

1 An Evaluation of the Performance of Sea-Bird Scientific's SeaFET™ Autonomous pH Sensor:  
2 Considerations for the Broader Oceanographic Community

3  
4 Cale A. Miller<sup>1,3</sup>, Katie Pocock<sup>2</sup>, Wiley Evans<sup>2</sup>, and Amanda L. Kelley<sup>1\*</sup>

5  
6 1. College of Fisheries and Ocean Sciences, University of Alaska Fairbanks, Fairbanks, AK,  
7 USA

8  
9 2. Hakai Institute, Heriot Bay, BC, Canada

10  
11 3. **Present address:** Department of Evolution and Ecology, College of Biological Sciences,  
12 University of California Davis, CA, USA

13  
14 \*Correspondence to: Amanda L. Kelley (alkelley@alaska.edu)

15  
16  
17 **Abstract**

18  
19 The commercially available Sea-Bird SeaFET™ provides an accessible way for a broad  
20 community of researchers to study ocean acidification and obtain robust measurements of  
21 seawater pH via the use of an *in situ* autonomous sensor. There are pitfalls, however, that have  
22 been detailed in previous best practices for sensor care, deployment, and data handling. Here, we  
23 took advantage of two distinctly different coastal settings to evaluate the Sea-Bird SeaFET™ and  
24 examine the multitude of scenarios in which problems may arise confounding the accuracy of  
25 measured pH. High-resolution temporal measurements of pH were obtained during 3- to 5-month  
26 field deployments in three separate locations (two in south-central, Alaska, USA, and one in  
27 British Columbia, Canada) spanning a broad range of nearshore temperature and salinity  
28 conditions. Both the internal and external electrodes onboard the SeaFET™ were evaluated  
29 against robust benchtop measurements for accuracy utilizing either the factory calibration, an *in*  
30 *situ* single-point calibration, or *in situ* multi-point calibration. In addition, two sensors deployed  
31 in parallel in Kasitsna Bay, Alaska, USA, were compared for inter-sensor variability in order to  
32 quantify other factors contributing to the sensor's intrinsic inaccuracies. Based on our results, the  
33 multi-point calibration method provided the highest accuracy (< 0.025 difference in pH) of pH  
34 when compared against benchtop measurements. Spectral analysis of time series data showed  
35 that during spring in Alaskan waters, a range of tidal frequencies dominated pH variability, while  
36 seasonal oceanographic conditions were the dominant driver in Canadian waters. Further, it is  
37 suggested that spectral analysis performed on initial deployments may be able to act as an *a*  
38 *posteriori* method to better identify appropriate calibration regimes. Based on this evaluation, we  
39 provide a comprehensive assessment of the potential sources of uncertainty associated with  
40 accuracy and precision of the SeaFET™ electrodes.

41  
42 **1 Introduction**

43  
44 The intrusion of excess anthropogenic CO<sub>2</sub> into the global oceans—referred to as ocean  
45 acidification (OA)—induces a series of geochemical reactions that increases seawater hydrogen  
46 ion concentration [H<sup>+</sup>] (lowering pH) while concomitantly reducing the ocean's overall buffering

47 capacity by reducing the carbonate concentration [ $\text{CO}_3^{2-}$ ] (Caldeira and Wickett, 2003; Orr et al.,  
48 2005). Due to more dynamic natural physical and chemical processes in the coastal ocean, a  
49 differentiation exists between open-ocean acidification and nearshore coastal acidification.  
50 Open-ocean acidification of surface waters is predominately a function of equilibration with  
51 atmospheric  $p\text{CO}_2$ , thus increasing on yearly and decadal timescales as anthropogenic sources of  
52  $\text{CO}_2$  production continue (Hofmann et al., 2011; Orr et al., 2005). Coastal acidification, however,  
53 can manifest on short time and space scales driven by riverine input and its chemical constituents  
54 (e.g., organic carbon, nutrients, and organic alkalinity), community metabolism and organization,  
55 tidal cycles, upwelling, and groundwater input (Duarte et al., 2013; Sunda and Cai, 2012;  
56 Waldbusser and Salisbury, 2014), all of which can act in conjunction with increasing  
57 atmospheric  $\text{CO}_2$ , leading to more frequent, intense, and longer-lasting acidification events  
58 (Hales et al., 2016; Harris et al., 2013). In the face of rapidly changing coastal conditions,  
59 tracking and quantifying the progression of OA requires precise and accurate measurements of  
60 carbonate chemistry over long periods of time; these can be achieved by appropriately  
61 constraining the carbonate system by measuring at least two of the system's parameters: total  
62 dissolved inorganic carbon ( $\text{TCO}_2$ ), total alkalinity (TA), pH, and the partial pressure of  $\text{CO}_2$   
63 ( $p\text{CO}_2$ ). Despite the marked increase in OA research over the past decade (Riebesell and Gattuso,  
64 2015; Rudd, 2017), nearshore monitoring efforts—particularly in estuarine waters—have been  
65 slow to ramp up, however, efforts are beginning to intensify as technological advancements are  
66 made (Feely et al., 2010, 2016; Hales et al., 2016; Harris et al., 2013; Newton et al., 2012;  
67 Waldbusser and Salisbury, 2014; Chan et al., 2017).

68  
69 Acidification of Alaskan coastal waters is predicted to progress rapidly relative to other  
70 regions within the next 50 years, and negatively impact the social-ecological structure of Alaskan  
71 marine resources by disrupting the Alaska Native subsistence and commercial fisheries (Ekstrom  
72 et al., 2015; Mathis et al., 2015b). The ocean waters present along the Alaskan coastline  
73 experience chemical and physical drivers of seawater chemistry that are unique to this region.  
74 The low seawater temperatures inherently have higher concentrations of dissolved  $\text{CO}_2$ , and  
75 chemical and physical oceanic processes unique to Alaskan waters such as sea ice melt, glacial  
76 discharge, and benthic pelagic coupling across shallow shelves are likely to exacerbate  
77 acidification in this region (Evans et al., 2014; Mathis et al., 2011a, 2011b, 2012). Recently, an  
78 OA monitoring initiative has been setup by the Alaska Ocean Observing Network (AOOS) to  
79 track and provide accessible material dedicated to acidification research in Alaskan waters  
80 (<http://www.aos.org/alaska-ocean-acidification-network>). Along the Pacific coast of Alaska, a  
81 robust benchtop system known as a Burke-o-Lator (BoL), which measures  $\text{TCO}_2$  and  $p\text{CO}_2$   
82 either continuously in a flow-through environment or from discrete seawater samples (Bandstra  
83 et al., 2006; Barton et al., 2012; Hales et al., 2016) has been installed in several locations,  
84 including the OceansAlaska Shellfish Hatchery in Ketchikan, the Alutiiq Pride Shellfish  
85 Hatchery in Seward (Evans et al., 2015), and at the Sitka Tribe of Alaska Environmental  
86 Research Center (real-time data from Alaskan and other BoLs:  
87 [http://www.ipacoa.org/Explorer?action=oiw:fixed\\_platform](http://www.ipacoa.org/Explorer?action=oiw:fixed_platform)). Nominal analytical uncertainty for  
88  $\text{TCO}_2$  determinations from this system is 0.2% based on the reproducibility of sample and  
89 certified reference material (CRM; provided by A. Dickson analyses). For  $p\text{CO}_2$  determinations,  
90 analytical uncertainty is 1.5% based on the inaccuracy of calculated CRM alkalinity relative to  
91 the certified value. While the BoL has significant advantages for achieving robust OA  
92 measurements in nearshore waters, the physical constraints of a benchtop system limit the spatial

93 dimension of which carbonate chemistry parameters can be measured. One potential resolution  
94 to diminish the gap in coverage of OA monitoring is to utilize autonomous pH sensors, which are  
95 far more versatile in their ability to monitor hard-to-reach areas.  
96

97 Recent assessments regarding OA monitoring efforts have specifically highlighted the  
98 benefits of accessibility by the commercially produced SeaFET<sup>TM</sup> pH sensor utilizing Honeywell  
99 Durafet technology (Martz et al., 2015). The SeaFET<sup>TM</sup> was originally developed at the  
100 Monterey Bay Aquarium Research Institute (Martz et al., 2010), but since has been  
101 manufactured and distributed by Satlantic (<http://www.satlantic.com>), which is now incorporated  
102 into Sea-Bird Scientific (<http://www.seabird.com>). The partnership between MBARI, Scripps  
103 Institute of Oceanography, and Satlantic led the way for commercial availability of the  
104 SeaFET<sup>TM</sup>, providing a ready-to-deploy-factory calibration, quick start manual, and user-friendly  
105 interface. The first generation of SeaFETs<sup>TM</sup> (not distributed by Sea-Bird, but by Dr. Todd Martz  
106 at Scripps Institute of Oceanography) have been deployed in numerous field studies and were  
107 heavily scrutinized in order to provide robust best practices for appropriate calibration and  
108 deployment procedures (Bresnahan et al., 2014; Hofmann et al., 2011; Kapsenberg and  
109 Hofmann, 2016; Martz et al., 2010; Matson et al., 2011; Yu et al., 2011). More recent studies  
110 have expanded the scope of SeaFET<sup>TM</sup> accuracy, inter-sensor variability, operator experience,  
111 and multi-point calibration techniques (Gonski et al., 2018; Johnson et al., 2017; Kapsenberg et  
112 al., 2017; McLaughlin et al., 2017). Given the multitude of information regarding SeaFET<sup>TM</sup>  
113 performance, coalescing all the potential sources of uncertainty in measurements (e.g., inter-  
114 sensor variability and calibration method) can be logistically challenging for non-experienced  
115 oceanographers who now have access to SeaFETs<sup>TM</sup> distributed by Sea-Bird.  
116

117 In this study, we aimed to take advantage of two distinct coastal settings in order to  
118 deploy and evaluate the commercially available Sea-Bird SeaFET<sup>TM</sup>, and the potential  
119 uncertainties that can arise with time series pH<sub>t</sub> (total scale) measurements. For this evaluation,  
120 SeaFETs<sup>TM</sup> were co-deployed side-by-side to quantify inter-sensor variability, discrepancies  
121 were examined between factory calibration, *in situ* single-point calibration, and *in situ* multi-  
122 point calibration pH<sub>t</sub> values, and anomalous data associated with sensor conditioning times were  
123 detailed and considered as potential sources of measurement inaccuracies. All evaluations of  
124 SeaFET<sup>TM</sup> performance were under non-controlled source water conditions (i.e., non-  
125 manipulated seawater) or by *in situ* deployments. Three pH sensors were deployed in coastal  
126 waters and were subjected to tidal influences and freshwater input, while a fourth was compared  
127 to pH<sub>t</sub> values derived from measurements obtained by a BoL. Finally, a spectral analysis of the  
128 quality-controlled data was performed in order to identify the driving mechanism of pH<sub>t</sub>  
129 variability between these divergent sites and consider possible un-accounted for calibration  
130 errors that could occur in dynamic settings that might not be resolved using a specific calibration  
131 method.  
132

