditions (Tagliarolo and McQuaid, 2015, Monaco and McQuaid, 2018).
along the French coast of the eastern English Channel found no significant correlation between air’s and mussel’s body temperature, but demonstrated a rather positively significant positive correlation between the body temperature and the hard substrate’s (i.e. rocks) temperature. However, in the present study the tide effect on processes that are affected by the thermal equation (k(T)) is considered indirectly through the metabolic depression (details in section 2.4). Sara et al. (2011) following the method developed by Kearney et al. (2010), who coupled a DEB model with a biophysical model (Kearney et al., 2010), incorporating the change of mussel’s body temperature during emersion, by using information of various climatological variables (i.e. solar radiation, air temperature, wind speed, wave height), but the ignored the temperature sensitivity on the physiological processes was ignored. In a future study, a combined similar approach by coupling the present DEB-accumulation model with a biophysical model, which includes both the tide effect on the physiological processes and the mussel’s body temperature respectively, could be followed and lead to a more detailed simulation of the intertidal mussel’s body temperature.

5. Conclusions

In a future study the model should be corroborated further by using a larger dataset of validated against more frequent field data regarding the MPs accumulation, with sampling of mussels of among various sizes and life stages. Currently, the model is mainly limited by the insuffient validation, as a larger dataset could be also used for a better model calibration, as for now it cannot be reliable in conducting predictions within accepted precision. However, this study provides a new approach in studying the accumulation of MPs by filter feeders and reveals the relations between characteristics of the mussel’s surrounding environment and the MPs accumulation, which is presented with high seasonal fluctuations. Additionally, in a future study the DEB-accumulation model will be coupled to with a coupled hydrodynamic-biochemical model (e.g., Petihakis et al., 2002, 2012, Triantafyllou et al., 2003, Tsiaras et al., 2014, Ciavatta et al., 2019, Kalaroni et al., 2020) and a MPs distribution model (Kalaroni et al., 2019) that will provide fields of temperature, food availability and MPs concentration respectively at the Mediterranean scale, and eventually lead to an integrated representation of the MPs accumulation by mussels (Daewel et al., 2008). This fully coupled model will be downscaled to the Cretan Sea SuperSite, while the parameterization of important biological processes will be redesigned based on the new data which will be acquired in the framework of the JERICO S3 project (http://www.jerico-ri.eu). The present study highlights the urgent need for adopting a multi-disciplinary monitoring activity by measuring physical, biological and chemical parameters that are crucial for mapping the MPs distribution, assessing the contamination level of the marine organisms and investigating the impact on the
health status. Overall, despite the significant limitations that were mentioned before, taken into account that plastics are one of the global hot issues, this particular study could help for the design of next efforts, since it provides indications on the future priority related issues.

Author Contribution:
Natalia Stamatakis, Yannis Hatzonikolakis, Kostas Tsiaras, Catherine Tsangaris, George Petihakis, Sarantis Sofianos, George Triantafyllou

G.T. conceived the basic idea of the present study and was responsible for the management and coordination of the research planning and execution. N.S. and Y.H. developed the model code with the contribution of K.T.. N.S. collected the existing information on the subject and performed the simulations of the present study with the help of Y.H. when needed. G.T., G.P., K.T., Y.H. and N.S. contributed to the interpretation of the results. C.T. provided the field data of the mussel’s microplastic accumulation in the North Ionian Sea. N.S. prepared the manuscript, with critical review, commentary and revision contributed from all co-authors.

Competing interests:
The authors declare that they have no conflict of interest.

Acknowledgments:
This work was partially funded by the national project ‘Blue growth with innovation and application in Greek Seas’ (MIS 5002438) and the EU H2020 CLAIM project (G.A. n° 774586). This study has been conducted using E.U. Copernicus Marine Service Information (http://marine.copernicus.eu/). Part of this work was supported by the JERICO- NEXT project. This project has received funding from the European Union’s Horizon 2020 research and innovation programme under grant agreement no. 654410.
<table>
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<th>References</th>
</tr>
</thead>
</table>


 GlobColour data (http://globcolour.info) used in this study has been developed, validated, and distributed by ACR-ST, France.


### Tables & Figures

\[ \frac{dE}{dt} = \dot{p}_A - \dot{p}_C \]  
\[ \frac{dV}{dt} = \frac{k \cdot \dot{p}_C - \dot{p}_M}{[E_g]} \]  
\[ \frac{dR}{dt} = (1 - k) \cdot \dot{p}_C - \left[ \frac{1 - k}{k} \right] \cdot \min(V, V_p) \cdot \dot{p}_M \]  
\[ \dot{p}_A = \{\dot{p}_{Am}\} \cdot f \cdot k(T) \cdot V^{\frac{2}{3}} \]  
\[ f = \frac{x}{x + K_y} \quad \text{where} \quad K_y = X_K \cdot (1 + \frac{y}{y_K}) \]  
\[ \dot{p}_C = \frac{[E]}{[E_g] + k \cdot [E]} \cdot \left( \frac{[E_g] \cdot \{ \dot{p}_{Am}\} \cdot k(T) \cdot V^{\frac{2}{3}}}{[E_m]} + \dot{p}_M \cdot V \right) \]  
\[ [E] = \frac{E}{V} \]  
\[ \{\dot{p}_M\} = k(T) \cdot \{\dot{p}_M\}_m \]  
\[ k(T) = \frac{\exp\left(\frac{r_A}{V_i} \cdot \frac{T_A}{T_i}\right)}{1 + \exp\left(\frac{r_A}{V_i} \cdot \frac{T_A}{T_i}\right) + \exp\left(\frac{r_A}{V_i} \cdot \frac{T_A}{T_i}\right)} \]  
\[ L = \frac{V^{\frac{1}{3}}}{\delta_m} \]  
\[ W = d \cdot \left( V + \frac{E}{[E_g]} \right) + \frac{R}{\mu E} \]  
\[ \dot{C}_R = \frac{[\dot{C}_{Rm}]}{1 + \sum_i^{\mathcal{X}} \left[ \frac{\dot{C}_{Rm}}{([P_{Xifm}] \cdot [P_{Xifm}])} \right] \cdot k(T) \cdot V^{\frac{2}{3}}} \cdot i = \begin{cases} 1 & \text{for CHL - a} \\ 2 & \text{for MPS} \end{cases} \]  
\[ \dot{p}_{Xif} = \dot{C}_R \cdot X_i \]  
\[ \dot{p}_{Xif} = \frac{p_{Xif} \cdot \dot{p}_{Xif}}{1 + \sum_i^{\mathcal{X}} \left[ \frac{p_{Xif} \cdot \dot{p}_{Xif}}{[P_{Xif}] + \sum_i^{\mathcal{X}} \left[ \frac{p_{Xif} \cdot \dot{p}_{Xif}}{[P_{Xif}] \cdot [P_{Xif}]} \right] \right]} \]  
\[ \dot{p}_{Xli} = \dot{p}_{Xli} - \dot{p}_{Xli} \]  
\[ \dot{f} = \dot{p}_{Xli} - \dot{p}_{A} \]
Table 1. Dynamic energy budget model: equations. See Table 2 for model variables, Table 3 for parameters and Table 4 for initial values.

\[
GSI = \frac{R}{d\left(V + \frac{E}{E_g}\right) + \frac{R}{\mu E}}
\]  

(17)

\text{Variable} & \text{Description} & \text{Units} \\
\hline
\text{V} & \text{Structural volume} & \text{cm}^3 \\
\text{E} & \text{Energy reserves} & \text{J} \\
\text{R} & \text{Energy allocated to development and reproduction} & \text{J} \\
\text{C} & \text{Microplastics accumulation} & \text{particles individual}^{-1} \\
\text{\dot{p}_a} & \text{Assimilation energy rate} & \text{J d}^{-1} \\
\text{\dot{p}_c} & \text{Utilization energy rate} & \text{J d}^{-1} \\
\text{\dot{C}_R} & \text{Clearance rate} & \text{m}^3\text{d}^{-1} \\
\text{C_{env}} & \text{Microplastics concentration} & \text{particles L}^{-1} \\
\text{\dot{p}_{XF}} & \text{Filtration rate} & \text{J d}^{-1} \text{ or g d}^{-1} \\
\text{\dot{p}_{Xi}} & \text{Ingestion rate} & \text{J d}^{-1} \text{ or g d}^{-1} \\
\text{\dot{f}_{pi}} & \text{Pseudofaeces production rate} & \text{J d}^{-1} \text{ or g d}^{-1} \\
\text{\dot{f}_f} & \text{Faeces production rate} & \text{J d}^{-1} \\
\text{f} & \text{Functional response function} & - \\
\text{\chi_i} & \text{Food or MPs density} & \text{mg chla m}^{-3} \text{ or g m}^{-3} \\
\text{[\dot{p}_M]} & \text{Maintenance costs} & \text{J cm}^{-3} \text{ d}^{-1} \\
\text{T} & \text{Temperature} & \text{K} \\
\text{k(T)} & \text{Temperature dependence} & - \\
\text{L} & \text{Shell length} & \text{cm} \\
\text{W} & \text{Fresh tissue mass} & \text{g} \\
\text{GSI} & \text{Gonado-somatic index} & - \\

Table 2. Dynamic energy budget model: variables
<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Description</th>
<th>Value</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>${\dot{p}_{Am}}$</td>
<td>J cm$^{-2}$ d$^{-1}$</td>
<td>Maximum surface area-specific assimilation rate</td>
<td>147.6</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>${C_{Rm}}$</td>
<td>m$^3$ cm$^{-2}$d$^{-1}$</td>
<td>Maximum surface area-specific clearance rate</td>
<td>0.096</td>
<td>Saraiva et al. (2011a)</td>
</tr>
<tr>
<td>${p_{X,Fm}}$</td>
<td>mg chl-a cm$^{-2}$d$^{-1}$</td>
<td>Algal maximum surface area-specific filtration rate*</td>
<td>0.1152</td>
<td>Rosland et al. (2009)</td>
</tr>
<tr>
<td>${\dot{p}_{X,Fm}}$</td>
<td>g cm$^{-2}$d$^{-1}$</td>
<td>Silt maximum surface area-specific filtration rate</td>
<td>3.5</td>
<td>Saraiva et al. (2011a)</td>
</tr>
<tr>
<td>${\dot{p}_{X,im}}$</td>
<td>mg chl-a d$^{-1}$</td>
<td>Algae maximum ingestion rate*</td>
<td>3.12·10$^6$</td>
<td>Saraiva et al. (2011b)</td>
</tr>
<tr>
<td>${\dot{p}_{X,im}}$</td>
<td>g d$^{-1}$</td>
<td>Silt maximum ingestion rate</td>
<td>0.11</td>
<td>Saraiva et al. (2011b)</td>
</tr>
<tr>
<td>$\rho_1$</td>
<td>-</td>
<td>Algae binding probability</td>
<td>0.99</td>
<td>Saraiva et al. (2011a)</td>
</tr>
<tr>
<td>$\rho_2$</td>
<td>-</td>
<td>Inorganic material binding probability</td>
<td>0.45</td>
<td>Saraiva et al. (2011a)</td>
</tr>
<tr>
<td>$k_f$</td>
<td>d$^{-1}$</td>
<td>Post-ingestive losses through faeces</td>
<td>Calibrated</td>
<td>-</td>
</tr>
<tr>
<td>$X_k$</td>
<td>mg chl m$^{-3}$</td>
<td>Half saturation coefficient</td>
<td>Calibrated</td>
<td>-</td>
</tr>
<tr>
<td>$T_A$</td>
<td>K</td>
<td>Arrhenius temperature</td>
<td>5800</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$T_i$</td>
<td>K</td>
<td>Reference temperature</td>
<td>293</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$T_L$</td>
<td>K</td>
<td>Lower boundary of tolerance rate</td>
<td>275</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$T_H$</td>
<td>K</td>
<td>Upper boundary of tolerance rate</td>
<td>296</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$T_{Al}$</td>
<td>K</td>
<td>Rate of decrease of upper boundary</td>
<td>45430</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$T_{AH}$</td>
<td>K</td>
<td>Rate of decrease of lower boundary</td>
<td>31376</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$[\dot{p}_{m}]_m$</td>
<td>J cm$^3$d$^{-1}$</td>
<td>Volume specific maintenance costs</td>
<td>24</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$[E_m]$</td>
<td>J cm$^3$</td>
<td>Maximum energy density</td>
<td>2190</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$k$</td>
<td>-</td>
<td>Fraction of utilized energy spent on maintenance/growth</td>
<td>0.7</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$V_p$</td>
<td>cm$^3$</td>
<td>Volume at start of reproductive stage</td>
<td>0.06</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>GSI$_{in}$</td>
<td>-</td>
<td>Gonado-somatic index triggering spawning</td>
<td>0.28</td>
<td>Van der Veer et al. (2006)</td>
</tr>
<tr>
<td>$\delta_m$</td>
<td>-</td>
<td>Shape coefficient</td>
<td>0.25</td>
<td>Casas &amp; Bacher (2006)</td>
</tr>
<tr>
<td>$d$</td>
<td>g cm$^{-3}$</td>
<td>Specific density</td>
<td>1.0</td>
<td>Kooijman (2000)</td>
</tr>
<tr>
<td>$\mu_E$</td>
<td>J g$^{-1}$</td>
<td>Energy content of reserves</td>
<td>6750</td>
<td>Casas &amp; Bacher (2006)</td>
</tr>
<tr>
<td>$\lambda$</td>
<td>J mg chl-a$^{-1}$</td>
<td>Conversion factor</td>
<td>2387.73</td>
<td>Rosland et al. (2009)</td>
</tr>
</tbody>
</table>

Table 3. Dynamic energy budget model: parameters
units mol C converted to mg CHL-a by multiplying with the factor \( \frac{12 \times 10^5}{50} \) assuming Carbon:CHL-a ratio of 50 (Hatzonikolakis et al., 2017).