## 133 **2 Methods**

### 134 **2.1 Apparatus: SeaFET<sup>TM</sup>**

135 The commercially available Sea-Bird SeaFET<sup>TM</sup> has retained the basic design of the original  
136 sensor developed at MBARI (Martz et al., 2010). This pH sensor utilizes the ion sensitive field  
137  
138

139 effect transistor (ISFET) technology, and is outfitted with an internal Honeywell Durafet and an  
 140 external solid-state chloride selective electrode (Cl-ISE) along with an internal thermistor, which  
 141 derives temperature using the Steinhart and Hart (1968) equation. The internal reference  
 142 electrode is intrinsically insensitive to salinity over a tested range from 30 – 36 (Bresnahan et al.,  
 143 2014), with recent work even suggesting near-ideal Nernstian response to salinity as low as ~9.0  
 144 (Gonski et al., 2018). This is in converse to the chloride sensitive external electrode, which is  
 145 salinity dependent. Both electrodes demonstrate exceptional stability over a range of moderate  
 146 salinity (30 – 36) and broad temperature (-1 to 35 °C) (Bresnahan et al., 2014; Kapsenberg et al.,  
 147 2015; Martz et al., 2014, 2010). The range of salinity sensitivity for the external electrode has  
 148 even been extended down to 20, where it displays a near-ideal Nernst slope (Takeshita et al.,  
 149 2014). Sea-Bird suggests that the external reference electrode provides the more accurate and  
 150 stable pH<sub>t</sub> measurement given that chloride concentration can be precisely determined from  
 151 accurate salinity measurements. This is in agreement with previous research demonstrating that  
 152 the external electrode has a more robust stability (Martz et al., 2010). In dynamic nearshore  
 153 environments (e.g., estuaries with strong tidal and riverine fluxes), however, the pH<sub>t</sub> derived  
 154 from the internal electrode is recommended (Sea-Bird Scientific’s Branham, C., pers. comm.)  
 155 despite the potential of thermodynamic hysteresis (Martz et al., 2010). Bresnahan et al. (2014)  
 156 demonstrated that the internal electrode is of the highest quality and under most scenarios  
 157 remains nearly as stable as the external electrode—this was further corroborated by Gonski et al.  
 158 (2018) with SeapHOx deployments in the Murderkill estuary, Delaware.

## 159 160 2.2 Calibration

161 Currently, three different calibration methods are present for the SeaFET™: a factory pre-  
 162 deployment single-point calibration, *in situ* single-point calibration, and an *in situ* multi-point  
 163 calibration (Bresnahan et al., 2014; Gonski et al., 2018). To properly calculate pH<sub>t</sub> from sensor  
 164 voltage readings, an appropriate calibration coefficient is required. The applied calibration  
 165 coefficients from the factory are a single-point, pre-deployment calibration. Given that a  
 166 conditioning period is required for the sensor (Bresnahan et al., 2014), these coefficients are  
 167 likely not adequate once the sensor becomes conditioned to the environment to which it is  
 168 deployed. For the internal electrode, the new calibration coefficient  $k_{0i}$  can be determined as

$$171 \quad k_{0i} = -S_{nernst} * pH_t + V_{int} - k_{2i} * T, \quad (1)$$

172 and  $k_{0e}$  for the external electrode

$$173 \quad k_{0e} = V_{ext} - pH_t + \log\left(1 + \frac{S_T}{K_s}\right) - 2 * \log(\gamma_{HCl}) - \log(Cl_T) * S_{nernst} + k_{2e} * T \quad (2)$$

174 where  $V_{int|ext}$  is the voltage from the electrode and  $k_{2i|e}$  is the temperature coefficient ( $dE^*/dT$ )  
 175 applied to all SeaFETs™ (Martz et al., 2010). For detailed definitions of  $S_{nernst}$  and the salinity  
 176 dependent constants  $\gamma_{HCl}$  (HCl activity coefficient),  $Cl_T$  (total chloride),  $S_T$  (total sulfate), and the  
 177 HSO<sub>4</sub><sup>-</sup> dissociation constant  $K_s$  (Dickson et al., 2007; Khoo et al., 1977) in equations 1 and 2, we  
 178 refer readers to Martz et al. (2010), Bresnahan et al. (2014), and Sea-Bird Scientific SeaFET™  
 179 Product Manual 2.0.0. In the literature, SeaFET™ calibration coefficients have been denoted as  
 180  $E^*_{int}$  and  $E^*_{ext}$  (Martz et al. 2010, Bresnahan et al. 2014), however, for the purpose of this  
 181 182 183

184 evaluation—which specifically examines Sea-Bird SeaFETs™—the adoption of  $k_0$  and  $k_2$  is in  
185 accordance with the preferred nomenclature from the manufacturer.  
186

187 Unlike the factory pre-deployment single-point calibration, the *in situ* single-point  
188 calibration occurs after the sensor has been deployed in the field. At the operator’s discretion, a  
189 discrete sample will be collected in direct proximity to the deployed sensor at the same time that  
190 it is actively making a measurement, and then measured for  $\text{pH}_t$  at *in situ* temperature and  
191 salinity. The known  $\text{pH}_t$  would then be used in the above equations as the “ $\text{pH}_t$ ” variable. Similar  
192 to the single-point *in situ* calibration, the multi-point calibration derives a series of calibration  
193 coefficients over a short period of time that is long enough to capture environment variability  
194 such as tidal fluxes, and then a single calibration coefficient is averaged. Both single-point  
195 calibration methods—pre-deployment and *in situ*—appear to be suitable for fairly static  
196 environmental conditions, whereas the multi-point *in situ* calibration is best suited for dynamic  
197 nearshore environments (Bresnahan et al., 2014; Gonski et al., 2018).  
198

### 199 **2.3 SeaFET™ conditioning: test tank deployments** 200

201 A series of three separate test tank deployments for three SeaFETs™<sub>395, 396, 397</sub> were conducted in  
202 order to determine the conditioning period for each sensor. Initial sensor deployments took place  
203 in October 2016 at the Alutiiq Pride Shellfish Hatchery (APSH) in Seward, Alaska. Sensors were  
204 deployed for a duration of 72 hours in a flow-through 60 L tank where seawater taken from a  
205 depth of ~75 m in Resurrection Bay was sand-filtered, UV treated, and finally run through a 5  
206  $\mu\text{m}$  mesh. All three sensors were programmed with identical sampling settings (Table 1). The  
207 onboard internal thermistor was used to calculate temperature, and measurements of seawater  
208 salinity incoming to the hatchery were collected by a Sea-Bird Scientific SBE 45 MicroTSG  
209 Thermosalinograph that is paired with the BoL and are available on the Alaska Ocean Observing  
210 System (<http://portal.aos.org/real-time-sensors.php#map>). Factory calibration coefficients for  
211 the internal ( $k_{0i}$ ,  $k_{2i}$ ) and external ( $k_{0e}$ ,  $k_{2e}$ ) electrodes were retained when processing raw voltage  
212 data.  
213

214 A second tank deployment for the same three sensors<sub>395, 396, 397</sub> were deployed at the  
215 University of Alaska, Fairbanks, in the Ocean Acidification Research Center (OARC). Seawater  
216 collected from the APSH was delivered to the OARC test tank, ~370 L in a half-filled tank.  
217 Seawater in the tank was circulated continuously and covered to aid in the prevention of  
218 evaporation and photosynthesis. A co-deployed Sea-Bird SBE 16plusV2 SeaCAT (recently  
219 serviced by Sea-Bird) collected temperature and salinity readings every 5 minutes. Sensors<sub>395,</sub>  
220<sub>396, 397</sub> were deployed for a duration of nine days in continuous operation mode which forgoes the  
221 ability to set frames per burst; average number of reads was identical between all sensors (Table  
222 1). From 1 – 4 November 2016, duplicate discrete bottle samples were collected in 250 ml glass  
223 bottles with screw caps at ~00:00 and 17:00 UTC per day. Bottle samples were preserved with  
224 20  $\mu\text{l}$  of saturated  $\text{HgCl}_2$  and processed at a later date for  $\text{TCO}_2$  and TA with a VINDTA 3C  
225 (Versatile Instrument for the Determination of total inorganic carbon and titration alkalinity).  
226 The VINDTA 3C has an uncertainty typically near 0.05% (Mathis et al., 2014, 2015a). Bottle  
227 sample  $\text{pH}_t$  was calculated using CO2SYS with known  $\text{TCO}_2$  and TA using the constants  
228 provided by Uppström (1974) and Lueker et al. (2000); derived  $\text{pH}_t$  was then compared against  
229 sensor  $\text{pH}_t$  to test the accuracy of both internal and external electrodes, assuming the discrete

230 bottle samples were the “true pH” of the seawater. Upon recovery, all sensors<sub>395, 396, 397</sub> were  
231 placed into polled mode and stored with wet caps filled with tris buffer (salinity 34, pH 8.09 at  
232 room temperature, 25 °C). Again, the factory calibration coefficients for the internal and external  
233 electrodes were retained when raw voltage was processed. Since the SBE 16plusV2 sampled  
234 every 5 min, salinity and temperature measured by the SBE at each 5-minute point was repeated  
235 for the following 4 minutes in order to calculate continuous minute readings by sensors<sub>395, 396,</sub>  
236 <sub>397</sub>.

237  
238 A final test tank deployment of sensors<sub>395, 396, 397</sub> at OARC was conducted after an  
239 assumed adequate conditioning period of nine days (first OARC deployment). All three sensors  
240 <sub>395, 396, 397</sub> had been set to polled mode after the end of the previous deployment and, therefore,  
241 were sleeping for 83 days until this final seven day deployment. The sampling settings were  
242 identical to the first OARC deployment for all three sensors<sub>395, 396, 397</sub> (Table 1). Similar to the  
243 previous OARC tank deployment, a co-deployed Sea-Bird SBE 16plusV2 SeaCAT collected  
244 temperature and salinity mirroring the sensor sampling interval of 3 hrs.