<table>
<thead>
<tr>
<th>Area</th>
<th>Xₖ value (mg m⁻³)</th>
<th>CHL-a range (mg m⁻³)</th>
<th>CHL-a mean (mg m⁻³)</th>
<th>Temperature range(°C)</th>
<th>Length after one year±SD (cm)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maliakos Gulf</td>
<td>0.72</td>
<td>0.87-5.59</td>
<td>1.80</td>
<td>12.0-26.0</td>
<td>7.06 ± 0.46</td>
<td>Hatzonikolakis et al., 2017</td>
</tr>
<tr>
<td>Thermaikos Gulf</td>
<td>0.56</td>
<td>1.04-2.76</td>
<td>1.89</td>
<td>11.5-24.5</td>
<td>7.0 ± 0.47</td>
<td>Hatzonikolakis et al., 2017</td>
</tr>
<tr>
<td>Black Sea</td>
<td>Calibrated: 0.96</td>
<td>0.53-16.30</td>
<td>3.07</td>
<td>6.5-25.0</td>
<td>7.5 ± 0.1</td>
<td>Karayucel et al., 2010</td>
</tr>
<tr>
<td>Bizerte lagoon</td>
<td>3.829</td>
<td>4.00-7.70</td>
<td>5.20</td>
<td>12.0-28.0</td>
<td>7.26 ± 0.46</td>
<td>Béjaoui-Omri et al., 2014</td>
</tr>
</tbody>
</table>

Table 4. Half saturation tuned values (Xₖ) and mussel growth data (Length) in different areas of the Mediterranean and Black Seas.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Value</th>
<th>Variable</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Northern Ionian Sea</td>
<td></td>
<td>North Sea</td>
<td></td>
</tr>
<tr>
<td>Start date</td>
<td>20 Nov 2010</td>
<td>Start date</td>
<td>1 Jul 2007</td>
</tr>
<tr>
<td>L</td>
<td>0.85 cm</td>
<td>L</td>
<td>0.15 cm</td>
</tr>
<tr>
<td>W</td>
<td>0.1938 g</td>
<td>W</td>
<td>0.0055 g</td>
</tr>
<tr>
<td>V</td>
<td>0.0096 cm³</td>
<td>V</td>
<td>5.3×10⁻⁵ cm³</td>
</tr>
<tr>
<td>E</td>
<td>350 J</td>
<td>E</td>
<td>10 J</td>
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<td>R</td>
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<td>C</td>
<td>0 particles individual⁻¹</td>
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Table 5. Dynamic energy budget-accumulation model: initial values. L: shell length; W: fresh tissue mass; V: structural volume; E: energy reserves; R: energy allocated to reproduction; C: Microplastics accumulation
Fig. 1. Environmental data used for the forcing of the Dynamic Energy Budget model (DEB) in the North Sea simulation, showing temperature (top) and chlorophyll a concentration against in situ data from the ICES database (bottom).

Fig. 2. Environmental data used for the forcing of the dynamic energy budget model in the Northern Ionian Sea simulation, showing temperature (top) and chlorophyll a concentration (bottom).
Fig. 3. Simulated mussel shell length (L) (top) and fresh tissue mass (W) (bottom) against North Sea data (red star: mean ± SD), using chlorophyll a (X = [CHL-a]) in the mussel diet.

Fig. 4. Simulated mussel shell length (L) (top) and fresh tissue mass (W) (bottom) against North Sea data (red star: mean ± SD), using chlorophyll a (X = [CHL-a]) in the mussel diet.
Fig. 5A. Target diagram of simulated shell length ($L$) and fresh mass tissue weight ($W$) against field data from Thermaikos and Maliakos Gulf (eastern Mediterranean Sea), Black Sea and Bizerte Lagoon (southwestern Mediterranean Sea), using the power ($L_1, W_1$), exponential ($L_2, W_2$) and linear ($L_3, W_3$) function of the half saturation coefficient. The model bias is indicated on the y-axis while the unbiased root-mean-square-deviation (RMSD) is indicated on the x-axis.
Fig. 6. Microplastics (MPs) accumulation by the mussel (blue line) against field data (red star: mean ± SD), using daily environmental concentration of MPs ($C_{\text{env}}$ mean value ± SD: 0.4 ± 0.3 particles L$^{-1}$) in the North Sea.

Fig. 7. Microplastics (MPs) accumulation by the mussel (blue line) against field data (red star: mean value ± SD), using daily environmental concentration of MPs ($C_{\text{env}}$ mean value ± SD: 0.0012 ± 0.024 particles L$^{-1}$) in the Ionian Sea.

Fig. 8. Depuration phase of the cultured Mytilus galloprovincialis (red line) and wild Mytilus edulis (black line) using zero environmental concentration of microplastics ($C_{\text{env}}$=0) after 1 year and 4 years of simulation time at the Northern Ionian Sea and North Sea respectively.
Fig. 9. Mean seasonally values and standard deviation of microplastics (MPs) accumulation (red error bars: mean value ± SD) by the mussel in North Sea derived from 15 model runs with different constant values of environmental MPs concentration ($C_{env}$ range: 0.1-0.8 particles L$^{-1}$); Mean hourly simulated data (black line) and standard deviation (blue lines) of microplastics accumulation derived from 3 model runs with stochastic sequences of daily random $C_{env}$ values.

Fig. 10. Mean monthly values and standard deviation of microplastics accumulation (red error bars: mean value ± SD) by the mussel in Northern Ionian Sea derived from 13 model runs with different constant values of environmental MPs concentration ($C_{env}$ range: 0.0012-0.024 particles L$^{-1}$); Mean hourly simulated data (black line) and standard deviation (blue lines) of microplastics accumulation derived from 3 model runs with stochastic sequences of daily random $C_{env}$ values.
(blue lines) of microplastics accumulation derived from 3 model runs with stochastic sequences of daily random $C_{env}$ values.

Fig. 11. Sensitivity index of MPs accumulation on the wild mussel of the North Sea when variables (CHL-a, temperature, $C_{env}$) and parameters ($X_k$, $Y_k$) are perturbed ± 10%. The notation (s) refers to the permanently submerged mussel.

Fig. 12. Sensitivity index of MPs accumulation on the cultured mussel of the Northern Ionian Sea when variables (CHL-a, temperature, $C_{env}$) and parameters ($X_k$, $Y_k$) are perturbed ± 10%.
Fig. 13. Prediction of seawater microplastics concentration by using Eq. 20 for the Northern Ionian Sea, North Sea (present study) and 6 areas around U.K. (Filey, Hastings-A&B, Brighton, Plymouth, Wallasey; Li et al. (2018)).
Modelling mussel (*Mytilus spp.*) microplastic accumulation.

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Abstract: Microplastics (MPs) are a contaminant of growing concern due to their widespread distribution and interactions with marine species, such as filter feeders. To investigate the MPs accumulation in wild and cultured mussels, a Dynamic Energy Budget (DEB) model was developed and validated with the available field data of *Mytilus edulis* (*M. edulis*, wild) from the North Sea and *Mytilus galloprovincialis* (*M. galloprovincialis*, cultured) from the Northern Ionian Sea. Towards a generic DEB model, the site-specific model parameter, half saturation coefficient (*X*\textsubscript{k}) was applied as a power function of food density for the cultured mussel, while for the wild mussel it was calibrated to a constant value. The DEB-accumulation model simulated the uptake and excretion rate of MPs, taking into account of environmental characteristics (temperature and chlorophyll-a). An accumulation of MPs equal to 0.5364 particles individual\textsuperscript{-1} (fresh tissue mass 1.9 g) and 0.91 particles individual\textsuperscript{-1} (fresh tissue mass 3.34 g) was simulated for the wild and cultured mussel after 4 years and 1 year respectively, in agreement with the field data. The inverse experiments investigating the depuration time of the wild and cultured mussel in a clean from MPs environment showed a 90\% removal of MPs load after 2.53 and 1.24 days, respectively. Furthermore, sensitivity tests on model parameters and forcing functions highlighted that besides MPs concentration, the accumulation is highly depended on temperature and chlorophyll-a of the surrounding environment. For this reason, an empirical equation was found, directly relating directly the environmental concentration of MPs in seawater, with the seawater’s temperature, chlorophyll-a and the MPs accumulation in mussel’s soft tissue, MPs load, temperature and chlorophyll-a.
1. Introduction

Microplastic particles (MPs) are synthetic organic polymers with size below 5 mm (Arthur et al., 2009) that originate from a variety of sources including—mainly—those particles that are manufactured for particular household or industrial activities, such as facial scrubs, toothpastes and resin pellets used in the plastic industry (primary MPs), and those formed from the fragmentation of larger plastic items (secondary MPs) (GESAMP, 2015). Eriksen et al. (2014) estimated that more than 5 trillion microplastic particles, weighing over 250,000 tons, float in the oceans. Due to their composition, density and shape, MPs are highly persistent in the environment and are, therefore, accumulating in different marine compartments at increasing rates: surface and deeper layers in the water column, as well as at the seafloor and within the sediments (Moore et al., 2001, Lattin et al., 2004, Thompson, 2004, Lusher, 2015). Since the majority of MPs entering the marine environment, originate from the land (i.e. land-fills, littering of beaches and coastal areas, rivers, floodwaters, untreated municipal sewerage, industrial emissions), the threat of MPs pollution in the coastal zone puts considerable pressure on the coastal ecosystems (Cole et al., 2011, Andrady, 2011). In recent years, initiatives under various projects (i.e. CLAIM, DeFishGear) target at evaluating the threat and impact of marine litter pollution; the European framework of JERICO-RI focuses on a sustainable research infrastructure in the coastal area to support the monitoring, science and management of coastal marine areas (http://www.jerico-ri.eu/). In the framework of JERICO-NEXT, a recent study addressed the environmental threats and gaps within monitoring programmes in European coastal waters, including the marine litter (i.e. MPs) as one of the most commonly identified threat to the marine environment and highlighted the need for improved monitoring of the MPs distribution and their impacts in European coastal environments (Painting et al., 2019).

Numerous studies have revealed that MPs are ingested either directly or through lower trophic prey by animals atfrom all levels of the food web; from zooplankton (Cole et al., 2013), small pelagic fishes and mussels (Digka et al., 2018a) to mesopelagic fishes (Wieczorek et al., 2018) and large predators like tuna and swordfish (Romeo et al., 2015). Microplastic ingestion by marine animals can potentially affect animal health and raises toxicity concerns, since plastics can facilitate the transfer of chemical additives and/or hydrophobic organic contaminants to biota (Mato et al., 2001, Rios et al., 2007, Teuten et al., 2007, 2009, Hirai et al., 2011). Human, as a top predator, is also contaminated by MPs (Schwabl et al., 2019). Mussel and small fishes that are commonly consumed whole, without removing digestive tracts, where MPs are concentrated, are among the most likely pathways for MPs to embed in the human diet (Smith et al., 2018). Especially regarding marine organisms (i.e. mussels), it is notable that the levels of their contamination has been added
to the European database (www.ecsafeseafooddbase.eu) as an environmental variable of growing concern, reflecting the health status (Marine Strategy Framework Directive (MSFD) Descriptor 10 – Marine Litter (Decision 2017/848/EU)) (De Witte et al., 2014, Vandermeersch et al., 2015, Digka et al., 2018a). Today, a series of studies have denoted the presence of MPs in mussels’ tissue intended for human consumption (Van Cauwenbergh and Janssen, 2014, Mathalon and Hill, 2014, Li et al., 2016, 2018, Hantoro et al., 2019). For instance, in a recent study, Li et al. (2018) sampled mussels from coastal waters and supermarkets in the U.K and estimated that a plate of 100g mussels contains 70 MPs that will be ingested by the consumer. The presence of MPs in mussels has been also demonstrated during laboratory trials in their faeces, intestinal tract (Von Moos et al., 2012, Van Cauwenbergh et al., 2015, Wegner et al., 2012, Khan and Prezant, 2018), as well as in their circulatory system (Browne et al., 2008). Other laboratory studies showed several effects of microplastic ingestion in laboratory exposed mussels, including histological changes, inflammatory responses, immunological alterations, lysosomal membrane destabilization, reduced filtering activity, neurotoxic effects, oxidative stress effects, increase in hemocyte mortality, dysplasia, genotoxicity and transcriptional responses (reviewed by Li et al., 2019). However, the tested concentrations of MPs in laboratory experiments are frequently unrealistic, being several orders of magnitude higher (2 to 7 orders of magnitude) than the observed seawater concentrations (Van Cauwenberge et al., 2015, Lenz et al. 2016).

Mussels, through their extensive filtering activity, feed on planktonic organisms that have similar size with MPs (Browne et al., 2007) and considering also their inability to select particles with high energy value (i.e. phytoplankton) during filtration (Vahl, 1972, Saraiva et al., 2011a), they are directly exposed to MPs’ contamination. Recent studies suggest a positive linear correlation between MPs concentration in mussels and surrounding waters (Capolupo et al., 2018, Qu et al. 2018, Li et al. 2019). The filtering activity of mussels, which directly affects the resulting MPs accumulation, is a complicated process that is controlled by other factors (food availability, temperature, tides etc.).

The purpose of the present work is to study the accumulation of MPs in mussels and reveal relations between the accumulated concentrations in mussels’ soft parts and environmental features. In this context, an accumulation model was developed based on Dynamic Energy Budget theory (DEB, Kooijman, 2000) and applied in two different regions, in two different modes of life (wild and cultivated): in the North Sea (Mytilus- Edulis (M. edulis), wild) and in the Northern Ionian Sea (Mytilus- Galloprovincialis (M. galloprovincialis), cultivated). DEB theory provides all the necessary detail to model the feeding processes and aspects of the mussel metabolism, taking into account the impact of the environmental variability on the simulated individual. Apart from modelling the growth of bivalves (Rosland et al., 2009, Sara et al., 2012, Thomas et al., 2011,
Saraiva et al., 2012, Hatzonikolakis et al., 2017, Monaco & McQuaid, 2018), DEB models have been used to study other processes as well, such as bioaccumulation of PCBs (Polychlorinated Biphenyls) and POPs (Persistent Organic Compounds) (Zaldivar, 2008), trace metals (Casas and Bacher, 2006) and the impact of climate change on individual’s physiology (Sara et al., 2014). However, to our knowledge this is the first time that a DEB-based model is used to assess the uptake and excretion rates of MPs in mussels.

2. Materials and Methods

2.1 Study areas and field data

The North Sea is a marginal large semi-enclosed sea on the continental shelf of north-west Europe with a total surface area of 850,000 km² and is bounded by the coastlines of 9 countries. The sea is shallow (mean depth 90 m), getting deeper towards the north (up to 725 meters) and the semi-diurnal tide (tidal range 0-5 m) is the dominant feature of the region (Otto et al., 1990). Major rivers, such as Rhine, Elbe, Weser, Ems and Thames discharge into the southern part of the sea (Lacroix et al., 2004), making this area a productive ecosystem. In this study, the area is limited along the French, Belgian and Dutch North Sea coast (N 50.98°-51.46°, W 1.75°-3.54°). This is located close to harbors, where shipping, industrial and agricultural activity is high, putting considerable pressure on the ecological systems of the region (Van Cauwenberghe et al., 2015).

The MPs concentration in mussels’ tissue and seawater that were used to validate and force the model respectively at its North Sea implementation were derived from Van Cauwenberghe et al. (2015). Van Cauwenberghe et al. (2015) examined the presence of MPs in wild mussels (M. edulis), and thus collected both biota and water at 6 sampling stations along the French, Belgian and Dutch North Sea coast in late summer of 2011. M. edulis (mean shell length: 4 ± 0.5 cm and wet weight (w.w.): 2 ± 0.7 g) and water samples were randomly collected on the local breakwaters, in order to assess the MPs concentration in the organisms and their habitat. MPs were present in all analyzed samples, both organisms and water. Seawater samples (N=12) had MPs (<1mm) on average 0.4 ± 0.3 particles L⁻¹ (range: 0.0 – 0.8 particles L⁻¹) and M. edulis contained on average 0.2 ± 0.3 particles g⁻¹w.w. (or 0.4 ± 0.3 particles individual⁻¹) (Van Cauwenberghe et al., 2015). The size range of MPs found within the mussels was 20-90 μm (size <1 mm).
The Northern Ionian Sea is located in the transition zone between the Adriatic and Ionian Sea. The long and complex coastline, presents a high diversity of hydrodynamic and sedimentary features. Rivers discharging into the Northern Ionian Sea include Kalamas/Thyamis (Greece) and Butrinto (Albania) (Skoulikidis et al., 2009), making the area suitable for aquaculture. Small farming sites and shellfish grounds are operating in Thesprotia (northwestern Ionian Sea) (Theodorou et al., 2011). The main source of marine litter inputs in the area originates from anthropogenic activities that mainly include shoreline tourism and recreational activities, poor wastewater management, agricultural practices, fisheries, aquacultures and shipping (Vlachogianni et al., 2017; Digka et al., 2018a). According to Politikos et al. (2020), the area around the Corfu island (Northern Ionian Sea) is characterized as a retention area of litter particles probably due to the prevailing weak coastal circulation. Furthermore, a northward current on the east Ionian Sea facilitates the transfer of litter particles towards the Adriatic Sea, which has been characterized as a hotspot of marine litter and one of the most affected areas in the Mediterranean Sea (Pasquini et al., 2016, Vlachogianni et al., 2017, Liubartseva et al., 2018, Politikos et al., 2020).

The field data used to validate the model output in the N. Ionian Sea were obtained from Digka et al. (2018b, 2018a). In the framework of the “DeFishGear” project, mussels (M. galloprovincialis) were collected by hand from a long line type mussel culture farm in Thesprotia (N 39.606567° E 20.149421°), in summer 2015 (end of June-July) at a sampling depth up to 3 m (Digka et al., 2018a). The average MPs accumulation was calculated from a total population of 40 mussels originated from the farm, with 18 of them found contaminated with MPs (46.25%). The average load of MPs (size <1 mm) per mussel (mean shell length 5.0 ± 0.3 cm) was 0.9 ± 0.2 particles individual⁻¹ and the size of MPs found in the mussel’s tissue ranged from 55 to 620 μm. Both clean and contaminated mussels were included in the calculated mean value in order to represent the mean state of the contamination level for the individual inhabiting the study area.

The seawater concentration of MPs for the N. Ionian Sea implementation was obtained from Digka et al. (2018b) and the DeFishGear project results (http://www.defishgear.net/project/main-lines-of-activities). In total, 12 manta net tows were conducted in the region, collecting a total number of n1=2,027 particles on October 2014 and n2=1,332 on April 2015, leading to an average of 280 particles per tow with size <1 mm and >330 μm (Digka et al., 2018b). In order to estimate the mean MPs concentration in the region, expressed as particles per volume, the dimensions of the manta net (W 60 cm H 24 cm, rectangular frame opening, mesh size 330 μm) and the sampling distance of each tow (~2 km) were used by multiplying the sample surface of the net by the trawled distance in meters (Maes et al., 2017), which resulted in a mean MPs concentration of 1.17 particles m⁻³ (233,333 particles km⁻²). Moreover, in the wider region of the Adriatic Sea, Zeri et al. (2018) found a mean density of 315,009 ± 568,578 particles km⁻² (1.58 ± 2.84 particles m⁻³), out of which
A relatively high value of standard deviation (one order of magnitude higher than the mean value) is adopted (0.0012 ± 0.024 particles L\(^{-1}\)), considering that the mussel farm is established in an enclosed gulf and close to the coast, since, according to Zeri et al. (2018), the abundance of MPs is one order of magnitude higher in inshore (<4 km) compared to offshore waters (>4 km). Furthermore, it may be assumed that the adopted range (standard deviation is also multiplied by a factor of 2) includes also the smaller particles sized between 50 μm and < 330 μm, which have been found in mussel’s tissue (Digka et al., 2018a), but were overlooked during the seawater sampling due to the manta net’s mesh size (> 330 μm). According to Enders et al. (2015) the relative abundance of small particles (50-300 μm) compared to particles larger than 300 μm is approximately 50%.

### 2.2 DEB model description

In the present study, a Dynamic Energy Budget (DEB, Kooijman, 2000, 2010) model is used as basis to simulate the accumulation of MPs by mussels. In DEB theory (Kooijman, 2000), the energy assimilated through food by the simulated individual is stored in a reserve compartment from where a fixed energy fraction \(\kappa\) is allocated for growth and somatic maintenance, with a priority for maintenance. The remaining energy \((1 - \kappa)\) is spent on maturity maintenance and reproduction. The individual’s condition is defined by the dynamics of three state variables: energy reserves \(E\) (joules), structural volume \(V\) (cm\(^3\)) and energy allocated to reproduction \(R\) (joules). The energy flow through the organism is controlled by the fluctuations of the available food density and temperature characterizing the surrounding environment.

The DEB model implemented here is an extended version of the model described in Hatzonikolakis et al. (2017), where the growth of the Mediterranean mussel is simulated by taking into account only the assimilation rate of the individual. Since the present study focuses on simulating the MPs accumulation, it is crucial to include a detailed representation of the mussel’s feeding mechanism. In this context, the DEB model was extended by including the clearance \((C_r)\), filtration \((\dot{p}_{XIF})\) and ingestion \((\dot{p}_{XII})\) rates of the mussel, following Saraiva et al. (2011a), assuming that all parameters referred to silt (or inedible particles) are applicable also to MPs particles represented by the silt variable. In this approach, a pre-ingestive selection occurs between filtration and ingestion, returning the rejected material in the water through pseudofaeces \((J_{pf})\). Consequently, energy is assimilated through food while the non-assimilated particles are excreted through the faeces production \((J_f)\). The model’s equations, variables and parameters are shown in Table 1, 2 and 3 respectively. The scaled functional response \(f\) (Eq. 5, Table 1), which regulates the...
assimilation rate, is modified following Kooijman (2006) to include an inorganic term representing the non-digestible matter i.e. microplastics: \( f = \frac{X}{(X + K_y)} \) and \( K_y = X_K \cdot (1 + Y/Y_K) \) where \( Y \) and \( Y_K \) are the concentration of MPs, converted from particles \( \text{L}^{-1} \) to \( \text{g m}^{-3} \) (Everaert et al., 2018) and the half saturation coefficient of inorganic particles here represented by MPs (\( \text{g m}^{-3} \)), respectively. Thus, the assimilation rate that is regulated by \( f \) is decreasing when the concentration of MPs is increased. The same approach is followed by other authors who considered inedible particles in the mussel’s diet (Ren, 2009, Troost et al., 2010). During the filtration process the same clearance rate for all particles is used (\( \hat{C}_r \)), representing the same searching rate for food that depends on the organism maximum capacity (\( \hat{C}_{rm} \)) and environmental particle concentrations (Vahl, 1972, Widdows et al., 1979, Cucci et al., 1989). During the ingestion process the mussel is able to selectively ingest food particles and reject inedible material, in order to increase the organic content of the ingested material ((Kiørboe & Møhlenberg, 1981, Jørgensen et al., 1990, Prins et al., 1991, Maire et al., 2007, Ren, 2009, Saraiva et al., 2011a). This selection is reflected by the different binding probabilities adopted for each type of particle (\( \rho_I \) for algae particles and \( \rho_2 \) for inorganic particles i.e. MPs, see Eq. 14 and table 3). The equations representing the feeding processes handle each type of particle separately, while there is interference between the simultaneous handling of different particle types (Eq. 12-14, Table 1) (Saraiva et al., 2011a). Finally, during the assimilation process, suspended matter (i.e. MPs) that the mussel is not able to assimilate due to its different chemical composition from the reserve compartment (Saraiva et al., 2011a) or incipient saturation at high algal concentrations (Riisgard et al., 2011) results in the faeces production (Eq. 16, Table 1).

### 2.3 Microplastics accumulation sub-model

With the DEB model as a basis, a sub-model describing the microplastics (MPs) accumulation by the mussel was developed, assuming that the presence of MPs in the ambient water does not cause a significant adverse effect on the organisms’ overall energy budget, in accordance with laboratory experiments, conducted in mussel species (Van Cauwenbergh et al., 2015: Mytilus edulis, Santana et al., 2018: mussel Perna perna). Additionally, it was assumed that the mussel filtrates MPs present in the water, without the ability of selecting between the high energetic valued particles and the MPs during the filtration process (Van Cauwenbergh et al., 2015, Von Moos et al., 2012, Browne et al., 2008, Digka et al., 2018a among others). The uptake of MPs from the environment is taken into account through the process of clearance/filtration rate, while the excretion of the contaminant is derived from two processes: (i) pseudofaeces production and (ii) faeces production. The resulting MPs accumulation is influenced by external environmental factors.
(MPs concentration, food availability, temperature) and internal biological processes (clearance, filtration, ingestion, growth). All these are described by the following differential equation describes the change of the individual MPs accumulation \( C \) (particles individual\(^{-1} \)), taking into account the processes mentioned above:

\[
\frac{dC}{dt} = C_{\text{env}} \cdot \dot{C}_R - \dot{j}_{pf2} - k_f \cdot \frac{j_f}{p_x11} \cdot C \\
\text{(Eq. 18)}
\]

where \( \dot{C}_R \) is the clearance rate for water (L h\(^{-1} \)), containing a concentration of MPs \( C_{\text{env}} \) (particles L\(^{-1} \)). The terms of \( \dot{j}_{pf2} \) and \( \frac{j_f}{p_x11} \) represent the elimination rate of MPs through pseudofaeces (particles h\(^{-1} \)) and the non-dimensional rate of faeces production with respect to the ingestion rate, respectively (see Table 1, Eq. 15-16). The parameter \( k_f \) represents the post-ingestive selection mechanism utilized by the mussel to incorporate indigestible material (i.e. MPs) into faeces and was calibrated using the available field data of mussel’s MPs accumulation from both study areas (Table 3). Mussel is able to discriminate among particles in the gut based on size, density and chemical properties of the particles (i.e. between microalgae and inorganic material) and thus to eliminate them through faeces (Ward et al., 2019a and references therein). In this context, the pseudofaeces production incorporates the rejected MPs prior to the ingestion, while the faeces production includes MPs that are rejected along with the food particles that are not assimilated by the mussel. The model’s time step has been set to one hour in order to capture the dynamics of the rapidly changing processes, such as feeding and excretion.