245  
246 The internal thermistor of each sensor<sub>395, 396, 397</sub> was tested for accuracy by comparing its  
247 derived *in situ* temperature to that collected by the Sea-Bird SBE 16plusV2 during the test tank  
248 deployments. The temperature difference between the internal thermistor and the SBE 16plusV2  
249 was used to calculate the average and maximum discrepancy between the two temperature  
250 readings. The temperature discrepancy was then applied to a combination of TA: TCO<sub>2</sub> ratios  
251 over a range of salinity (20 – 35) in CO<sub>2</sub>SYN (constants: Uppström, 1974; Lueker et al., 2000),  
252 which produced two different pH<sub>t</sub> values. The difference between these two pH<sub>t</sub> values were,  
253 therefore, concluded to be a result of the temperature discrepancy.

## 254 255 **2.4 SeaFET™ performance: field deployments**

256  
257 In late boreal winter 2017—32 days post final tank deployment—SeaFET™<sub>397</sub> was deployed at  
258 the APSH and the two remaining sensors (SeaFET™<sub>395, 396</sub>) in Kasitsna Bay within greater  
259 Kachemak Bay, Alaska (Fig. 1). At the APSH (60° 5' 55.59"N, 149° 26' 39.80"W), incoming  
260 seawater from Resurrection Bay at a depth of 75 m is split before running through a series of  
261 hatchery water filters so that an unfiltered line is run directly to the BoL. The incoming line to  
262 the BoL was then split to feed an ~11.5 L conical tank housing sensor<sub>397</sub> fit with the copper bio-  
263 fouling guard; tank residence time was ~7.5 min. The sensor<sub>397</sub> at this location was deployed on  
264 6 March 2017 with a robust sampling setting (Table 1). Two calibration methods were applied to  
265 this sensor<sub>397</sub>, an *in situ* single-point calibration and an *in situ* multi-point calibration. Both  
266 calibrations were performed 50 days after deployment on 25 April 2017 once the BoL had  
267 completed service maintenance. The single-point *in situ* calibration was taken during midday tide  
268 transition in Resurrection Bay, while the multi-point *in situ* approach used five (sensor sampling  
269 3 h intervals) time points spanning an entire tidal cycle. The single-point *in situ* calibration was  
270 used to derive  $k_{0i}$  for the internal electrode (eq.1) and  $k_{0e}$  for the external electrode (eq. 2). The  
271 multi-point *in situ* calibration followed the same formulations with the difference being the final  
272 calibration coefficient calculated was the average of the five independently calculated calibration  
273 coefficients. Three final pH<sub>t</sub> values for the sensor<sub>397</sub> were, therefore, calculated based upon the  
274 different calibration coefficients (factory, single-point and multi-point *in situ* calibration) and  
275 compared against the pH<sub>t</sub> determined from continuous  $p\text{CO}_2$  measurements by the BoL and

276 derived TA (TA-S equation, Evans et al. 2015) using CO2SYS with constants provided by  
277 Uppström (1974) and Lueker et al. (2000).  $pH_t$  uncertainty from the BoL using this combination  
278 of measured and derived parameters is 0.007 units based on propagating the error of the BoL  
279  $pCO_2$  uncertainty reported above with the RMSE ( $17 \mu mol kg^{-1}$ ) of the regional TA-S  
280 relationship (Orr, et al., *in prep*).

281  
282 Inter-sensor variability was examined between two SeaFETs<sup>TM</sup><sub>395, 396</sub> deployed off the  
283 pier at the Kasitsna Bay laboratory in Kachemak Bay ( $59^{\circ}28' 6.71''N$ ,  $151^{\circ}33'11.12''W$ )  $\sim 1.5$  m  
284 from the bottom: depth at this location fluctuates between  $\sim 7.5 - 16.8$  m (Fig. 1). On 18 March  
285 2017—44 days post final tank deployment—the sensors<sub>395, 396</sub> were attached to the pier piling  
286 directly beside one another on a single mooring frame. Both sensors were wrapped with pipe  
287 tape to minimize biofouling and fit with their respective copper biofouling guards which had a  
288 tributyltin plug attached to the inside of the guard. The sampling settings for both sensors<sub>395, 396</sub>  
289 were identical to the one at the APSH (Table 1). Five discrete reference samples were taken in  
290 duplicate: one sample on day of deployment (UTC: 3-18-17, 18:00), two samples 1-day post-  
291 deployment (UTC: 19 March 2017, 03:00 and 15:00), and two samples 2- and 1-day pre-  
292 recovery of the sensors<sub>395, 396</sub> (UTC 3 June 2017, 03:00; 6 June 2017, 03:00). Reference samples  
293 were collected within 30 s of the instrument sampling time period via a diver's hand Niskin,  
294 measured for temperature and salinity with a YSI 3100 conductivity instrument, stored in 250 ml  
295 glass bottles with screw caps, poisoned with 100  $\mu l$  of saturated  $HgCl_2$ , and secured with teflon  
296 tape around the bottleneck threading and Parafilm wrapped on the outside of the cap. Calibration  
297 samples were processed for  $TCO_2$  and TA with a VINDTA 3C and  $pH_t$  calculated using  
298 CO2SYS with the constants provided by Uppström (1974) and Lueker et al. (2000). Salinity  
299 measurements collected by the Kachemak Bay National Estuarine Research Reserve data sonde,  
300 10 km SE of the deployed sensors ( $59^{\circ}26' 26.87''N$ ,  $151^{\circ}43'15.21''W$ ), were used along with the  
301 sensor's internal thermistor readings to calculate  $pH_t$  from the raw voltage data in order to  
302 capture representative environmental conditions providing relevance for the  $pH_t$  time series in  
303 this location. A static salinity of 32 was also used for all calculations of  $pH_t$  as an assessment of  
304 variability due to salinity measured from a data sonde 10 km away. A total of four different  $pH_t$   
305 values for both sensors<sub>395, 396</sub> were calculated based on calibration method (factory pre-  
306 deployment single-point calibration and the *in situ* single-point) and conditioning: either  
307 conditioned or non-conditioned to the environment. All calculated  $pH_t$  values from the sensors  
308 <sub>395, 396</sub> were then compared against the remaining discrete reference bottle samples not used for  
309 calibration. This was done in order to examine the accuracy and inter-sensor variability  
310 difference between conditioned and non-conditioned to the environment electrodes. Because the  
311 Kachemak Bay data sonde was located 10 km from the deployed sensors<sub>395, 396</sub>, the measured  
312 temperature and salinity from the discrete reference samples were used to determine  $pH_t$  for the  
313 internal and external electrodes at those specific time points. That is, sensor accuracy for these  
314 two sensors<sub>395, 396</sub> was only assessed with accurate temperature and salinity values determined  
315 from the discrete bottle samples.

316  
317 A fourth SeaFET<sup>TM</sup><sub>268</sub> operated by the Hakai Institute was deployed on Environment  
318 Canada's Sentry Shoal weather buoy in the Northern Strait of Georgia, BC, Canada:  $49^{\circ} 54'$   
319  $24.00''N$ ,  $124^{\circ} 59' 5.99''W$  (Fig.1). The Sentry Shoal mooring site is in a water depth of 15 m and  
320 the sensor<sub>268</sub> was affixed at a depth of 1 m. A pre-deployment bucket test was conducted for 24  
321 h at a sampling interval of 30 min with an average of 10 samples per frame and 30 frames per

322 burst from 28 – 29 June 2016. Sensor<sub>268</sub> was outfitted with a copper housing guard and wrapped  
 323 with copper tape. Sensor<sub>268</sub> underwent two separate deployments, an initial deployment, and a  
 324 redeployment (6 July and 27 August 2016) that occurred after the sensor was retrieved for  
 325 cleaning and maintenance. Two separate calibration samples (taken in triplicate) were taken in  
 326 accordance with each deployment, and occurred 13 and 7 days after each deployment (19 July  
 327 and 2 September 2016). For each deployment, sensor<sub>268</sub> settings were similar to the others at the  
 328 APSH and in Kasitsna Bay (Table 1). All calibration samples were taken in triplicate at a depth  
 329 of 1 m via CTD and Niskin bottle castings and collected in 350 ml amber glass bottles with  
 330 polyurethane-lined crimp-sealed metal caps and poisoned with 200 µl of saturated HgCl<sub>2</sub>, and  
 331 then processed for TCO<sub>2</sub> and pCO<sub>2</sub> with a BoL at the Hakai Institute’s Quadra Island Field  
 332 Station. The measured values were used to derive pH<sub>t</sub> using CO2SYS with the constants  
 333 provided by (Uppström, 1974) and (Lueker et al., 2000) in order to perform a single-point *in situ*  
 334 calibration. Uncertainty in pH determinations from BoL pCO<sub>2</sub> and TCO<sub>2</sub> measurements was  
 335 0.006 units. After sensor<sub>268</sub> deployment and calibration, a total of three, triplicate, reference  
 336 sample sets were taken and processed for pH<sub>t</sub> following the procedure used for calibration  
 337 samples, then compared against sensor pH<sub>t</sub>.

## 338 2.5 Quantifying pH<sub>t</sub> and intrinsic sensor uncertainties

339  
 340 Calculating pH<sub>t</sub> from the SeaFET’s<sup>TM</sup> raw voltage reading is dependent on temperature, salinity  
 341 and an ideal 100% Nernstian response. The software application SeaFETcom permits the  
 342 operator to automatically calculate pH<sub>t</sub> by assigning the calibration coefficient either written to  
 343 the sensor’s header file or the one provided on the CD-ROM (these should be identical).  
 344 Determination of final pH<sub>t</sub> values from the first test tank deployment at the APSH were  
 345 calculated by two different operators and two sources for the factory pre-deployment single-point  
 346 calibration coefficients: header file and CD-ROM disc file. Aside from that exception, all other  
 347 final pH<sub>t</sub> values for the internal and external electrodes were calculated with the Mathworks  
 348 software MATLAB (V. 2016a) and Microsoft excel (v. 2016) using the following equations for  
 349 the internal electrode

$$350 \quad \quad \quad 351 \quad \quad \quad pH_{int} = \frac{V_{int} - k_{0i} - k_{2i} * T}{S_{nernst}}, \quad (3)$$

352 and the external electrode

$$353 \quad \quad \quad pH_{ext} = \frac{V_{ext} - k_{0e} - k_{2e} * T}{S_{nernst}} + \log(Cl_T) + 2 * \log(\gamma_{HCl}) - \log\left(1 + \frac{S_T}{K_s}\right) \quad (4)$$

354 where V<sub>int/ext</sub> is the voltage from the electrode and k<sub>2i/e</sub> is the temperature coefficient (dE\*/dT)  
 355 applied to all SeaFETs<sup>TM</sup> (Martz et al. 2010). Again, for detailed definitions of S<sub>nernst</sub> and the  
 356 salinity dependent constants γ<sub>HCl</sub> (HCl activity coefficient), Cl<sub>T</sub> (total chloride), S<sub>T</sub> (total sulfate),  
 357 and the HSO<sub>4</sub><sup>-</sup> dissociation constant K<sub>s</sub> (Khoo et al. 1977, Dickson et al. 2007) in equations 3 and  
 358 4, we refer readers to Martz et al. (2010), Bresnahan et al. (2014), and Sea-Bird Scientific  
 359 SeaFET<sup>TM</sup> Product Manual 2.0.0.