The accumulation of MPs in the individual is represented by the state variable \( C \) (particles individual\(^{-1} \)) which is computed at every model time step. This has been set to one hour, in order to properly resolve the dynamics of the rapidly changing processes, such as feeding and excretion.

### 2.4 Environmental drivers

Besides MPs concentration in the seawater, the DEB model is forced by sea surface temperature (SST) and food availability, represented defined by chlorophyll-a concentrations (CHL-a, an index of phytoplankton biomass). *M. edulis* has been demonstrated to filter suspended particles greater than 1μm, a size class that includes all of the phytoplankton, zooplankton and much of the detritus (Vahl, 1972, Mohlenberg and Riisgård, 1978, Saraiva et al., 2011a, Strohmeier et al., 2012), including even aggregated picoplankton-size particles (i.e. marine snow) (Kach and Ward, 2008, Ward and Kach, 2009). CHL-a has been considered the most reliable food quantifier for the calculation of DEB shellfish parameters (Pouvreau et al., 2006, Sara et al., 2012,
Hatzonikolakis et al. (2017 and references therein). Hatzonikolakis et al. (2017) have tested the performance of the model, considering also particulate organic carbon (POC) in the mussel’s diet, which, however, did not have an important impact on the model’s skill against field data in the Mediterranean Sea study areas. This outcome agrees with Troost et al. (2010) demonstration that POC contributes to the mussel’s diet when CHL-a concentrations are low at the southwest of Netherlands. Thus, in the present study, only CHL-a is considered as the available food source for mussels originated from the Southern North Sea and the Northern Ionian Sea. Thus, only CHL-a is considered as the available food source. For both study areas SST and CHL-a are derived from daily satellite data, a method also used by other authors (i.e. Thomas et al., 2011, Monaco & McQuaid, 2018).

In the North Sea, SST data were obtained from daily satellite images provided by Copernicus Marine Environmental Monitoring Service (CMEMS) at 0.04 degree spatial resolution and CHL-a data obtained from the Globcolour daily multi-sensor product provided by CMEMS–Globcolour database at 1 km spatial resolution, based on the OC5 algorithm of Gohin et al. (2002) (http://marine.copernicus.eu/, generated using CMEMS Products, production center ACRI-ST). The environmental forcing data (SST, CHL-a) were averaged over the study area (51.08°-51.44° N, 2.19°-3.45° E), covering the period 2007-2011 (5 years), in order to realistically simulate the wild mussel’s growth harvested in late summer 2011 (Van Cauwenbergh et al., 2015). It is notable that the study area of the North Sea belongs to CASE II waters (coastal region), where algorithms tend to overestimate CHL-a concentrations. In optically-complex Case II waters, CHL-a cannot readily be distinguished from particulate matter and/or yellow substances (dissolved organic matter) and so global chlorophyll algorithms are less reliable (IOCCG, 2000). However, the CHL-a dataset that was used was found in good agreement with available in situ data from ICES database (https://www.ices.dk/data/Pages/default.aspx) for the specific study area and time period, showing a relatively smaller bias and better time-space coverage, as compared with other tested remote sensing datasets (not shown) (i.e. regional chlorophyll product available for the North West Shelf-Seas in the CMEMS catalogue, https://resources.marine.copernicus.eu/).

In the North Ionian Sea, daily satellite SST data were also obtained from the CMEMS database for the Mediterranean Sea with 0.04 degree spatial resolution, while CHL-a daily data were derived from the Globcolour multi-sensor (i.e. SeaWiFs, MERIS, MODIS, VIIRS and OLCI-a) merged product (http://globcolour.info) at 1 km spatial resolution based on the OC5 algorithm suitable for coastal regions (Gohin et al., 2002) while CHL-a data were derived from the merged product of many satellites (i.e. SeaWiFs, Meris, Modis, Viirs and Oci-a) provided by Globcolour web interface (http://globcolour.info) at a daily temporal resolution and 1 km spatial resolution. The forcing data were averaged over the study area (N 39.49°-39.65°, E 20.09°-20.23°) covering the
period 2014-2015 (2 years), when the cultured mussel is ready for the market. The chosen CHL-a dataset was found preferable, as compared with other available remote sensing datasets (i.e. CMEMS chlorophyll product for Mediterranean Sea), since it presented a better spatial and temporal coverage (Hourany et al., 2019, Garnesson et al., 2019) and a slightly lower error, as compared with the very few available in situ data in the study area (not shown). Unfortunately, these were very scarce and therefore an extended comparison between remote and in situ data could not be conducted. The satellite-derived CHL-a data were estimated based on the OC5 algorithm of Gohin et al. (2002) in both study areas, which is regarded as suitable for coastal waters. Satellite data have facilitated large scale ecological studies by providing maps of phytoplankton functional types and sea surface temperature (Raitos et al., 2005, 2008, 2012, 2014, Palacz et al., 2013, Di Cicco et al., 2017, Brewin et al., 2017). The daily environmental forcing data are shown in Fig. 1 and Fig. 2 for the North Sea and the N. Ionian Sea, respectively. The two coastal environments present some important differences regarding both CHL-a and SST. Specifically, in the N. Ionian Sea, CHL-a is relatively low (annual mean ~ 0.88 mg chl-a m⁻³) and peaks during winter (maximum ~2.64 mg chl-a m⁻³ at December 2014), while in the North Sea CHL-a is about four times higher (annual mean 4.25 mg chl-a m⁻³), peaking in April every year (maximum range 29.44-33.38 mg chl-a m⁻³), as soon as light availability reaches a critical level (Van Beusekom et al., 2009). The higher productivity during spring season in the North Sea is related with the nutrient inputs from the English Channel, the North Atlantic and particularly the river discharge of nutrient-rich waters along the Belgian-French-Dutch coastline, which peaks earlier, during winter period (Van Beusekom et al., 2009). The SST sea surface temperature peaks during August in both areas (Fig. 1 and Fig. 2), but is significantly higher in the N. Ionian Sea (maximum 28.8°C), as compared to the North Sea (maximum 18-19.3°C).

The environmental concentration of MPs, $C_{env}$ (particles L⁻¹) was obtained also at a daily time step as randomly generated values of the Gaussian distribution that is determined by the mean value and standard deviation of the observed field data (0.4 ± 0.3 particles L⁻¹, North Sea, Van Cauwenberghhe et al., 2015, 0.0012 ± 0.024 particles L⁻¹, N. Ionian Sea, Digka et al., 2018a). Considering that these values originate from surface waters and that mussels live in the near surface layer (0-5 m), $C_{env}$ is estimated as a mean value of the upper layer with the methods described by Kooi et al. (2016), who studied the vertical distribution of MPs, considering an exponential decrease with depth. Specifically, in the N. Ionian Sea, mussels were collected from a depth up to 3 m (Digka et al., 2018a), while in the North Sea (Van Cauwenberghhe et al., 2015), there is no information and thus a maximum depth of 5 m is adopted.

In the North Sea simulation, the effect of tides is taken into account by considering that the mussel originated from the intertidal zone, is submerged 12 hours during the day (Van
Cauwenberghe et al., 2015). In the N. Ionian Sea simulation, tides are not considered, given the very small tide amplitude (few centimeters) in the Mediterranean (i.e. Sara et al., 2011; Hatzonikolakis et al., 2017) and thus the cultured mussel is assumed permanently submerged. *In situ* hourly tide data (2007-2011) from the coastal zone of the region (Dunkerque station N 51.04820°, E 2.36650°) obtained from Coriolis and Copernicus data provider (http://marine.copernicus.eu, http://www.coriolis.eu.org), showed that mussels experience alternating periods of aerial exposure and submergence at approximately every 6 hours (2 high and 2 low tides). During aerial exposure the model suspends the feeding processes (Sara et al., 2011) and simulates metabolic depression (Monaco & McQuaid, 2018) where, the Arrhenius thermal sensitivity equation (Eq. 9) is corrected by a metabolic depression constant ($M_d = 0.15$), a value representative for *M. galloprovincialis* and here applied also for *M. edulis*. In the present study, the mussel’s body temperature change during low tide is ignored, inducing a model error. The mussel’s body temperature (i.e. surrounding water temperature for submerged mussels) during air exposure depends on many factors, such as solar radiation, air’s temperature, wind speed and wave height, according to studies investigating the temperature effect on intertidal mussels (Kearney et al., 2010, Sara et al., 2011). However, the present study aims to primarily examine the MPs accumulation and thus the intertidal mussel’s body temperature was not thoroughly examined. Nonetheless, the time that the mussel is able to filter, ingest and excrete the suspended matter (i.e. food and MPs particles) and the effect on the mussel’s growth through the modified relation of $k(T)$ are included, since the assimilation process occurs whether the mussel is submerged or not (Kearney et al., 2010).

### 2.5 Parameter values

Most of the DEB model parameters were obtained from Van der Veer et al. (2006) and are referred to the blue mussel *M. edulis* in the northeast Atlantic (see Table 3 for the exceptions).

This assumption has also been adopted in previous studies which showed that this parameter set for *M. edulis* applies also for *M. galloprovincialis* (i.e. Casas and Bacher, 2006, Hatzonikolakis et al., 2017). The half saturation coefficient $X_k$ represents the density of food at which the food uptake rate reaches half of its maximum value and should be treated as a site – specific parameter (Troost et al., 2010, Pouvreau et al., 2006). In order to estimate the value of $X_k$, a different approach was followed for each study area.

For the North Sea simulation, $X_k$ was tuned so that the simulated individual has the recorded size at the corresponding estimated age (Van Cauwenberghe et al., 2015) growing with the representative growth rates of wild *M. edulis* at the region (Saraiva et al., 2012, Sukhotin et al., 2007). For the N. Ionian Sea simulation, an alternative method was adopted, aiming to generalize
the DEB model to overcome the problem of site-specific parameterization. The DEB model was
tuned against literature field data for cultured mussels originated from different areas in the
Mediterranean and Black Seas, where the average CHL-a concentration ranged between 1.0 and 5.0
mg chl-a m$^{-3}$, and one $X_k$ value was found for each area. The four areas used, their characteristics
and the corresponding value of $X_k$ adopted, are shown in Table 4. These values of $X_k$ are related to
the prevailing CHL-a concentration of each area ([CHL-a]) through three different functions:
linear: $f(x) = a * [CHL - a] + b$ exponential: $f(x) = a * \exp(b * [CHL - a])$ and power: $f(x) =
a * [CHL - a]^b + c$. The curve fitting app of Matlab (Matlab R2015a) was used for the
determination of $a$, $b$ and $c$ of each function taking into account the 95% confidence level. The
score of each function regarding the somatic/mussel growth simulation in all four regions is tested
through target diagrams (Jolliff et al., 2009) by computing the bias and unbiased root-mean-square-deviation (RMSD) between field and simulated data of all 4 regions and the function with the best
score is adopted. A similar approach was followed by Alunno-Bruscia et al. (2011) for the oyster
Crassostrea gigas in six Atlantic ecosystems who expressed the $X_k$ as a linear function of food
density (e.g. phytoplankton). Unfortunately, the approach described for the N. Ionian Sea
simulation could not be applied in the North Sea, as the limited amount of growth data from the
literature for wild M. edulis in similar environments did not permit a statistically significant fit of a
similar function ($X_k = f(chl - a)$).

2.6 Simulation of reproduction-Initialization of the model

The reproductive buffer (R) is assumed to be completely emptied at spawning (R = 0)
(Sprung, 1983, Van Haren et al., 1994). In order to simulate mussel’s spawning, the gonado-somatic
index (GSI) defined as gonad dry mass over total dry flesh mass was computed at every model’s
time step (Eq. 17 Table 1; the water content of the fresh tissue mass was assumed 80% according to
Thomas et al. (2011)). Spawning was induced by a critical value of GSI (GSI$^{th}$, Table 3) and a
minimum temperature threshold (T$^{th}$) at each study area, obtained from the literature. In the North
Sea implementation, T$^{th}$ was set at 9.6 °C (Saraiva et al., 2012), while in the N. Ionian Sea, at 15 °C
(Honkoop and Van der Meer, 1998). This kind of formulation for the spawning event in bivalves
has been used in previous studies (i.e. Pouvreau et al., 2006, Troost et al., 2010, Thomas et al.,
2011, Monaco & McQuaid, 2018). The simulated abrupt losses of the mussel's tissue mass
correspond to spawning events and the model’s prediction was compared with the available
literature data regarding the spawning period in each study area. Theodorou et al. (2011)
demonstrated that the spawning events occur during winter for M. galloprovincialis in the mussel
farms of Greece, while in the North Sea the spawning period for *M. edulis* is extended from the end of April until the end of June (Sprung, 1983, Cardoso et al., 2007).

In both areas, the model was initialized so that the simulated individual is in the juvenile phase (\(V < V_p\); Table 3) and the reproductive buffer can be considered to be empty (\(R = 0\)) (Thomas et al., 2011). As stated by Jacobs et al. (2015) amongst others, juvenile mussels (*M. edulis*) range between 1.5-25 mm in size. Specifically, in the North Sea the settlement of mussel larvae (*M. edulis*) takes place in June and the juveniles grow to a maximum size of 25 mm within 4 months (Jacobs et al., 2014). In the N. Ionian Sea, the operating mussel farms follow the life cycle of *M. galloprovincialis*, starting the operational cycle each year by dropping seed collectors from late November until March and the juvenile mussels grow up to 6-6.5 cm after approximately one year according to the information obtained from the local farms in the region and Theodorou et al. (2011). The initial fresh tissue mass was distributed between the structural volume (\(V\)) and reserves energy (\(E\)). Energy allocated to those two compartments was firstly constrained by the initial length (\(L\)) and then energy allocated to \(V\) was in Eq. 10 (Table 1). The initial value of \(E\) was set so that the simulated individual has an initial weight that corresponds to the juvenile phase (\(V < V_p\)) (Table 5). Finally, for both model implementations, the initial accumulation of MPs in the mussel’s tissue (\(C\)) was set to zero.