### 360 2.5.1 Sensor uncertainty

361 The overall accuracy of every SeaFET<sup>TM</sup> sensor was evaluated by quantifying all sources of



362 potential uncertainty when calculating a final  $\text{pH}_t$  from the sensor (Table 2). The  $\text{pH}_t$  uncertainty  
363 introduced by calibration method was calculated as the absolute difference between the “true  
364  $\text{pH}_t$ ” and the final sensor  $\text{pH}_t$  derived from either factory calibration, the single-point *in situ*  
365 calibration, or multi-point *in situ* calibration. The “true  $\text{pH}_t$ ” was calculated using CO2SYS  
366 dissociation constants by Lueker et al., (2000) and Uppström, (1974) with measured  $\text{TCO}_2$  and  
367 TA via the VINDTA 3C,  $\text{TCO}_2$  and  $p\text{CO}_2$  measured by the BoL for discrete samples (e.g.,  
368 sensor<sub>268</sub>), and  $p\text{CO}_2$  and TA (TA-S equation, Evans et al. 2015) for continuous samples  
369 (sensor<sub>397</sub>). A one-way analysis of variance (ANOVA) and the root mean square error (RMSE)  
370 were run and calculated in order to compare the  $\text{pH}_t$  values from both electrodes on sensor<sub>397</sub>  
371 across calibration methods against the  $\text{pH}_t$  values from the BoL. The BoL at the APSH sampled  
372 every 5 min which produced 256 comparable sample points with a time alignment disparity that  
373 ranged from 0 – 120 s against sensor<sub>397</sub>. The potential  $\text{pH}_t$  uncertainty based on the thermistor  
374 was calculated by using the absolute difference between the thermistor derived temperature and  
375 that measured by the SBE 16plusV2 ( $T_{\text{diff}}$ ) from the OARC test tank deployments and the  
376 Kasitsna Bay sensors<sub>395, 396</sub> against the Seldovia data sonde 10 km away. Finally, an average  
377 inter-sensor variability uncertainty term was calculated as the difference between the two  
378 sensors<sub>395, 396</sub> deployed side-by-side in Kasitsna Bay after a single-point *in situ* calibration was  
379 performed. All uncertainty terms were calculated and collated based on our evaluations from the  
380 Alaska deployed sensors<sub>395, 396, 397</sub>, while sensor<sub>268</sub> deployed at Sentry Shoal was only included  
381 when determining the accuracy uncertainty term. Due to the disparity between reference samples  
382 for the Kasitsna Bay sensors<sub>395, 396</sub> and the Sentry Shoal sensor<sub>268</sub> (two discrete reference  
383 samples) to that of sensor<sub>397</sub> at the APSH (256 reference samples), only the average calculated  
384 difference (SeaFET™  $\text{pH}_t$  – “true  $\text{pH}_t$ ”) for each calibration method and electrode was used from  
385 the APSH sensor<sub>397</sub> and then collated with the other reference points from the Kasitsna Bay and  
386 Sentry Shoal sensors<sub>395, 396, 268</sub>.

## 387 2.5.2 $\text{pH}_t$ time series analysis

388  
389 Final time series analysis was examined in the time and frequency domain using the Mathworks  
390 software MATLAB (V. 2016a). Power spectral density was determined via Welch’s method  
391 using the pwelch function in MATLAB. Time series data was resampled and linearly  
392 interpolated in order to compensate for the missing data points that occurred when sensors  
393 arbitrarily stopped sampling.

## 394 3 Results

### 395 3.1 Test tank and field conditions

396  
397  
398  
399 Finalized (i.e., calibrated)  $\text{pH}_t$  values from the first test tank deployment produced two different  
400 values, of which each was dependent on whether the calibration coefficient from the header file  
401 or the disc file was selected, the result was a difference of ~0.0011 units for both the internal and  
402 external electrodes. Because sensors were stored in tris buffer that lacked the addition of bromide  
403 between tank deployments and before field deployments, an environmental conditioning period  
404 was required for each of the Alaska sensors<sub>395, 396, 397</sub> once submerged in their respective field  
405 sites. Thus, any determination of SeaFET™  $\text{pH}_t$  accuracy and conditioning period from tank

406 deployments were inconclusive and will not be considered henceforth. No sensors<sub>395, 396, 397, 268</sub>  
407 displayed signs of biofouling or low battery power upon recovery.

408  
409 Sensor<sub>397</sub> deployed in parallel with the BoL at the APSH experienced a tank failure on 8  
410 April 2017 resulting in the sensor's emergence for 24 h. In addition, missing temperature and  
411 salinity values resulted in gaps of pH<sub>t</sub> measurements over the entire deployment. The BoL  
412 experienced flow control issues when initial deployment occurred on 6 March 2017 and was not  
413 online until 18 April 2017 but, then, operated nearly consistently until 24 May 2017. All pH<sub>t</sub> and  
414 temperature comparisons were, therefore, made beginning on 18 April 2017.

415  
416 Due to the *in situ* environmental conditioning period of the Kasitsna Bay sensors<sub>395, 396</sub>,  
417 calibration was performed using the initial reference sample collected on 18 March 2017, 03:00  
418 UTC and again with the reference sample collected on 3 June 2017, 03:00 UTC. Due to high  
419 variance between duplicate reference samples (SD: 0.08 pH<sub>t</sub>) on 19 March 2017, 15:00 UTC,  
420 this reference was discarded and not used for comparison or calibration. The Sentry Shoal  
421 sensor<sub>268</sub> underwent one maintenance and cleaning procedure, including a battery change,  
422 during the ~5-month deployment (Table 1). One calibration sample (19 July 2016) and one  
423 reference sample (9 November 2016) were averaged from duplicate rather than triplicate  
424 replicates due to large variance from one of the replicate samples. The reference sample taken on  
425 23 August 2016, 17:00 UTC was discarded as temperature and salinity data were missing and  
426 sensor<sub>268</sub> pH<sub>t</sub> could not be calculated. The final reference sample (UTC: 9 November 2016,  
427 17:05) was taken 5 min after sensor<sub>268</sub> sampled on 9 November 2016, 17:00 UTC.

### 428 429 **3.2 Thermistor response: test tank deployment**

430  
431 The internal thermistor amongst the sensors<sub>395, 396, 397</sub> had a difference of less than 0.2 °C over  
432 the entirety of the second and third tank deployments. All thermistor derived temperature values  
433 had good alignment with the SBE 16plusV2 temperature, and consistently recorded a slightly  
434 higher temperature. The discrepancy between the thermistor temperature and SBE16plusV2 was  
435 minimal, and reached a maximum of 0.378 (logged by sensor<sub>395</sub>) during any time over all tank  
436 deployments. The average discrepancy, however, was ~0.21 °C when averaging across all  
437 sensors<sub>395, 396, 397</sub> and all times—resulting in a 0.003 pH uncertainty.

### 438 439 **3.3 Field performance**

440  
441 Sensor<sub>397</sub> deployed alongside the BoL appeared stable throughout its entire deployment and  
442 tracked the pH<sub>t</sub> derived from the BoL well (Fig. 2). Errant spikes were present from both  
443 electrodes throughout periods before 18 April 2017, which were a result of plumbing changes  
444 that occurred to the APSH incoming seawater. On 10 April 2017 the internal thermistor, BoL  
445 temp, and BoL salinity fluctuated by 3 °C and 14, respectively, over a 12 h period. These  
446 anomalies were removed from analysis. Salinity remained relatively stable throughout the rest of  
447 the deployment and ranged from 30.0 – 32.1. The pH<sub>t</sub> uncertainty decreased, and the accuracy of  
448 the sensor's<sub>397</sub> internal electrode improved once the *in situ* single-point and multi-point  
449 calibrations were performed with a RMSE decreasing from 0.5455 pH<sub>t</sub> units under factory  
450 calibration, 0.0361 pH<sub>t</sub> units for *in situ* single-point calibration and 0.0273 pH<sub>t</sub> units for the *in*  
451 *situ* multi-point calibration. The external electrode also improved accuracy with *in situ* single-

452 point and multi-point calibrations with an RMSE of 0.1077 under factory calibration, 0.0390 for  
453 *in situ* single-point calibration and 0.0388 for the *in situ* multi-point calibration (Fig. 2). There  
454 was a significant difference in the reduction of the  $\text{pH}_t$  uncertainty for both the internal and  
455 external electrodes when utilizing *in situ* single-point and multi-point calibration coefficients  
456 compared to the factory calibration coefficients (Table 3). In addition, there was a significant  
457 decrease in the  $\text{pH}_t$  uncertainty when using the *in situ* multi-point calibration coefficients rather  
458 than the *in situ* single-point method for the internal electrode, but not for the external electrode  
459 (Table 3). The  $\text{pH}_t$  uncertainty of the internal electrode decreased from 0.0294 units with an *in*  
460 *situ* single-point calibration to 0.0224 units after an *in situ* multi-point calibration. It should be  
461 noted that the time alignment disparity which ranged from 0 – 120 s is not considered a  
462 significant source of discrepancy as only 4 sample points out of the 256 comparable points were  
463 > 0.03 units (i.e., only 4 comparable points greater than the average  $\text{pH}_t$  uncertainty found after  
464 calibration) between any one 5 min sample taken by the BoL. The internal thermistor of  
465 sensor<sub>397</sub> tracked the recorded BoL temperature trend fairly (Fig. 3), but had a greater magnitude  
466 discrepancy than its test tank deployment (~0.21 °C). On average, the thermistor temperature had  
467 an absolute difference of 2.83 °C (SD 0.35) from 18 April 2017 – 6 June 2017, which would  
468 result in a  $\text{pH}_t$  uncertainty of ~0.044 units. Sensor<sub>397</sub> was not fully submerged in the conical tank  
469 leaving the top portion susceptible to air temperature fluctuations which could have affected the  
470 thermistor readings.