### 2.7 Simulation Runs

The DEB-accumulation model simulates at an hourly basis the growth and MPs accumulation of the wild mussel from the North Sea and the cultured mussel from the N. Ionian Sea. Initially, a model run is performed at each study area during the periods July 2007 to August 2011 (4 years) for the North Sea simulation and late November 2014 to January 2016 (~ 1 year) for the N. Ionian Sea simulation. Additionally, the inverse simulations were performed in order to evaluate the depuration phase of both cultured and wild mussel, by setting the environmental MPs concentration equal to zero (\(C_{env}=0\)), after a period of 1 year simulation at the N. Ionian Sea, when the cultured mussel has the appropriate size for market, and after 4 years at the North Sea, when literature field data are available (Van Cauwenberghe et al., 2015). In this simulation, the mussel’s gut clearance is achieved by the excretion of MPs through faeces (3rd term of Eq. 18), and thus it is necessary to maintain the existence of food in the mussel’s environment in order to ensure that the feeding-excretion processes will occur.

Furthermore, to examine the model’s uncertainty related to the environmental MPs concentration, a series of 15 and 13 simulations were performed in the North Sea and N. Ionian Sea
respectively, adopting different constant values of $C_{env}$ within the observed range of each area.

Finally, the effect of the environmental forcing data and some model’s parameters on the resulting MPs accumulation by both mussels was explored through sensitivity experiments. These were used to derive a new function that predicts the level of MPs pollution in the environment.

### 2.8 Sensitivity tests and Regression analysis

The effect of the environmental data (CHL-a, temperature, $C_{env}$) and two parameters representative of mussel’s growth ($X_k$, $Y_k$) on the MPs accumulation by the mussel for each study area was examined through sensitivity experiments with the DEB-accumulation model. Each variable (CHL-a, $T$, $C_{env}$) and parameter ($X_k$, $Y_k$) was perturbed by ±10% to examine their effect on the simulated MPs accumulation, and the results of each run were analyzed using the sensitivity index ($SI$), which calculates the percentage change of the mussel’s MPs accumulation $SI = \frac{1}{n} \sum_{t=1}^{n} \left( \frac{|c_t^1 - c_t^0|}{c_t^0} \right) \cdot 100 \%$, where $n$ is the simulated time steps, $c_t^0$ is the MPs accumulation predicted with the standard simulation at time $t$ and $c_t^1$ is the MPs accumulation with a perturbed variable/parameter at time $t$; for details see Bacher and Gangnery, (2006). The same method has been also applied to other studies, which examined the model’s sensitivity on specific variables/parameters regarding the mussel growth (Casas and Bacher, 2006, Rosland et al., 2009, Béjaoui-Omri et al., 2014, Hatzonikolakis et al., 2017). In order to also examine the effect of tides, in the North Sea implementation, the sensitivity experiments were conducted twice: the first time assuming that the mussel is permanently submerged and the second time assuming that the mussel is periodically exposed to the air.

Preliminary sensitivity experiments showed that the MPs accumulation is highly depended on the prevailing conditions regarding the CHL-a, temperature and $C_{env}$ and the mussel’s growth that is regulated by the half saturation coefficient ($X_k$). Therefore an attempt was made using the model’s output to describe the MPs accumulation as a function of these variables through a custom regression model:

$$y = b_1 \times W + b_2 \times \exp \left( \frac{1}{T} \right) + b_3 \times \frac{1}{[CHL-a]} + b_4 \times C_{env} \quad \text{(Eq. 19)}$$

where $y$ (particles/individual) is the response variable and represent the predicted MPs accumulation by the mussel; $W$ (g) the mussel’s fresh tissue mass, $T$ (K) the sea surface temperature, CHL-a and $C_{env}$ are the concentrations of chlorophyll-a and MPs in the water respectively, which are the predictor variables. The values of coefficients $b_1$, $b_2$, $b_3$, and $b_4$ are...
calculated using the nonlinear regression function (nlinfit, Matlab R2015a) which attempts to find values of the parameters $b$ that minimize the least squared differences between the model’s MPs accumulation output $C$ and the predictions of the regression model $y = f(W, T, [chl a], Cenv, b)$.

The ultimate aim of this analysis, once coefficients are determined, is to use Eq. 19 to obtain the environmental MPs concentration:

$$C_{env} = \frac{1}{W_{Mussels} b_4} \left( C - b_1 + b_2 \exp \left( \frac{1}{T} \right) - b_3 \left[ CHL-a \right] \right)$$

(Eq. 20)

which could be a very useful tool to predict the MPs concentration in the environment, when all involved variables are known (mussel size, mussel’s accumulated MPs ($C$), wet weight ($W$) temperature ($T$) and CHL-a), using the mussel as a potential bioindicator (Li et al., 2016, Li et al., 2019). The score of this custom model was tested by applying Eq. 20 in our study areas and 6 more areas around the U.K., where information of mussel’s wet weight and both mussels’ and environment’s MPs load is available (Li et al., 2018). CHL-a and temperature, which were not included in Li et al. (2018), were obtained from daily satellite images (same source as in the North Sea, see 2.4 section), covering the period that the mussels were harvested (Li et al., 2018).

3. Results

3.1 Growth simulations

The growth simulations of $M. edulis$ and $M. galloprovincialis$ for the North Sea and the N. Ionian Sea are shown in Fig. 34 and Fig. 45 respectively. In the North Sea implementation, $X_k$ was tuned to a constant value: $X_k=8$ mg chl-a m$^{-3}$ ($\pm 1.5$ mg m$^{-3}$). The fitted value was slightly higher, as compared to the one ($X_k=3.88$ mg m$^{-3}$ joint) used by Casas and Bacher (2006) in productive areas of the French Mediterranean shoreline (average CHL-a concentration 1.45 mg m$^{-3}$ joint; maximum peak at 20 mg m$^{-3}$ joint), as a consequence of given the even higher productivity in the North Sea (average CHL-a concentration 4.25 mg m$^{-3}$ joint; maximum peak at ~33.40 mg m$^{-3}$ joint). The high value of $X_k$ could also be explained by the presence of silt and other inedible particles (i.e. MPs) which led result to lower quality food in the mussel’s diet compared with an assumed “clean of inedible particles” site environment (Kooijman, 2006, Ren, 2009). In the present study the inedible particles (i.e. MPs) have been incorporated in the mussel’s diet through
the modified relation of the functional response \( f \) (Eq. 5, Table 1), which regulates the assimilation rate and thus the mussel’s growth. However, the DEB model applied at the French site, did not account inedible particles in the mussel’s food. Furthermore, it has been reported that wild mussels grow considerably slower than farmed mussels (~1.7 times) (Sukhotin and Kulakowski, 1992) and thus, a higher value of \( X_k \) promotes a lower mussel growth, which is the case of the North Sea mussel. The simulated mussel shell length after 4 years, in August, is 4.35 cm and the fresh tissue mass is 1.87 gr, in agreement with Van Cauwenberghe et al. (2015) and other studies conducted on wild mussels (Sukhotin et al., 2007, Saraiva et al., 2012, MarLIN, 2016). In particular, Saraiva et al. (2012) found that after 16 years of simulation, the wild mussel of the Wadden Sea (North Sea) is 7 cm long, while according to Bayne and Worral (1980) a mussel with shell length 4 cm corresponds to the age of 4 years, in agreement with the current study. The simulated growth presents a strong seasonal pattern, being higher during spring and summer season, as compared to autumn and winter, which is consistent with the seasonal cycle of temperature and CHL-a concentration, for a typical year in the region (Fig. 1). The increase of food availability and temperature during spring (April) results in high mussel growth for a 4-month period, while the decrease of CHL-a from summer until the end of the year, in conjunction with the temperature decrease in autumn, result in a lower mussel growth. Spawning events that occurred each year in late April and beginning of May (30 April-2 May) each year, are responsible for the sharp decline in mussel’s fresh tissue mass, shown in Fig. 4 (Handa et al., 2011; Zaldivar, 2008) and in agreement with the literature (Sprung, 1983, Cardoso et al., 2007, Saraiva et al., 2012). The predicted weight loss due to spawning was around 7% at the first year of simulation, while the second, third and fourth year the percentage of weight loss increased gradually to 8.3%, 12.6% and 14.4% respectively. Bayne and Worral (1980) demonstrated that the weight losses on spawning for individuals of 1 g weight vary between 2.1% and 39.8%, presenting a weight-specific increase with size.

In the N. Ionian Sea implementation, \( X_k \) is applied as a function of CHL-a concentration through the method described in section 2.5. The target diagram showing the performance of each tested function (linear: \( f(x) = a \times [CHL - a] + b \), where \( a = 0.959 \) and \( b = -1.420 \); exponential: \( f(x) = a \times \exp(b \times [CHL - a]) \) where \( a = 0.2 \) and \( b = 0.567 \); power: \( f(x) = a \times [CHL - a]^b + c \) where \( a = 0.01, b = 3.529 \) and \( c = 0.480 \) ) is shown in Figure 5. The linear and power function of \( X_k \) present a good skill, with the power function leading to the most successful simulation of the cultured mussel’s growth in all four areas (diagram marks for mussel length and fresh tissue mass are closer to the target’s center). The power function applied in the N. Ionian Sea, resulted in mussel’s shell length 5.8 cm and fresh tissue mass 5.92 gr after one year simulation, in
agreement with Theodorou et al. (2011). The spawning event occurred at the beginning of December (Theodorou et al., 2011) and was illustrated by a 12.6% tissue mass decline.

3.2 Microplastics accumulation and depuration phase

The hourly simulated MPs accumulation by the mussel in the North Sea and N. Ionian Sea are shown in Fig. 6 and Fig. 7 respectively. Calibration of the parameter $k_f$ ($1.2 \text{ d}^{-1}$) led to a model which was well fitted to the observed MPs accumulation in the mussel of both study areas. In the North Sea, a 4-year-old wild mussel ($L=4.35 \text{ cm}, W=1.87 \text{ g}$) contains 0.5364 particles individual$^{-1}$ in August within the range value found by Van Cauwenberghe et al. (2015) (0.4 ± 0.3 particles individual$^{-1}$), although the model overestimated the data range reproducing a seasonal increase that was not observed. This is most likely due to the fact that Van Cauwenberghe et al. (2015) allowed a 24 h clearance period, before analyzing the mussels' tissue for MPs, possibly resulting in slightly lower MPs accumulation than the model’s prediction. The MPs egested through faeces by the 4 year old mussel after 24 h were 0.2 ± 0.2 particles individual$^{-1}$ (Van Cauwenberghe et al., 2015), which agree also with model’s output (0.3 particles individual$^{-1}$, Fig. 8) regarding the depuration phase and could compensate for the observed difference in mussel’s MPs load between the simulated and field data. In the N. Ionian Sea, the simulated MPs accumulation by the cultured mussel with $L=4.858 \text{ cm}$ and $W=3.343 \text{ g}$ was 0.91 particles individual$^{-1}$ in the end of June/July, in agreement with field observations obtained from Digka et al. (2018a) (0.9 ± 0.2 particles individual$^{-1}$). Overall, the developed model simulated the MPs accumulation by both mussels in the two different areas, using the same parameter set (see Table 3 for the exceptions), under the assumption that parameters referred to silt particles (i.e. inedible particles) may be used to describe also the MPs accumulation. Both simulations were in good agreement with the available field data, with a small deviation for the North Sea. In both regions, the model computed MPs accumulation, assuming that the mussel treats MPs as silt particles (i.e. inedible particles) and is in agreement with the available field data. This may lead to the assumption suggests that mussels probably present a common behavior against all inedible particles. In model’s results, based on the uptake and excretion rates of MPs by the mussels in both study areas, the majority of MPs are rejected through pseudofaeces and fewer through faeces production (not shown). This is in agreement with Woods et al. (2018) who found that most microplastic fibers (71%) were quickly rejected as pseudofaeces and < 1% excreted in faeces.

The small-scale (daily) fluctuations of MPs in the mussel (wild and cultivated) reflect the adopted random variability of the environmental MPs concentration $C_{env}$ and the daily fluctuations
of the environmental forcing (CHL-a, temperature). The large-scale (seasonal) variability follows mainly the variability of the clearance rate. The seasonal variability of the CHL-a concentration and temperature greatly determines the variability of the clearance rate and hence the variability of MPs in the individual. Moreover, the model predicts that mussel’s energy needs are increased as it grows and therefore the clearance rate is increased, resulting in higher MPs accumulation.

The simulated time needed to clean the mussel’s gut from the MPs load for both areas is shown in Fig. 8. In both areas, the cleaning follows an exponential decay, in agreement with laboratory experiments by Woods et al. (2018). In particular, the model predicts a 90% mussel’s cleaning after 284330 hours (~124 days) and 5663 hours (~2.53 days) for the N. Ionian Sea and North Sea respectively. The cleaning process is more rapid in the North Sea simulation, which can be attributed to the higher CHL-a concentration found in this area, leading to increased production of faeces by the mussel and hence faster excretion of the accumulated MPs. In the N. Ionian Sea, on the other hand, the rate of the mussel’s cleaning is slower, due to the limited food availability.