471  
472 The sensors<sub>395, 396</sub> in Kasitsna Bay improved their accuracy after an *in situ* single-point  
473 calibration was performed (Fig. 4), however, this was only the case when sensors were not  
474 conditioned as calibration performed after the conditioning period reduced accuracy (Fig. 5)  
475 when comparing against discrete reference samples. It should be noted that only the  $\text{pH}_t$  recorded  
476 by both sensors<sub>395, 396</sub> at times of the reference samples had precise salinity and temperature  
477 (temperature and salinity recorded with reference sample rather than thermistor derived  
478 temperature) measurements as all other measurements were calculated from salinity measured by  
479 the data sonde 10 km away, and with temperature derived from the onboard thermistor. The  $\text{pH}_t$   
480 recorded by the external electrode at a fixed salinity displayed little to no variance relative to  $\text{pH}_t$   
481 calculated with data sonde salinity (< 0.02  $\text{pH}_t$  difference: average whether conditioned or non-  
482 conditioned to environment). The average  $\text{pH}_t$  uncertainty from both sensors<sub>395, 396</sub> reduced by  
483 approximately half for the internal electrode when not conditioned to the environment after an *in*  
484 *situ* single-point calibration was performed (0.1072 and 0.1394 to 0.0475 and 0.0741 units,  
485 respectively), while the external electrode improved only minimally from 0.0988 and 0.0963 to  
486 0.0610 and 0.0894 units, respectively (Fig. 4). When *in situ* single-point calibration was  
487 performed after the sensors<sub>395, 396</sub> were conditioned (i.e., calibrated with reference sample taken  
488 on 4 June 2017, 03:00 UTC), the  $\text{pH}_t$  uncertainty for the internal electrode reduced only  
489 minimally from factory calibration: 0.1072 and 0.1394 to 0.0896 and 0.1240 units, respectively  
490 (Fig. 5a, b). Conversely, the  $\text{pH}_t$  error for the external electrode increased from 0.0988 and  
491 0.0963 to 0.1011 and 0.1480, respectively (Fig 5c, d).

492  
493 Both sensors<sub>395, 396</sub> displayed low inter-sensor variability for the internal electrode, and  
494 high for the external electrode after *in situ* single-point calibration was performed on sensors not  
495 conditioned to the environment (Fig. 6, gray circles). The mean anomaly between both  
496 sensor's<sub>395, 396</sub> internal electrodes was 0.0525 units, whereas the external mean anomaly was  
497 0.145 units. When measurements taken before the sensor was conditioned to the environment

498 (blue shaded region Fig. 6) were removed from analysis, the mean anomaly changed by  $< 0.006$   
499 units for both electrodes. Inter-sensor variability for both electrodes once conditioned, and after  
500 *in situ* single-point calibration, was  $< 0.05$  units: 0.0409 and 0.0461 units for the internal and  
501 external electrodes, respectively (Fig. 6, black circles). When measurements recorded before the  
502 sensors were conditioned to the environment were removed (blue shaded region Fig. 10), the  
503 anomaly decreased further,  $< 0.015$  units for both electrodes.  
504

505 Thermistor readings on both sensors<sub>395, 396</sub> tracked the temperature at the Seldovia site  
506 well, however errant spikes occurred around 18 April 2017 and again around 10 May 2017, and  
507 continued till the end of the deployment (Fig. 7). The absolute average difference between the  
508 thermistor values and the Seldovia data sonde was  $0.281\text{ }^{\circ}\text{C}$  (SD 0.295), nearly identical to the  
509 difference displayed during the test tank deployments, average  $0.21\text{ }^{\circ}\text{C}$ .  
510

511 At Sentry Shoal, temperature and salinity seasonally fluctuated and ranged from  $8.71 -$   
512  $21.8\text{ }^{\circ}\text{C}$  and  $23.4 - 29.4$ , respectively. Based on the overall accuracy of the internal and external  
513 electrodes, there was no clear distinction as to which provided the more robust measurement  
514 after *in situ* single-point calibration was performed. While the external electrode did display a  
515 lower  $\text{pH}_t$  average uncertainty, this was based on only two reference points, one of which had a  
516 time discrepancy of 5 min (9 November 2016, 17:05 UTC). Only two reference samples were  
517 comparable against sensor<sub>268</sub>  $\text{pH}_t$  due to the loss of salinity and temperature data on 23 August  
518 2016, 17:00 UTC. Reference samples on 26 September 2016 and 9 November 2016 were,  
519 therefore, compared using the new calibration coefficients determined after redeployment on 27  
520 August 2016. The average  $\text{pH}_t$  uncertainty was  $< 0.0115$  units for both electrodes (Fig. 8)  
521 compared to average  $\text{pH}_t$  uncertainties of 0.0244 and 0.0560 units for the internal and external  
522 electrodes, respectively, if initial calibration coefficients from 19 July 2016 were retained. The  
523 low  $\text{pH}_t$  uncertainty ( $< 0.0137$  units) determined after the *in situ* single-point calibration,  
524 however, was still greater than the average  $\text{pH}_t$  uncertainty under factory calibration:  $< 0.005$   
525 units for both electrodes (Fig 8).  
526

### 527 **3.4 Spectral analysis**

528  
529 All sensors<sub>395, 396, 397, 268</sub> displayed a mixed semi-diurnal tidal response during all field  
530 deployments (Fig. 9). SeaFETs<sup>TM</sup><sub>395, 396</sub> at Kasitsna Bay had a stronger amplitude response at a  
531 frequency of two cycles  $\text{d}^{-1}$ , whereas sensor<sub>397</sub> had a greater amplitude at one cycle  $\text{d}^{-1}$  (Fig. 9a,  
532 c, d). All three sensors<sub>395, 396, 397</sub> in Alaskan waters had a strong amplitude signal of 1 cycle  
533 every 21 days, with an addition signal of one cycle every three days for SeaFET<sup>TM</sup><sub>397</sub>. The  
534 amplitude signal for sensor<sub>397</sub> shifted depending on source of measurement (BoL, internal or  
535 external electrode), however, all measurement sources followed the same frequency pattern (Fig  
536 9a). Sensor<sub>268</sub> at Sentry Shoal displayed a strong signal at a frequency of zero as well as at one  
537 and two cycles  $\text{d}^{-1}$  (Fig 9a).  
538

### 539 **3.5 Intrinsic uncertainty and accuracy**

540  
541 Among the calculated potential sources of uncertainty in  $\text{pH}_t$ , inter-sensor variability (difference  
542 between SeaFET's<sup>TM</sup>  $\text{pH}_t$ ) and sensor accuracy produced the greatest uncertainty discrepancies  
543 for the internal and external electrodes under factory calibration (Fig. 10). The  $\text{pH}_t$  uncertainty

544 (i.e., overall sensor accuracy) for the internal electrode reduced a greater degree than the external  
545 electrode at every ordinal calibration method: factory, *in situ* single-point, to *in situ* multi-point  
546 calibration (Fig. 10). This was not the case for the external electrode, however, as the overall  $\text{pH}_t$   
547 accuracy was greater when factory calibration was used compared to an *in situ* single-point  
548 calibration was performed after the sensor was conditioned. The thermistor uncertainty (i.e.,  
549 uncertainty when calculating  $\text{pH}_t$  based on the thermistor temperature rather than a more accurate  
550 temperature gauge) produced a  $\text{pH}_t$  uncertainty of 0.0044 units, and was based on the recorded  
551 values by sensors<sub>395, 396</sub>. Even though the temperature-derived values from the thermistor of  
552 sensors<sub>395, 396</sub> were compared against a data sonde 10 km away, the average  $T_{\text{diff}}$  values were  
553 consistent with the  $T_{\text{diff}}$  calculated from the test tank deployments (within 0.07°C) and, therefore,  
554 provided an adequate resolution to determine a thermistor uncertainty value.

555

#### 556 4 Discussion

557

558 Obtaining accurate and precise measurements of pH in nearshore coastal waters is crucial for  
559 understanding changing trends, dynamics, and current baselines of acidification in these—  
560 “susceptible to change”—marine domains. For dynamic nearshore systems, the current standard  
561 of OA weather (carbonate chemistry variability on timescales of days to months) accuracy  
562 should have an uncertainty no greater than 0.02 pH units according to the Global Ocean  
563 Acidification Observing Network (Newton et al. 2015). Previous evaluations of the SeaFET™  
564 sensor package have demonstrated accuracy for both electrodes to be better than 0.02 pH units,  
565 with a range between 0.01 – 0.04 units for the internal electrode in more dynamic environments  
566 (Bresnahan et al., 2014; Gonski, 2018; Martz et al., 2010). Based on our findings, we observed  
567 an accuracy range of 0.009 – 0.148  $\text{pH}_t$  units after sensors were conditioned and *in situ* single-  
568 point or multi-point calibrations were performed for the internal and external electrodes. This  
569 range decreased when SeaFETs™<sub>395, 396</sub> from Kasitsna Bay were calibrated with reference  
570 samples taken at initial deployment (i.e., non-conditioned to environment). For SeaFET™<sub>397</sub>, the  
571 internal electrode’s accuracy was nearly identical to that of the external electrode after an *in situ*  
572 multi-point calibration (Fig. 2), suggesting that the internal electrode can produce a highly  
573 precise  $\text{pH}_t$  measurement comparable to the BoL with an accuracy meeting the standards of the  
574 OA weather measurements (Newton et al. 2015). This is not to suggest that the SeaFET™ can  
575 replace the BoL, particularly because the BoL can capture multiple carbonate chemistry  
576 measurements thereby fully constraining the system and identifying potential decoupling of the  
577 carbonate system in estuarine waters (Bandstra et al., 2006; Hales et al., 2016). Nonetheless, the  
578 SeaFET™ can provide an accurate measurement of  $\text{pH}_t$  in nearshore waters when SeaFET™  
579 operation is executed with high precision.

580

581 Sensors<sub>397, 268</sub> deployed at the APSH and at Sentry Shoal displayed the lowest  
582 uncertainty and greatest precision of  $\text{pH}_t$  measurements (Fig. 2 and 8). In both instances, the  
583 sensors<sub>397, 268</sub> were adequately conditioned (i.e., subjected to *in situ* conditions for ~50 days)  
584 before calibration was performed. The greater overall accuracy displayed by sensor<sub>268</sub> at Sentry  
585 Shoal may be due to the fact that the sensor was exposed to *in situ* conditions for a longer period  
586 of time and re-calibrated multiple times to the same environment. Further, calibration and  
587 reference sample  $\text{pH}_t$  was derived from  $\text{TCO}_2$  and  $p\text{CO}_2$  processed by the BoL at Sentry Shoal  
588 and from  $p\text{CO}_2$  (also measured by BoL) and the TA-salinity relationship (Evans et al. 2015) at  
589 the APSH. It is unclear as to why the sensor accuracy of both Kasitsna Bay sensors<sub>395, 396</sub> was

590 substantially less than the sensors<sub>397, 268</sub> at the APSH or Sentry Shoal. A potential reason for the  
591 low accuracy may be that sensors were calibrated at a reference point that was extreme relative  
592 to the time series  $\text{pH}_t$  signal—that is, calibrated at a time of high variability. In this case,  
593 performing an *in situ* multiple-point calibration could have reduced the uncertainty and increased  
594 the accuracy. While previous studies have found that collection and preservation of calibration  
595 and reference samples can result in a decrease in accuracy depending on operator experience  
596 (McLaughlin et al., 2017), the operator in this study was considered to have substantial  
597 experience conducting such operations used in this evaluation. In addition, given the increased  
598  $\text{pH}_t$  variability over a short temporal period—which can be seen at the end of the Kasitsna Bay  
599 deployment (Fig. 4 and 5)—and the low discrepancy between duplicate reference samples, the  
600 former reasoning (i.e., calibrated to an extreme reference point) is a more reasonable explanation  
601 for the reduced accuracy by the Kasitsna Bay sensors<sub>395, 396</sub> than operator experience. We re-  
602 iterate here that reference sample temperature and salinity were used to calculate SeaFET™  $\text{pH}_t$   
603 at the time points in which sensor  $\text{pH}_t$  and reference sample  $\text{pH}_t$  were compared, thus salinity  
604 was not a confounding factor.

605  
606 Despite the lower accuracy of the Kasitsna Bay SeaFETs™<sub>395, 396</sub>, the two sensors  
607 provided a better insight of inter-sensor variability for electrodes non-conditioned and electrodes  
608 conditioned to the environment. After *in situ* single-point calibration for conditioned sensors, the  
609 average inter-sensor variability decreased for the internal electrode by ~80%, and >300% for the  
610 external electrode (Fig. 6). The inter-sensor variability reported here was still greater than  
611 previous findings (Kapsenberg et al., 2017), however, the comparison made in this study was  
612 done in the field compared to controlled laboratory conditions as in Kapsenberg et al. (2017).  
613 And while non-homogenized water could lead to anomalies in  $\text{pH}_t$  measurements by the Kasitsna  
614 Bay sensors<sub>395, 396</sub>, it is unlikely that water was consistently non-homogenized over the entirety  
615 of a deployment at a distance of < 20 cm (distance between electrodes on each sensor).  
616 Furthermore, due to the dynamic nature of Kachemak Bay, where the tidal exchanges are  
617 extreme, averaging 4.73 m, it is unlikely that micro-heterogeneity of seawater is the driving force  
618 behind the observed differences in  $\text{pH}_t$  measurements that were observed between sensors<sub>395, 396</sub>.  
619 There was a tradeoff for a decrease in inter-sensor variability, as the *in situ* single-point  
620 calibration performed after sensors were conditioned resulted in a decrease in accuracy compared  
621 to an *in situ* single-point calibration performed for sensors not conditioned to the environment. It  
622 should be noted that we do not consider salinity to be a potential source of uncertainty for inter-  
623 sensor variability because the  $\text{pH}_t$  difference using data sonde salinity compared to a fixed  
624 salinity resulted in an anomaly of < 0.005 units.

625  
626 The influence of rapid environmental variability should be acknowledged here as this can  
627 create uncertainty in autonomous sensor operation and accuracy (Tamburri et al. 2011).  
628 While the temperature changes due to rapid environmental change in Kasitsna Bay equate to a  
629 potential 0.011 discrepancy in  $\text{pH}$ , previous evaluation of these sensors show that rapid response  
630 to temperature changes should be negligible and result in uncertainties below the accuracy  
631 assured when applying an average temperature coefficient ( $k_2$ ), which can result in discrepancies  
632 of <0.015  $\text{pH}$  units (Bresnahan et al. 2014). Rapid changes in salinity could also result in  
633 uncertainties regarding SeaFET™ accuracy and may be responsible for the noisier signal  
634 observed by the external electrode for the sensors<sub>395, 396</sub> deployed in Kasitsna Bay. The greatest  
635 salinity change within a 3 h period observed in Kasitsna Bay was 3.90. Given that the mean

636 salinity at the deployment site was 31.8, a mismatch in timing here, or lag in response, could  
637 equate to pH changes as great as 0.053 units—although this likely not a realistic change as this  
638 was the maximum difference within a 3 h period. It should be noted that rapid salinity changes  
639 would only affect the external electrode as the internal electrode is insensitive to changes in  
640 salinity. Due to the uncertainties that can emerge from rapid environmental variability, we  
641 reiterate the benefits of an operator understanding the deployment site as this will enhance data  
642 collection by the SeaFET™.

643  
644 The Sentry Shoal sensor<sub>268</sub> had the lowest average  $pH_t$  uncertainty for both electrodes  
645 after *in situ* single-point calibration was performed, however, these were still greater than the  $pH_t$   
646 uncertainty determined using the factory calibration coefficients. This specific example  
647 highlights two possibilities: (1) the role of inter-sensor variability, as this may be a coincidental  
648 case given the uncertainty observed when quantifying inter-sensor variability, and (2) the  
649 influence of variance within a calibration sample set. For the case of SeaFET™<sub>268</sub>, the replicate  
650 calibration samples collected on 19 July 2016 and 2 September 2016 for the first and second  
651 deployments had standard deviations of 0.016 and 0.005  $pH_t$  units, respectively. When factory  
652 and *in situ* calibrated data produce final  $pH_t$  values in close agreement, it is important to  
653 recognize that the variance in the calibration sample set may contribute to better agreement  
654 between factory calibrated sensor  $pH_t$  data and average discrete sample  $pH_t$  measurements. It  
655 should also be noted that pre-deployment calibration can provide highly accurate measurements  
656 by the Honeywell Durafet (internal electrode), however, matching exact conditions to those at  
657 the field site are necessary (Johnson et al., 2017), and this was not likely the case for the factory  
658 provided calibration coefficients.

659  
660 The evaluation of SeaFET™ performance presented here corroborates and contrasts with  
661 previous studies examining the overall accuracy and precision of  $pH_t$  measurements made by  
662 these oceanographic instruments. While the accuracy of two sensors<sub>397, 268</sub> fall well within the  
663 range determined from previous studies, the accuracy of sensors<sub>395, 396</sub> at Kasitsna Bay lay  
664 outside the bounds of what has been reported in the primary literature (Bresnahan et al., 2014;  
665 Gonski et al., 2018; Johnson et al., 2017; Kapsenberg et al., 2017; Martz et al., 2010). For  
666 example, Bresnahan et al. (2014) describes intrinsic Durafet uncertainties less than 0.03 units,  
667 but this varied depending on the validating reference source (e.g., spectrophotometric pH or  
668 estimated pH from O<sub>2</sub>). One reason as to why the Kasitsna Bay SeaFET's™ uncertainties  
669 differed from Bresnahan et al. (2014) may be due to the fact that calibration was performed ~78  
670 days after deployment. Thus, we suggest that in a highly dynamic area such as Kasitsna Bay,  
671 calibration should be performed immediately after conditioning. While there is no way to  
672 officially conclude that this could have reduced uncertainty, it is one potential source of  
673 discrepancy. Following current best practices in Bresnahan et al. (2014) may yield robust  
674 measurements, however, the utility of our assessment describes the importance of knowing when  
675 to take calibration samples as a means to decrease uncertainties. Nevertheless, it is relevant to  
676 report the potential uncertainties possible when operating SeaFETs™ as a multitude of factors  
677 can influence the overall accuracy (e.g., operator, sample preservation, electrode conditioning,  
678 calibration measurements), therefore, the potential uncertainties calculated in this study represent  
679 the upper limit of an average uncertainty compiled from four different SeaFETs™ (Fig. 10). The  
680 utility of such an analysis provides a confidence in SeaFET™ operation, and highlights all the  
681 potential uncertainties that need to be considered when deploying the sensors in the field. For

682 example, we have included a thermistor uncertainty term determined from the test tank and field  
683 deployments of the Alaska sensors<sub>395, 396, 397</sub>, even though a suitable solution around this issue  
684 would be to apply an offset to the thermistor temperature given it was compared to more robust  
685 temperature measurements conducted before field deployment. It should be noted, that in this  
686 case, the thermistor uncertainty observed from sensor<sub>397</sub> against the BoL was excluded as the  
687 lag time between thermistor response and tank residence time likely confounded the comparison.  
688 The potential pH<sub>t</sub> uncertainties presented here should serve as a guide for SeaFET™ operators in  
689 order to better understand the source of an uncertainty and take the necessary steps to improve  
690 SeaFET™ measurements. Bresnahan et al. (2014) acknowledged that relying on the SeaFET™  
691 for an accurate pH measurement should be viewed cautiously if additional biogeochemical  
692 sensors are not co-deployed to cross-validate the stability and accuracy of the SeaFET's™  
693 electrodes, therefore, being fully aware of all the potential uncertainties presented here will only  
694 further aid SeaFET™ operators.

695  
696 The time series data provided by the SeaFET™ deployments in this study have expanded  
697 the extent of recorded pH<sub>t</sub> variability along the North American west coast. The sensors<sub>395, 396</sub>  
698 deployed in Kasitsna Bay provide some of the first high temporal resolution measurements of  
699 pH<sub>t</sub> in this region. During this spring deployment, it appears that semi-diurnal tidal fluctuations  
700 are the dominant contributor to pH<sub>t</sub> variability with an additional cycle occurring every 21 days  
701 coinciding with the seasonal spring and neap tides (Fig. 9). The sensor<sub>268</sub> at Sentry Shoal also  
702 displays a strong pH<sub>t</sub> response to the semi-diurnal mixed tidal cycle. A strong signal is also  
703 present at a frequency of zero, and is likely a result of the long, across-season, time series. That  
704 is, over the course of the entire deployment which went from summer into late fall, seasonal  
705 drivers of pH<sub>t</sub> (e.g., decrease in water temperature) confounded repetitive frequency patterns. In  
706 addition, Sentry Shoal may have a weaker tidal signature relative to other pH<sub>t</sub> modulators that do  
707 not follow a cyclical pattern such as water mass intrusion, inconsistent metabolic cycles from the  
708 end of summer into the fall season, and a shift to the rainy season.