3.3 Model’s uncertainty regarding the environmental microplastics concentration

The MPs concentration in the environment presents a strong variability in both temporal and spatial scales. To examine the model’s uncertainty related to the environmental MPs concentration ($C_{env}$), a series of 15 and 13 simulations were performed in the North Sea and N. Ionian Sea respectively, adopting different values of $C_{env}$ within the observed range of each area. In the North Sea, the adopted $C_{env}$ ranged between 0.1 and 0.8 particles L$^{-1}$ with a step of 0.05 (15 runs), while in the N. Ionian Sea $C_{env}$ ranged between 0.0012 and 0.0252 particles L$^{-1}$ with a step of 0.002 (13 runs). The mean seasonal values and standard deviation of the 15 simulations in the North Sea and the mean monthly values and standard deviation of the 13 simulations in the N. Ionian Sea were computed and plotted in Fig. 9 and Fig. 10, respectively. Each error bar represents the uncertainty of the simulated accumulation at the specific time, related to the environmental MPs concentration.

In both case studies, the uncertainty of the model appears to increase as the MPs accumulation is increased. As the mussel grows in the North Sea, the mean value and standard deviation of MPs accumulation is increased during the same season every year, illustrating the effect of the mussel’s weight. Moreover, the seasonal variability of the MPs accumulation appears to be related with should be caused by the seasonality of CHL-a concentration. This is apparent during each year’s spring: when CHL-a concentration peaks at its maximum value (~30 mg m$^{-3}$; see Fig. 1), the filtration rate is decreased (Riisgard et al., 2003, 2011), leading to lower MPs accumulation by the mussel and thus lower model’s uncertainty. In the N. Ionian Sea, the effect of the mussel’s weight is
more apparent in the early months (~ 6 months), resulting on higher MPs accumulation and model uncertainty as the mussel grows. Afterwards, the seasonality of both CHL-a concentration and temperature plays the major role. During summer, when the CHL-a concentration is progressively decreased, reaching minimum values (~0.7 mg /m^3) and temperature is increased (>20º C), the filtration rate is significantly decreased or stopped, resulting in lower MPs accumulation and lower model’s uncertainty. This is in line with studies reporting that the mussel suspends the filtering activity and thus closes its valves until better conditions occur (Pascoe et al., 2009, Rissgard et al., 2011). Overall, the available field data lie within the model’s uncertainty, apart from the North Sea case, where the range of field data variability and model uncertainty do not overlap significantly at the time of the observations for both study areas.

Moreover, to evaluate the scenario adopted with the set-up of the previous experiments (random C_{env} at a daily time step) 3 additional model runs are performed in each study area, adopting each time different stochastic sequences of daily random C_{env} values within the observed range, which is considered to reflect the high spatial and temporal variability of the environmental MPs concentration. The mean value and standard deviation of these “stochastic” runs lie most of the time within the standard deviation of the overall model’s uncertainty in both case study areas (Fig. 9 and Fig. 10).

### 3.4 Sensitivity and Regression analysis results

The results of the sensitivity experiments regarding the MPs accumulation by the mussels are shown in Fig. 11 and 12 for the North Sea and N. Ionian Sea respectively. The comparison between the intertidal and subtidal mussel of the North Sea revealed that both +10% and -10% perturbation of CHL-a and X_k have a slightly lower effect on the MPs accumulation by the intertidal mussel which is probably attributed to the intermittent feeding periods experienced by the individual due to the tide effect. As far as the temperature effect, both +10% and -10% perturbed value led to higher sensitivity on the MPs accumulation by the intertidal mussel, due to the adopted modified temperature relation during low tide. Especially, if the mussel’s body temperature change during air exposure would be considered, the perturbed temperature will probably affect even more the MPs accumulation on the intertidal than the subtidal mussel. The sensitivity effect of the C_{env} is slightly higher and lower on the MPs accumulation by the intertidal mussel when perturbed either +10\% or -10\% is almost the same for the intertidal and subtidal mussel, respectively, however the difference of the sensitivity index (%) between the two mussels (intertidal vs. subtidal) is small.
indicating that the environmental MPs concentration affects similarly both mussels, regardless the continuous or intermittent feeding-excretion process.

The comparison between the mussel sensitivity indexes in the N. Ionian and the North Sea (in conditions of submergence) study areas reveals some important differences. Generally, most of the perturbed (either +10% or -10%) variables and parameters (i.e. CHL-a, temperature, $X_k$) present higher sensitivity on the MPs accumulation by the mussel from the N. Ionian Sea. This is attributed to the prevailing environmental conditions and specifically the lower food availability (CHL-a) and the higher temperature range in the N. Ionian Sea compared to the North Sea, which greatly determine the feeding processes, the mussel’s growth and hence the MPs accumulation. The perturbed $C_{env}$ in both study areas appears to affect similarly the MPs accumulation on both mussels ($\sim$10%), with the small difference (<2%) probably attributed to the higher abundance of seawater’s MPs present in the North Sea compared to the N. Ionian Sea. Finally, the half saturation coefficient for the inorganic particles ($Y_k$) has no effect on the MPs accumulation of both North Sea and N. Ionian Sea mussels, indicating that the amount of inedible particles (i.e. MPs) is relatively low in both areas and thus the $Y_k$ does not affect the way that the organic particles are being ingested (Kooijman, 2006). According to Ren (2009), when the inorganic matter is low, the $K(y)$ (Eq. 5; Table 1) is approximately equal to $X_k$ and then $Y_k$ is the least sensitive parameter for the ingestion rate and thus growth.

The DEB-accumulation model output was used to determine the coefficients in Eq. 19 by the nonlinear regression analysis: $b_1 = 0.1909 \pm 0.0006$, $b_2 = 0.0412 \pm 0.0019$, $b_3 = 0.1315 \pm 0.0021$ and $b_4 = 1.1060 \pm 0.0253$. The accurate estimation of the confidence intervals for the estimated coefficients ($b_1$, $b_2$, $b_3$, $b_4$) is indicated by the low confidence intervals, while the mean squared error of the regression model appears also sufficiently small—small enough which indicates an accurate estimation of them and the mean squared error of the regression model is small enough (MSE=0.0523). Subsequently, as shown in Figure 13, Equation 20 may be used to predict the MPs concentration of the environment where mussels live, being in most cases, the predicted MPs concentration is found within the standard deviation of the field data. Two exceptions are shown in Hastings-A and Plymouth areas. The reasons behind these discrepancies may be related to the environmental conditions prevailing in each area at the sampling time. For example, Eq. 20 does not take into account the impact of tides that may affect the mussel’s MPs load ($C$) and the lack of information on the exact sampling date led to using a mean SST and CHL-a value representative of the given sampling time period (Li et al., 2018). Although, Eq. 20 does not account for the tide effect, however, the sensitivity analysis (Fig. 11) showed that the effect of $C_{env}$ on the mussel’s MPs accumulation was the same for both intertidal and subtidal mussel in the North Sea. This result may also apply at the two exceptions areas, leading to the assumption that the
discrepancies are due to the lack of the ambient temperature and CHL-a information during the sampling date. In any case, however, this is just a first rough demonstration of the method and should be implemented in more environments in order to be further validated.

4. Discussion

A DEB-accumulation model was developed and validated with the only available data from the North Sea and the N. Ionian Sea, to study the MPs accumulation by wild M. edulis and cultured M. galloprovincialis, as they grown in different, representative environments. Although the study is limited by scarce validation data, it should be noted the MPs accumulation model parameter set, except one tuning parameter \( k_f \), was extracted from the literature (Table 3), assuming it is notable that the accumulation submodel’s parameters are extracted from literature (Table 3) illustrating that mussels adopt a common defensive mechanism against inedible particles (i.e. silt, MPs). Thus, the theoretical background constructed by Saraiva et al., (2011a) (based on Kooijman, 2010) regarding the feeding and excretion processes of the mussel remains unspoiled.

Through the strong theoretical background of DEB theory, this study highlights that the accumulation of MPs by the mussel is highly depended on the prevailing environmental conditions which control the amount of MPs that the mussel filtrates and excretes.

Towards a generic DEB model

Beginning with the generalization of the DEB model regarding the site-specific parameter \( X_k \) in the N. Ionian Sea simulation, the applied function of the half saturation coefficient \( f(x) = a \star [CHL - a]^b + c \) successfully captures the physiological responses and thus the growth rate of the cultured mussel at the N. Ionian Sea implementation. In the current study, a demonstration of this method led conducted leading to a robust and generic DEB growth model able to simulate robust enough with a sufficiently generic nature for the simulation of the mussel growth in representative mussel habitats of the Mediterranean Sea, covering a range of productivity and sea surface temperature. This approach supports and takes a step further of Bourles et al. (2008) suggestion about a suggested for oyster growth (Crassostrea gigas) that a seasonally varied half saturation coefficient, demonstrating an improvement could improve the accuracy of the food quantifier. The applied function of \( X_k \) considers the daily CHL-a fluctuations, and thus the seasonal variation of the seawater composition, because seawater composition is closely related to the season. As more field data becomes available from various environments, such an approach could result to more generic formulations for the site-
specific parameter $X_k$, so that the model could be applied in several areas of interest, where field
growth data are absent and/or to simulate the potential mussel growth in the 2D space.

The simulation of MPs accumulation by the mussels, using the DEB-accumulation model, is
in good agreement with the available field data (Fig. 3 and Fig. 4). The simulated values lie within
the observed field data range (mean ±SD), although the seasonal increase reproduced by the model
at the North Sea implementation did not exactly overlap with the field data at the time of
observations. This could be attributed to the clearance period (24 h) that allowed mussels to excrete
MPs through faeces (0.2 ± 0.2 particles individual$^{-1}$) before the mussel’s tissue analysis (Van
Cauwenberghe et al., 2015). The measured loss of mussel’s MPs is in agreement with the model’s
result on the depuration experiment after 24 h. The MPs accumulation by the cultivated mussel
(fresh tissue mass 3.3342 g) originated from the N. Ionian Sea with mean $C_{env} = 0.0012 ± 0.024$
particles L$^{-1}$, is 0.91 particles individual$^{-1}$ and by the wild mussel (fresh tissue mass 1.87 g) from the
North Sea with mean $C_{env} = 0.4 ± 0.3$ particles L$^{-1}$ is 0.5364 particles individual$^{-1}$. If these
concentrations are expressed per gram of wet tissue of mussels, the cultivated mussel contamination
(0.27 particles g$^{-1}$w.w.) is comparable with the wild mussel (0.2834 particles g$^{-1}$w.w.), despite the
much lower environmental MPs concentration ($C_{env}$) in the N. Ionian Sea than the North Sea. This
comparison aims to highlight the significant impact of the prevailing environmental conditions
(CHL-a and temperature) on the MPs accumulation by the mussels, although they originate from
different areas and lived different time period. The generally high abundance of CHL-a in the North
Sea simulation, contributes to a reduction of the filtering activity and hence of the MPs
accumulation. The threshold algal concentration for reduction of the mussel’s filtration rate
(incipient saturation) has been found to lie between 6.3 and 10.0 mg m$^{-3}$ μg chla L$^{-1}$ (Riisgard et al.,
2011), which is a range comparable to the CHL-a concentrations in the North Sea case.
Furthermore, in the N. Ionian Sea simulation, the filtration, ingestion, pseudofaeces and faeces
production rates are decreased during the summer season when the CHL-a and temperature has
downward and upward trend respectively, gradually leading to a decline of the mussel’s MPs
accumulation—Van Cauwenberghe and Janssen (2014) found that cultivated $M. edulis$ from the
North Sea contained on average 0.36 ± 0.07 particles g$^{-1}$w.w., a slightly higher similar value with
that found in the present study for the wild mussel of the North Sea (0.2834 particles g$^{-1}$w.w.). This
could be attributed to mussel farms acting as a potential source of MPs contamination for the
mussels due to plastic materials (i.e. plastic sock nets and polypropylene long lines) used during
cultivation (Mathalon and Hill, 2014, Santana et al., 2018). This probably highlights the small
contribution of mussel farms as a source of MPs pollution (Santana et al., 2018). Moreover, the
intertidal wild mussel (present study) is assumed to filter and excrete MPs half of the time in
comparison with the submerged cultured mussel in the North Sea, resulting though in similar
accumulation level. The model also predicts the time needed for the 90% gut clearance of both cultured (N. Ionian Sea) and wild (North Sea) mussel to be almost 284 hours and 565 hours (equivalent to 12 and 2.5 days) respectively, when MPs contamination is removed from their habitat. This is in line with a series of studies which demonstrated that the depuration time varies between 6-72 hours and can last up to 40 days depending on several factors such as species, environmental conditions (Bayne et al., 1987), size and type of MPs (Browne et al., 2008, Ward and Kach, 2009, Woods et al., 2018, Birnstiel et al., 2019).

The strong dependence of food (CHL-a), temperature and seawater’s MPs concentration on the MPs accumulation by the mussel, regarding its wet weight, is demonstrated through sensitivity experiments that were used to derive a rather simple nonlinear regression model (Eq. 1). The comparison of the regression model’s with the DEB model’s output resulted in a quite accurate estimation of the coefficients, which in turn sparked the idea of a ‘new’ relationship (Eq. 2) that could potentially predict the MPs concentration in the environment ($C_{env}$) when certain conditions are known (CHL-a, T, $C_{env}$, W). The latter equation was applied in 8 areas in total (2 from the present study areas and 6 from Li et al. (2018)), with relatively good results since there is general overlapping of regressed and observed MPs concentration in the environment ($C_{env}$), except for Hastings-A and Plymouth areas, probably due to missing information on the environmental conditions (CHL-a, SST) during the sampling.

The predicted value is within the observed range of field data in most regions, suggesting that making the mussels can be used as potential bioindicators. Mussels have been previously proposed as bioindicators for marine microplastic pollution (<1 mm), although the efficient gut clearance and selective feeding behavior limit their quantitative ability (Lusher et al., 2017, Brate et al., 2018, Beyer et al., 2017, Fossi et al., 2018, Li et al., 2019). The very recent study by Ward et al. (2019) demonstrated that bivalves are poor bioindicators of MPs pollution due to the particle selection during feeding and excretion processes that is based on the physical characteristics of the MPs. Considering that the MPs accumulation is site-dependent and that sampling of mussels is usually easier than seawater (Karlsson et al., 2017, Brate et al., 2018), models like the one described in Eq. 2 that besides the MPs accumulation take also into account characteristics of the environment, that which are crucial for the way that mussels accumulate MPs.

This method could be possibly used at global level and allow comparisons between various environments. However, the method described should be validated in more environments with more frequent field data to be able to provide secure results.

In addition to the scarce validation data regarding the MPs accumulation in mussels, in this study has some more limitations. First of all, the data regarding the concentration of MPs in the mussels’ environment are also scarce; since MPs is a relatively recent subject of study, the existing knowledge of the spatial and temporal distribution is still quite limited...
To overcome the lack of environmental MPs time series, a function of randomly generated values within the observed range of each area was applied and its uncertainty was examined through an ensemble forecasting. Specifically, the model’s uncertainty due to the environmental MPs concentration ($C_{env}$) was tested by performing a series of model runs forced by an envelope of representative values of $C_{env}$ and $T$.

The results (section 3.3) showed that the adopted stochastic scenario simulated quite satisfactorily realistically the MPs accumulation by the mussels, lying within the observed field range, although a slight overestimation was found in the North Sea and in agreement with the available field data. The approach used is assumed to represent the natural variability close to reality since it has been reported that MPs quantification in the water is rather a complicated procedure due to the influence of many factors such as tides, wind, wave action, ocean currents, river inputs and hydrodynamic features leading to high spatially and temporally variability of MPs distribution even in very small scales (Messinetti et al., 2018, Goldstein et al., 2013). In addition, the nature of the variable $C_{env}$ makes it difficult to estimate, presenting large observational errors, not only due to the intense physical variation but also due to different sampling and analysis techniques that were used. In a future work the DEB-accumulation model could be coupled with a high-resolution MPs distribution model (Kalaroni et al., 2019), being extensively validated against field data that will have been collected and processed according to a common scientifically defined protocol, to overcome this limitation. Moreover, the approach followed in calculating the value of MPs concentration in the near surface layer (0-5m depth) (Kooi et al., 2016), resulted in a representative value of the upper ocean layer. In depth knowledge of the MPs distribution, both horizontally and vertically, is essential to understand and mitigate their impact not only on the various marine compartments but also on the organisms inhabiting those compartments (Van Sebille et al., 2015, Kooi et al., 2016). For that reason, it is important to enhance the monitoring activity especially in the vulnerable coastal environments, adopting integrated cross-disciplinary approaches and monitoring of biological, physical and chemical parameters which provide information on the ecosystem function, in order to improve the assessment of emerging pollutants (i.e. MPs) and their impacts on biota (objective of JERICO-RI framework).

Further, the assumption that the mussel has the same filtration rate for all particles independently of their chemical composition, size and shape, is a simplification and an open contradictory theme of discussion (see Saraiva et al., 2011a for details). However, in our model application through the model, a pre-ingestive particle selection by the mussel is implied based on the organic-inorganic content of the suspended matter illustrating the different binding probabilities applied for algal and MPs particles during the ingestion process. Through an investigation of wild
mussel’s faeces and pseudofaeces production in laboratory conditions, Zhao et al. (2018) found that the length of MPs was significantly longer in pseudofaeces than in the digestive gland and faeces. Furthermore, Van Cauwenbergh et al. (2015) demonstrated that mussel’s faeces contained larger MPs (15–500 μm) compared to the mussel’s tissue (20–90 μm). Apparently, smaller sized MPs seem to be dominant within the mussels in comparison with the size of the MPs in the ambient environment (Li et al., 2018, Qu et al., 2018, Digka et al., 2018b), implying that the mussel is more prone to ingest and retain smaller sized MPs. As an example, Digka et al. (2018b) confirmed that the smaller MPs (<1 mm) occupy the 62.3%, 96.9% and 100% of the total MPs in seawater, sediments and mussels from the N. Ionian Sea respectively. In a future work this selection pattern regarding size, could be simulated by suitable preference weights among different MPs sizes. This will improve the knowledge of the feeding and excretion mechanisms used by the mussels against MPs pollution and the assessment of the ecological footprint (Rist et al., 2019).

Our use of the tide data led to some model bias. Furthermore, the tide data as considered in the present study impose model's bias, since the model does not take into account of the mussel's body temperature change when this is exposed to air was not taken into consideration. Assessing the mussel’s body temperature requires extended experiments in field conditions (Tagliarolo and McQuaid, 2015, Monaco and McQuaid, 2018). A very recent study by Seuront et al. (2019)
along the French coast of the eastern English Channel found no significant correlation between air’s and mussel’s body temperature, but demonstrated a rather positively significant positive correlation between the body temperature and with the hard substrate’s (i.e. rocks) temperature. However, in the present study the tide effect on processes that are affected by the thermal equation (k(T)) is considered indirectly through the metabolic depression (details in section 2.4). Sara et al. (2011) following the method developed by Kearney et al. (2010), who coupled a DEB model with a biophysical model (Kearney et al., 2010), incorporating the change of mussel’s body temperature during emersion, by using information of various climatological variables (i.e. solar radiation, air temperature, wind speed, wave height), but the temperature sensitivity on the physiological processes was ignored. In a future study, a combined similar approach by coupling the present DEB-accumulation model with a biophysical model, which includes both the tide effect on the physiological processes and the mussel’s body temperature respectively, could be followed and lead to a more detailed simulation of the intertidal mussel’s body temperature.

5. Conclusions

In a future study the model should be corroborated further by using a larger dataset of validated against more frequent field data regarding the MPs accumulation, with sampling of mussels of various sizes and life stages. Currently, the model is mainly limited by the insufficient validation, as a larger dataset could be also used for a better model calibration, as for now it cannot be reliable in conducting predictions within accepted precision. However, this study provides a new approach in studying the accumulation of MPs by filter feeders and reveals the relations between characteristics of the mussel’s surrounding environment and the MPs accumulation, which is presented with high seasonal fluctuations. Additionally, in a future study the DEB-accumulation model will be coupled to a coupled hydrodynamic-biochemical model (e.g., Petihakis et al., 2002, 2012, Triantafyllou et al., 2003, Tsiaras et al., 2014, Ciavatta et al., 2019, Kalaroni et al., 2020) and a MPs distribution model (Kalaroni et al., 2019) that will provide fields of temperature, food availability and MPs concentration respectively at the Mediterranean scale, and eventually lead to an integrated representation of the MPs accumulation by mussels (Daewel et al., 2008). This fully coupled model will be downscaled to the Cretan Sea SuperSite, while the parameterization of important biological processes will be redesigned based on the new data which will be acquired in the framework of the JERICO S3 project (http://www.jerico-ri.eu). The present study highlights the urgent need for adopting a multi-disciplinary monitoring activity by measuring physical, biological and chemical parameters that are crucial for mapping the MPs distribution, assessing the contamination level of the marine organisms and investigating the impact on the
health status. Overall, despite the significant limitations that were mentioned before, taken into account that plastics are one of the global hot issues, this particular study could help for the design of next efforts, since it provides indications on the future priority related issues.

Author Contribution:
Natalia Stamataki, Yannis Hatzonikolakis, Kostas Tsiaras, Catherine Tsangaris, George Petihakis, Sarantis Sofianos, George Triantafyllou

G.T. conceived the basic idea of the present study and was responsible for the management and coordination of the research planning and execution. N.S. and Y.H. developed the model code with the contribution of K.T.. N.S. collected the existing information on the subject and performed the simulations of the present study with the help of Y.H. when needed. G.T., G.P., K.T., Y.H. and N.S. contributed to the interpretation of the results. C.T. provided the field data of the mussel’s microplastic accumulation in the North Ionian Sea. N.S. prepared the manuscript, with critical review, commentary and revision contributed from all co-authors.

Competing interests:
The authors declare that they have no conflict of interest.

Acknowledgments:
This work was partially funded by the national project ‘Blue growth with innovation and application in Greek Seas’ (MIS 5002438) and the EU H2020 CLAIM project (G.A. n° 774586). This study has been conducted using E.U. Copernicus Marine Service Information (http://marine.copernicus.eu/). Part of this work was supported by the JERICO- NEXT project. This project has received funding from the European Union’s Horizon 2020 research and innovation programme under grant agreement no. 654410.
References


GlobColour data (http://globcolour.info) used in this study has been developed, validated, and distributed by ACRI-ST, France.


Tables & Figures

\[
\frac{dE}{dt} = \dot{p}_a - \dot{p}_c \tag{1}
\]

\[
\frac{dV}{dt} = \frac{k \dot{p}_c - [\dot{p}_M] V}{[E_g]} \tag{2}
\]

\[
\frac{dR}{dt} = (1 - k) \cdot \dot{p}_c - \left[ \frac{1-k}{k} \right] \cdot \min\left( V, V_p \right) \cdot [\dot{p}_M] \tag{3}
\]

\[
\dot{p}_a = \{\dot{p}_{Am}\} \cdot f \cdot k(T) \cdot V^2 \tag{4}
\]

\[
f = \frac{x}{x + K_y}, \quad \text{where} \quad K_y = X_k \cdot (1 + \frac{y}{y_k}) \tag{5}
\]

\[
\dot{p}_c = \frac{[E]}{[E_g] + k \cdot [E]} \cdot \left( \frac{[E_g] \cdot \{\dot{p}_{Am}\} \cdot k(T) \cdot V^2}{[E_m]} + [\dot{p}_M] \cdot V \right) \tag{6}
\]

\[
[E] = \frac{E}{V} \tag{7}
\]

\[
[\dot{p}_M] = k(T) \cdot [\dot{p}_M]_m \tag{8}
\]

\[
k(T) = \frac{\exp\left( \frac{\tau_A}{T_i} \cdot \tau_A \right)}{1 + \exp\left( \frac{\tau_A}{T_i} - \frac{\tau_A}{\tau_L} \right) + \exp\left( \frac{\tau_{AH}}{T_H} - \frac{\tau_{AH}}{T_i} \right)} \tag{9}
\]

\[
L = \frac{V^2}{\delta_m} \tag{10}
\]

\[
W = d \cdot \left( V + \frac{E}{[E_g]} \right) + \frac{R}{\mu_E} \tag{11}
\]

\[
\dot{C}_R = \frac{[C_{Rm}]}{1 + \Sigma_i^n [C_{Rm}]} \cdot k(T) \cdot V^2, \quad i = \begin{cases} 1 \text{ for CHL - a} \\ 2 \text{ for MPS} \end{cases} \tag{12}^a
\]

\[
\dot{p}_{Xif} = \dot{C}_R \cdot X_i \tag{13}^a
\]

\[
\dot{p}_{Xil} = \frac{\rho_{Xl} \cdot \dot{p}_{Xif}}{1 + \Sigma_i^n \rho_{Xl} \cdot \rho_{Xif}} \tag{14}^a
\]

\[
\dot{J}_{pfi} = \dot{p}_{Xif} - \dot{p}_{Xil} \tag{15}^a
\]

\[
\dot{J}_f = \dot{p}_{X1f} - \dot{p}_A \tag{16}
\]
Table 1. Dynamic energy budget model: equations. See Table 2 for model variables, Table 3 for parameters and Table 4 for initial values.

\[ GSI = \frac{\frac{R}{\mu_E}}{d\left(\frac{V}{E} + \frac{R}{\mu_E}\right)} \]  

(17)

\[ \hat{p}_a \] for assimilation energy rate
\[ \hat{p}_c \] for utilization energy rate
\[ \hat{c}_{env} \] for microplastics concentration
\[ \hat{p}_{XIF} \] for filtration rate
\[ \hat{p}_{XII} \] for ingestion rate
\[ \hat{F}_{PF} \] for pseudofaeces production rate
\[ F_f \] for faeces production rate
\[ f \] for functional response function
\[ X_i \] for food or MPs density
\[ [\hat{p}_M] \] for maintenance costs
\[ T \] for temperature
\[ k(T) \] for temperature dependence
\[ L \] for shell length
\[ W \] for fresh tissue mass
\[ GSI \] for gonado-somatic index