709  
710 As an elaboration on the power spectral density analysis, we suggest this form of  
711 frequency analysis can be utilized to better understand the system in which a SeaFET™ is  
712 deployed, thus informing the operator as to what the drivers of their system are, and when to  
713 calibrate the sensor. It is possible that in a highly dynamic setting, the sensor could re-condition  
714 over time periods not resolved in a multi-point calibration sampling scheme, and this could  
715 enhance sensor inaccuracies. For example, in Kasitsna Bay, a strong semi-diurnal tide cycle was  
716 present, so upon redeployment in this area, if possible, the best calibration approach would be an  
717 *in situ* multi-point calibration between the mixed semi-diurnal tidal cycle. Alternatively, if the  
718 system is not driven by a strong tidal signature (e.g., non-coastal region), an *in situ* single-point  
719 calibration may be a reasonable approach. It should be noted that while spectral analysis can be  
720 used as an additional tool to better calibrate the SeaFET™, specific coastal environments with  
721 dynamic storm frequencies or varying photosynthesis and respiration cycles could obscure a  
722 clear driving frequency of pH change. In these situations, capturing the dynamic range (i.e.,  
723 multiple calibration samples over this period) of one of these events may be sufficient to provide  
724 best approach for robust calibration.

## 725 726 **5 Conclusion**

727



728 The following evaluation of the Sea-Bird SeaFET™ helped elucidate the overall  
729 accuracy and highlighted the potential uncertainties and pitfalls of operating and obtaining pH<sub>t</sub>  
730 measurements by the internal and external electrode pair. We found that the internal electrode  
731 provided the more robust measurement in nearshore estuarine waters when an *in situ* multi-point  
732 calibration was performed (Fig. 10). The quantified potential pH<sub>t</sub> uncertainty is based  
733 specifically on our findings, whereas further results may minimize this uncertainty given  
734 additional evaluations. However, the results here provide an upper limit of the pH<sub>t</sub> uncertainty  
735 that may be observed when operating a Sea-Bird SeaFET™. Further, high temporal resolution  
736 pH<sub>t</sub> measurements in nearshore Canadian and Alaskan waters provide a better understanding of  
737 the drivers modulating pH on short timescales. Given the application, the Sea-Bird SeaFET™  
738 can provide a reliable and accurate pH<sub>t</sub> measurement which can be utilized to broaden the  
739 coverage of understanding pH variability in nearshore and open-ocean waters.  
740

## 741 **Acknowledgments**

742 The authors would like to thank Jeff Hetrick and Jacqueline Ramsey at the Alutiiq Pride  
743 Shellfish Hatchery for providing their facilities and services for this evaluation. We would also  
744 like to thank Angela Doroff at the Kasitsna Bay laboratory for providing facilities for SeaFET™  
745 deployments. Funding for this project was provided in part by the University of Alaska  
746 Fairbanks College of Fisheries and Ocean Sciences. WE and KP thank the Pacific Salmon  
747 Foundation and Environment Canada for providing the platform for deploying SeaFET 268, the  
748 University of Alaska Fairbanks Ocean Acidification Research Center for the long-term use of  
749 SeaFET 268, and the Tula Foundation for supporting their efforts with this work.  
750

## 751 **References**

- 752 Bandstra, L., Hales, B. and Takahashi, T.: High-frequency measurements of total CO<sub>2</sub>: Method  
753 development and first oceanographic observations, *Mar. Chem.*, 100(1–2), 24–38,  
754 doi:10.1016/j.marchem.2005.10.009, 2006.
- 756 Barton, A., Hales, B., Waldbusser, G. G., Langdon, C. and Feely, R. A.: The Pacific oyster,  
757 *Crassostrea gigas*, shows negative correlation to naturally elevated carbon dioxide levels:  
758 Implications for near-term ocean acidification effects, *Limnol. Oceanogr.*, 57(3), 698–710,  
759 doi:10.4319/lo.2012.57.3.0698, 2012.
- 760 Bresnahan, P. J., Martz, T. R., Takeshita, Y., Johnson, K. S. and LaShomb, M.: Best practices for  
761 autonomous measurement of seawater pH with the Honeywell Durafet, *Methods Oceanogr.*, 9,  
762 44–60, doi:10.1016/j.mio.2014.08.003, 2014.
- 763 Caldeira, K. and Wickett, M. E.: Anthropogenic carbon and ocean pH, *Nature*, 425(6956), 365–  
764 365, doi:10.1038/425365a, 2003.
- 765 Chan, F., Barth, J. A., Blanchette, C. A., Byrne, R. H., Chavez, F., Cheriton, O., Feely, R. A.,  
766 Friederich, G., Gaylord, B., Gouhier, T., Hacker, S., Hill, T., Hofmann, G., McManus, M. A.,  
767 Menge, B. A., Nielsen, K. J., Russell, A., Sanford, E., Sevadjan, J. and Washburn, L.: Persistent  
768 spatial structuring of coastal ocean acidification in the California Current System, *Sci. Rep.*, 7(1),  
769 2526, doi:10.1038/s41598-017-02777-y, 2017.

770 Dickson, A. G., Sabine, C. L. and Christian, J. R.: Guide to Best Practices for Ocean CO<sub>2</sub>  
771 Measurements., Report, North Pacific Marine Science Organization. [online] Available from:  
772 <http://www.oceandatapactices.net:80/handle/11329/249>, 2007.

773 Duarte, C. M., Hendriks, I. E., Moore, T. S., Olsen, Y. S., Steckbauer, A., Ramajo, L.,  
774 Carstensen, J., Trotter, J. A. and McCulloch, M.: Is Ocean Acidification an Open-Ocean  
775 Syndrome? Understanding Anthropogenic Impacts on Seawater pH, *Estuaries Coasts*, 36(2),  
776 221–236, doi:10.1007/s12237-013-9594-3, 2013.

777 Ekstrom, J. A., Suatoni, L., Cooley, S. R., Pendleton, L. H., Waldbusser, G. G., Cinner, J. E.,  
778 Ritter, J., Langdon, C., van Hooidek, R., Gledhill, D., Wellman, K., Beck, M. W., Brander, L.  
779 M., Rittschof, D., Doherty, C., Edwards, P. E. T. and Portela, R.: Vulnerability and adaptation of  
780 US shellfisheries to ocean acidification, *Nat. Clim. Change*, 5(3), 207–214,  
781 doi:10.1038/NCLIMATE2508, 2015.

782 Evans, W., Mathis, J. T. and Cross, J. N.: Calcium carbonate corrosivity in an Alaskan inland  
783 sea, *Biogeosciences*, 11(2), 365–379, doi:10.5194/bg-11-365-2014, 2014.

784 Evans, W., Mathis, J. T., Ramsay, J. and Hetrick, J.: On the Frontline: Tracking Ocean  
785 Acidification in an Alaskan Shellfish Hatchery, *PLOS ONE*, 10(7), e0130384,  
786 doi:10.1371/journal.pone.0130384, 2015.

787 Feely, R. A., Alin, S. R., Newton, J., Sabine, C. L., Warner, M., Devol, A., Krembs, C. and  
788 Maloy, C.: The combined effects of ocean acidification, mixing, and respiration on pH and  
789 carbonate saturation in an urbanized estuary, *Estuar. Coast. Shelf Sci.*, 88(4), 442–449,  
790 doi:10.1016/j.ecss.2010.05.004, 2010.

791 Feely, R. A., Alin, S. R., Carter, B., Bednaršek, N., Hales, B., Chan, F., Hill, T. M., Gaylord, B.,  
792 Sanford, E., Byrne, R. H., Sabine, C. L., Greeley, D. and Juranek, L.: Chemical and biological  
793 impacts of ocean acidification along the west coast of North America, *Estuar. Coast. Shelf Sci.*,  
794 183, Part A, 260–270, doi:10.1016/j.ecss.2016.08.043, 2016.

795 Gonski, S. F., Cai, W.-J., Ullman, W. J., Joesoef, A., Main, C. R., Pettay, D. T. and Martz, T. R.:  
796 Assessment of the suitability of Durafet-based sensors for pH measurement in dynamic estuarine  
797 environments, *Estuar. Coast. Shelf Sci.*, 200(Supplement C), 152–168,  
798 doi:10.1016/j.ecss.2017.10.020, 2018.

799 Hales, B., Suhrbier, A., Waldbusser, G. G., Feely, R. A. and Newton, J. A.: The Carbonate  
800 Chemistry of the “Fattening Line,” Willapa Bay, 2011–2014, *Estuaries Coasts*, 1–14,  
801 doi:10.1007/s12237-016-0136-7, 2016.

802 Harris, K. E., DeGrandpre, M. D. and Hales, B.: Aragonite saturation state dynamics in a coastal  
803 upwelling zone, *Geophys. Res. Lett.*, 40(11), 2720–2725, doi:10.1002/grl.50460, 2013.

804 Hofmann, G. E., Smith, J. E., Johnson, K. S., Send, U., Levin, L. A., Micheli, F., Paytan, A.,  
805 Price, N. N., Peterson, B., Takeshita, Y., Matson, P. G., Crook, E. D., Kroeker, K. J., Gambi, M.  
806 C., Rivest, E. B., Frieder, C. A., Yu, P. C. and Martz, T. R.: High-Frequency Dynamics of Ocean

807 pH: A Multi-Ecosystem Comparison, *Plos One*, 6(12), e28983,  
808 doi:10.1371/journal.pone.0028983, 2011.

809 Johnson, K. S., Plant, J. N., Coletti, L. J., Jannasch, H. W., Sakamoto, C. M., Riser, S. C., Swift,  
810 D. D., Williams, N. L., Boss, E., Haentjens, N., Talley, L. D. and Sarmiento, J. L.:  
811 Biogeochemical sensor performance in the SOCCOM profiling float array, *J. Geophys. Res.-*  
812 *Oceans*, 122(8), 6416–6436, doi:10.1002/2017JC012838, 2017.

813 Kapsenberg, L., Bockmon, E. E., Bresnahan, P. J., Kroeker, K. J., Gattuso, J.-P. and Martz, T.  
814 R.: Advancing Ocean Acidification Biology Using Durafet® pH Electrodes, *Front. Mar. Sci.*, 4,  
815 doi:10.3389/fmars.2017.00321, 2017.

816 Kapsenberg, L. and Hofmann, G. E.: Ocean pH time-series and drivers of variability along the  
817 northern Channel Islands, California, USA, *Limnol. Oceanogr.*, 61(3), 953–968,  
818 doi:10.1002/lno.10264, 2016.