Table 2. Dynamic energy budget model: variables

<table>
<thead>
<tr>
<th>Variable</th>
<th>Description</th>
<th>Units</th>
</tr>
</thead>
<tbody>
<tr>
<td>( V )</td>
<td>Structural volume</td>
<td>( \text{cm}^3 )</td>
</tr>
<tr>
<td>( E )</td>
<td>Energy reserves</td>
<td>( \text{J} )</td>
</tr>
<tr>
<td>( R )</td>
<td>Energy allocated to development and reproduction</td>
<td>( \text{J} )</td>
</tr>
<tr>
<td>( C )</td>
<td>Microplastics accumulation</td>
<td>particles individual(^{-1} )</td>
</tr>
<tr>
<td>( \dot{p}_a )</td>
<td>Assimilation energy rate</td>
<td>( \text{J d}^{-1} )</td>
</tr>
<tr>
<td>( \dot{p}_c )</td>
<td>Utilization energy rate</td>
<td>( \text{J d}^{-1} )</td>
</tr>
<tr>
<td>( \dot{c}_R )</td>
<td>Clearance rate</td>
<td>( \text{m}^3\text{d}^{-1} )</td>
</tr>
<tr>
<td>( c_{env} )</td>
<td>Microplastics concentration</td>
<td>particles ( \text{L}^{-1} )</td>
</tr>
<tr>
<td>( \dot{p}_{XIF} )</td>
<td>Filtration rate</td>
<td>( \text{J d}^{-1} ) or ( \text{g d}^{-1} )</td>
</tr>
<tr>
<td>( \dot{p}_{XII} )</td>
<td>Ingestion rate</td>
<td>( \text{J d}^{-1} ) or ( \text{g d}^{-1} )</td>
</tr>
<tr>
<td>( \dot{F}_{PF} )</td>
<td>Pseudofaeces production rate</td>
<td>( \text{J d}^{-1} ) or ( \text{g d}^{-1} )</td>
</tr>
<tr>
<td>( F_f )</td>
<td>Faeces production rate</td>
<td>( \text{J d}^{-1} )</td>
</tr>
<tr>
<td>( f )</td>
<td>Functional response function</td>
<td>-</td>
</tr>
<tr>
<td>( X_i )</td>
<td>Food or MPs density</td>
<td>mg chla m(^{-3} ) or g m(^{-3} )</td>
</tr>
<tr>
<td>( [\dot{p}_M] )</td>
<td>Maintenance costs</td>
<td>( \text{J cm}^3\text{d}^{-1} )</td>
</tr>
<tr>
<td>( T )</td>
<td>Temperature</td>
<td>K</td>
</tr>
<tr>
<td>( k(T) )</td>
<td>Temperature dependence</td>
<td>-</td>
</tr>
<tr>
<td>( L )</td>
<td>Shell length</td>
<td>cm</td>
</tr>
<tr>
<td>( W )</td>
<td>Fresh tissue mass</td>
<td>g</td>
</tr>
<tr>
<td>( GSI )</td>
<td>Gonado-somatic index</td>
<td>-</td>
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<tr>
<td>Parameter</td>
<td>Units</td>
<td>Description</td>
</tr>
<tr>
<td>-----------------------------------------------</td>
<td>------------------------------</td>
<td>----------------------------------------------------------------------------</td>
</tr>
<tr>
<td>${\dot{p}_{Am}}$</td>
<td>$J \text{ cm}^{-2} \text{ d}^{-1}$</td>
<td>Maximum surface area-specific assimilation rate</td>
</tr>
<tr>
<td>${\dot{C}_{Rm}}$</td>
<td>$m^3 \text{ cm}^{-2} \text{ d}^{-1}$</td>
<td>Maximum surface area-specific clearance rate</td>
</tr>
<tr>
<td>${\dot{p}<em>{X</em>{s}Fm}}$</td>
<td>$\text{ mg chl}_a \text{ cm}^{-2} \text{ d}^{-1}$</td>
<td>Algal maximum surface area-specific filtration rate* $0.1152$</td>
</tr>
<tr>
<td>${\dot{p}<em>{X</em>{s}Fm}}$</td>
<td>$g \text{ cm}^{-2} \text{ d}^{-1}$</td>
<td>Silt maximum surface area-specific filtration rate</td>
</tr>
<tr>
<td>${\dot{p}<em>{X</em>{a}m}}$</td>
<td>$\text{ mg chl}_a \text{ d}^{-1}$</td>
<td>Algae maximum ingestion rate*</td>
</tr>
<tr>
<td>${\dot{p}<em>{X</em>{a}m}}$</td>
<td>$g \text{ d}^{-1}$</td>
<td>Silt maximum ingestion rate</td>
</tr>
<tr>
<td>$\rho_1$</td>
<td>-</td>
<td>Algae binding probability</td>
</tr>
<tr>
<td>$\rho_2$</td>
<td>-</td>
<td>Inorganic material binding probability</td>
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<tr>
<td>$k_f$</td>
<td>$d^{-1}$</td>
<td>Post-ingestive losses through faeces</td>
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<tr>
<td>$X_K$</td>
<td>$\text{ mg chl}_a \text{ m}^3$</td>
<td>Half-saturation coefficient</td>
</tr>
<tr>
<td>$T_A$</td>
<td>$K$</td>
<td>Arrhenius temperature</td>
</tr>
<tr>
<td>$T_l$</td>
<td>$K$</td>
<td>Reference temperature</td>
</tr>
<tr>
<td>$T_L$</td>
<td>$K$</td>
<td>Lower boundary of tolerance rate</td>
</tr>
<tr>
<td>$T_H$</td>
<td>$K$</td>
<td>Upper boundary of tolerance rate</td>
</tr>
<tr>
<td>$T_{AL}$</td>
<td>$K$</td>
<td>Rate of decrease of upper boundary</td>
</tr>
<tr>
<td>$T_{AH}$</td>
<td>$K$</td>
<td>Rate of decrease of lower boundary</td>
</tr>
<tr>
<td>$[\dot{p}_{m}Lm]$</td>
<td>$J \text{ cm}^3 \text{ d}^{-1}$</td>
<td>Volume specific maintenance costs</td>
</tr>
<tr>
<td>$[E_G]$</td>
<td>$J \text{ cm}^3$</td>
<td>Volume specific growth costs</td>
</tr>
<tr>
<td>$[E_m]$</td>
<td>$J \text{ cm}^3$</td>
<td>Maximum energy density</td>
</tr>
<tr>
<td>$k$</td>
<td>-</td>
<td>Fraction of utilized energy spent on maintenance/growth</td>
</tr>
<tr>
<td>$V_p$</td>
<td>$cm^3$</td>
<td>Volume at start of reproductive stage</td>
</tr>
<tr>
<td>GSI$_{th}$</td>
<td>-</td>
<td>Gonado-somatic index triggering spawning</td>
</tr>
<tr>
<td>$\delta_m$</td>
<td>-</td>
<td>Shape coefficient</td>
</tr>
<tr>
<td>$d$</td>
<td>$g \text{ cm}^{-3}$</td>
<td>Specific density</td>
</tr>
<tr>
<td>$\mu_E$</td>
<td>$J \text{ g}^{-1}$</td>
<td>Energy content of reserves</td>
</tr>
<tr>
<td>$\lambda$</td>
<td>$J \text{ mg chl}_a^{-1}$</td>
<td>Conversion factor</td>
</tr>
</tbody>
</table>

*Table 3. Dynamic energy budget model: parameters
*units mol C converted to mg CHL-a by multiplying with the factor $\frac{12 \times 10^5}{50}$ assuming Carbon:CHL-a ratio of 50

(Hatzonikolakis et al., 2017).

<table>
<thead>
<tr>
<th>Area</th>
<th>$X_k$ value (mg m$^{-3}$)</th>
<th>CHL-a range (mg m$^{-3}$)</th>
<th>CHL-a mean (mg m$^{-3}$)</th>
<th>Temperature range($^\circ$C)</th>
<th>Length after one year±SD (cm)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maliakos Gulf</td>
<td>0.72</td>
<td>0.87-5.59</td>
<td>1.80</td>
<td>12.0-26.0</td>
<td>7.06 ± 0.46</td>
<td>Hatzonikolakis et al., 2017</td>
</tr>
<tr>
<td>Thermaikos Gulf</td>
<td>0.56</td>
<td>1.04-2.76</td>
<td>1.89</td>
<td>11.5-24.5</td>
<td>7.0 ± 0.47</td>
<td>Hatzonikolakis et al., 2017</td>
</tr>
<tr>
<td>Black Sea</td>
<td>Calibrated: 0.96</td>
<td>0.53-16.30</td>
<td>3.07</td>
<td>6.5-25.0</td>
<td>7.5 ± 0.1</td>
<td>Karayucel et al., 2010</td>
</tr>
<tr>
<td>Bizerte lagoon</td>
<td>3.829</td>
<td>4.00-7.70</td>
<td>5.20</td>
<td>12.0-28.0</td>
<td>7.26 ± 0.46</td>
<td>Béjaoui-Omri et al., 2014</td>
</tr>
</tbody>
</table>

Table 4. Half saturation tuned values ($X_k$) and mussel growth data (Length) in different areas of the Mediterranean and Black Seas.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Value</th>
<th>Variable</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Start date</td>
<td>20 Nov 2010</td>
<td>Start date</td>
<td>1 Jul 2007</td>
</tr>
<tr>
<td>L</td>
<td>0.85 cm</td>
<td>L</td>
<td>0.15 cm</td>
</tr>
<tr>
<td>W</td>
<td>0.1938 g</td>
<td>W</td>
<td>0.0055 g</td>
</tr>
<tr>
<td>V</td>
<td>0.0096 cm$^3$</td>
<td>V</td>
<td>5.3$\times$10$^{-5}$ cm$^3$</td>
</tr>
<tr>
<td>E</td>
<td>350 J</td>
<td>E</td>
<td>10 J</td>
</tr>
<tr>
<td>R</td>
<td>0 J</td>
<td>R</td>
<td>0 J</td>
</tr>
<tr>
<td>C</td>
<td>0 particles individual$^1$</td>
<td>C</td>
<td>0 particles individual$^1$</td>
</tr>
</tbody>
</table>

Table 5. Dynamic energy budget-accumulation model: initial values. L: shell length; W: fresh tissue mass; V: structural volume; E: energy reserves; R: energy allocated to reproduction; C: Microplastics accumulation
Fig. 1. Environmental data used for the forcing of the Dynamic Energy Budget model (DEB) in the North Sea simulation, showing temperature (top) and chlorophyll a concentration against in situ data from the ICES database (bottom).

Fig. 2. Environmental data used for the forcing of the dynamic energy budget model in the Northern Ionian Sea simulation, showing temperature (top) and chlorophyll a concentration (bottom).
Fig. 3. Simulated mussel shell length (L) (top) and fresh tissue mass (W) (bottom) against North Sea data (red star: mean ± SD), using chlorophyll a (X = CHL-a) in the mussel diet.

Fig. 4. Simulated mussel shell length (L) (top) and fresh tissue mass (W) (bottom) against North Sea data (red star: mean ± SD), using chlorophyll a (X = CHL-a) in the mussel diet.
Fig. 53. Target diagram of simulated shell length ($L$) and fresh mass tissue weight ($W$) against field data from Thermaikos and Maliakos Gulf (eastern Mediterranean Sea), Black Sea and Bizerte Lagoon (southwestern Mediterranean Sea), using the 

power ($L_1$, $W_1$), exponential ($L_2$, $W_2$) and linear ($L_3$, $W_3$) function of the half saturation coefficient. The model bias is indicated on the y-axis while the unbiased root-mean-square-deviation (RMSD) is indicated on the x-axis.
Fig. 6. Microplastics (MPs) accumulation by the mussel (blue line) against field data (red star: mean ± SD), using daily environmental concentration of MPs ($C_{env}$ mean value ± SD: 0.4 ± 0.3 particles L$^{-1}$) in the North Sea.

Fig. 7. Microplastics (MPs) accumulation by the mussel (blue line) against field data (red star: mean value ± SD), using daily environmental concentration of MPs ($C_{env}$ mean value ± SD: 0.0012 ± 0.024 particles L$^{-1}$) in the Northern Ionian Sea.

Fig. 8. Depuration phase of the cultured Mytilus galloprovincialis (red line) and wild Mytilus edulis (black line) using zero environmental concentration of microplastics ($C_{env}$=0) after 1 year and 4 years of simulation time at the Northern Ionian Sea and North Sea respectively.
Fig. 9. Mean seasonally values and standard deviation of microplastics (MPs) accumulation (red error bars: mean value ± SD) by the mussel in North Sea derived from 15 model runs with different constant values of environmental MPs concentration ($C_{env}$ range: 0.1-0.8 particles L$^{-1}$); Mean hourly simulated data (black line) and standard deviation (blue lines) of microplastics accumulation derived from 3 model runs with stochastic sequences of daily random $C_{env}$ values.

Fig. 10. Mean monthly values and standard deviation of microplastics accumulation (red error bars: mean value ± SD) by the mussel in Northern Ionian Sea derived from 13 model runs with different constant values of environmental MPs concentration ($C_{env}$ range: 0.0012-0.024 particles L$^{-1}$); Mean hourly simulated data (black line) and standard deviation
(blue lines) of microplastics accumulation derived from 3 model runs with stochastic sequences of daily random $C_{env}$ values.

Fig. 11. Sensitivity index of MPs accumulation on the wild mussel of the North Sea when variables ($CHL$-$a$, temperature, $C_{env}$) and parameters ($X_k$, $Y_k$) are perturbed ± 10%. The notation (s) refers to the permanently submerged mussel.

Fig. 12. Sensitivity index of MPs accumulation on the cultured mussel of the Northern Ionian Sea when variables ($CHL$-$a$, temperature, $C_{env}$) and parameters ($X_k$, $Y_k$) are perturbed ± 10%.
Fig. 13. Prediction of seawater microplastics concentration by using Eq. 20 for the Northern Ionian Sea, North Sea (present study) and 6 areas around U.K. (Filey, Hastings-A&B, Brighton, Plymouth, Wallasey; Li et al. (2018)).