819 Kapsenberg, L., Kelley, A. L., Shaw, E. C., Martz, T. R. and Hofmann, G. E.: Near-shore  
820 Antarctic pH variability has implications for the design of ocean acidification experiments, *Sci.*  
821 *Rep.*, 5, srep09638, doi:10.1038/srep09638, 2015.

822 Khoo, K. H., Ramette, R. W., Culbertson, C. H. and Bates, R. G.: Determination of hydrogen ion  
823 concentrations in seawater from 5 to 40.degree.C: standard potentials at salinities from 20 to 45  
824 ‰, *Anal. Chem.*, 49(1), 29–34, doi:10.1021/ac50009a016, 1977.

825 Lueker, T. J., Dickson, A. G. and Keeling, C. D.: Ocean pCO<sub>2</sub> calculated from dissolved  
826 inorganic carbon, alkalinity, and equations for K<sub>1</sub> and K<sub>2</sub>: validation based on laboratory  
827 measurements of CO<sub>2</sub> in gas and seawater at equilibrium, *Mar. Chem.*, 70(1–3), 105–119,  
828 doi:10.1016/S0304-4203(00)00022-0, 2000.

829 Martz, T., Send, U., Ohman, M. D., Takeshita, Y., Bresnahan, P., Kim, H.-J. and Nam, S.:  
830 Dynamic variability of biogeochemical ratios in the Southern California Current System,  
831 *Geophys. Res. Lett.*, 41(7), 2496–2501, doi:10.1002/2014GL059332, 2014.

832 Martz, T. R., Connery, J. G. and Johnson, K. S.: Testing the Honeywell Durafet® for seawater  
833 pH applications, *Limnol. Oceanogr. Methods*, 8(5), 172–184, doi:10.4319/lom.2010.8.172, 2010.

834 Martz, T. R., Daly, K. L., Byrne, R. H., Stillman, J. H. and Turk, D.: Technology for ocean  
835 acidification research: needs and availability, *Oceanography*, 28(2), 40–47, 2015.

836 Mathis, J. T., Cross, J. N. and Bates, N. R.: Coupling primary production and terrestrial runoff to  
837 ocean acidification and carbonate mineral suppression in the eastern Bering Sea, *J. Geophys.*  
838 *Res. Oceans*, 116(C2), C02030, doi:10.1029/2010JC006453, 2011a.

839 Mathis, J. T., Cross, J. N. and Bates, N. R.: The role of ocean acidification in systemic carbonate  
840 mineral suppression in the Bering Sea, *Geophys. Res. Lett.*, 38(19), L19602,  
841 doi:10.1029/2011GL048884, 2011b.

842 Mathis, J. T., Pickart, R. S., Byrne, R. H., McNeil, C. L., Moore, G. W. K., Juranek, L. W., Liu,  
843 X., Ma, J., Easley, R. A., Elliot, M. M., Cross, J. N., Reisdorph, S. C., Bahr, F., Morison, J.,  
844 Lichendorf, T. and Feely, R. A.: Storm-induced upwelling of high pCO<sub>2</sub> waters onto the  
845 continental shelf of the western Arctic Ocean and implications for carbonate mineral saturation  
846 states, *Geophys. Res. Lett.*, 39(7), L07606, doi:10.1029/2012GL051574, 2012.

847 Mathis, J. T., Cross, J. N., Monacci, N., Feely, R. A. and Stabeno, P.: Evidence of prolonged  
848 aragonite undersaturations in the bottom waters of the southern Bering Sea shelf from  
849 autonomous sensors, *Deep-Sea Res. Part-II. Stud. Oceanogr.*, 109, 125–133,  
850 doi:10.1016/j.dsr2.2013.07.019, 2014.

851 Mathis, J. T., Cross, J. N., Evans, W. and Doney, S. C.: Ocean Acidification in the Surface  
852 Waters of the Pacific-Arctic Boundary Regions, *Oceanography*, 28(2), 122–135,  
853 doi:10.5670/oceanog.2015.36, 2015a.

854 Mathis, J. T., Cooley, S. R., Lucey, N., Colt, S., Ekstrom, J., Hurst, T., Hauri, C., Evans, W.,  
855 Cross, J. N. and Feely, R. A.: Ocean acidification risk assessment for Alaska’s fishery sector,  
856 *Prog. Oceanogr.*, 136, 71–91, doi:10.1016/j.pocean.2014.07.001, 2015b.

857 Matson, P. G., Martz, T. R. and Hofmann, G. E.: High-frequency observations of pH under  
858 Antarctic sea ice in the southern Ross Sea, *Antarct. Sci.*, 23(6), 607–613,  
859 doi:10.1017/S0954102011000551, 2011.

860 McLaughlin, K., Dickson, A., Weisberg, S. B., Coale, K., Elrod, V., Hunter, C., Johnson, K. S.,  
861 Kram, S., Kudela, R., Martz, T., Negrey, K., Passow, U., Shaughnessy, F., Smith, J. E., Tadesse,  
862 D., Washburn, L. and Weis, K. R.: An evaluation of ISFET sensors for coastal pH monitoring  
863 applications, *Reg. Stud. Mar. Sci.*, 12, 11–18, doi:10.1016/j.rsma.2017.02.008, 2017.

864 Newton J.A., Feely R. A., Jewett E. B., Williamson P. & Mathis J.  
865 2015. Global Ocean Acidification Observing Network: Requirements and Governance Plan.  
866 Second Edition, GOA-ON, [http://www.goa-on.org/docs/GOA-ON\\_plan\\_print.pdf](http://www.goa-on.org/docs/GOA-ON_plan_print.pdf).  
867

868 Newton, J., Devol, A., Alford, M., Mickett, J., Sabine, C. and Sutton, A.: Nanoos Contributions  
869 to Understanding Ocean Acidification, *J. Shellfish Res.*, 31(1), 327–327, 2012.

870 Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. A., Gnanadesikan, A.,  
871 Gruber, N., Ishida, A., Joos, F., Key, R. M., Lindsay, K., Maier-Reimer, E., Matear, R., Monfray,  
872 P., Mouchet, A., Najjar, R. G., Plattner, G. K., Rodgers, K. B., Sabine, C. L., Sarmiento, J. L.,  
873 Schlitzer, R., Slater, R. D., Totterdell, I. J., Weirig, M. F., Yamanaka, Y. and Yool, A.:  
874 Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying  
875 organisms, *Nature*, 437(7059), 681–686, doi:10.1038/nature04095, 2005.

876 Orr, J. C., J.-M. Epitalon, A. G. Dickson, and J.-P. Gattuso: Routine uncertainty propagation for  
877 the marine carbon dioxide system, *Marine Chemistry*, *in prep.*  
878

879 Riebesell, U. and Gattuso, J.-P.: Lessons learned from ocean acidification research, *Nat. Clim.*  
880 *Change*, 5(1), 12–14, doi:10.1038/nclimate2456, 2015.

881 Rudd, M. A.: What a Decade (2006–15) Of Journal Abstracts Can Tell Us about Trends in  
882 Ocean and Coastal Sustainability Challenges and Solutions, *Front. Mar. Sci.*, 4,  
883 doi:10.3389/fmars.2017.00170, 2017.

884 Steinhart, J. S. and Hart, S. R.: Calibration curves for thermistors, *Deep-Sea Research*, 15(4),  
885 497–503, doi:10.1016/0011-7471(68)90057-0, 1968.

886 Sunda, W. G. and Cai, W.-J.: Eutrophication Induced CO<sub>2</sub>-Acidification of Subsurface Coastal  
887 Waters: Interactive Effects of Temperature, Salinity, and Atmospheric P-CO<sub>2</sub>, *Environ. Sci.*  
888 *Technol.*, 46(19), 10651–10659, doi:10.1021/es300626f, 2012.

889 Takeshita, Y., Martz, T. R., Johnson, K. S. and Dickson, A. G.: Characterization of an Ion  
890 Sensitive Field Effect Transistor and Chloride Ion Selective Electrodes for pH Measurements in  
891 Seawater, *Anal. Chem.*, 86(22), 11189–11195, doi:10.1021/ac502631z, 2014.

892• Tamburri, M. N., Johengen, T. H., Atkinson, M. J., Schar, D. W. H., Robertson, C. Y., Purcell,  
893 H., Smith, G. J., Pinchuk, A. and Buckley, E. N.: Alliance for Coastal Technologies, *Marine*  
894 *Technology Society Journal*, 45(1), 43–51, doi: 10.4031/MTSJ.45.1.4, 2011.

895 Uppström, L. R.: The boron/chlorinity ratio of deep-sea water from the Pacific Ocean, *Deep Sea*  
896 *Res. Oceanogr. Abstr.*, 21, 161–162, doi:10.1016/0011-7471(74)90074-6, 1974.

897 Waldbusser, G. G. and Salisbury, J. E.: Ocean Acidification in the Coastal Zone from an  
898 Organism’s Perspective: Multiple System Parameters, Frequency Domains, and Habitats,  
899 *Annu. Rev. Mar. Sci.*, 6(1), 221–247, doi:10.1146/annurev-marine-121211-172238, 2014.

900 Yu, P. C., Matson, P. G., Martz, T. R. and Hofmann, G. E.: The ocean acidification seascape and  
901 its relationship to the performance of calcifying marine invertebrates: Laboratory experiments on  
902 the development of urchin larvae framed by environmentally-relevant pCO<sub>2</sub>/pH, *J. Exp. Mar.*  
903 *Biol. Ecol.*, 400(1–2), 288–295, doi:10.1016/j.jembe.2011.02.016, 2011.

904  
905  
906  
907  
908  
909  
910  
911  
912  
913  
914  
915  
916  
917  
918  
919

920 **Table 1.** Deployment regime of all four SeaFETs™ including deployment location, date, and  
 921 calibration methods performed. \*Non-controlled source water pumped directly from  
 922 Resurrection Bay, AK, USA.

923  
 924

| Location<br>(Tank or Field)  | Date                                      | SeaFET ID     | Average<br>reads frame <sup>-1</sup> | Frames<br>Burst <sup>-1</sup> | Sampling<br>Freq. (min) | Calibration method                   |
|------------------------------|---|---------------|--------------------------------------|-------------------------------|-------------------------|--------------------------------------|
| APSH — <i>Tank</i>           | Oct. 5 – 8, 2016                          | 395, 396, 397 | 1                                    | 10                            | 5                       | Factory                              |
| OARC — <i>Tank</i>           | Oct. 26 – Nov. 3,<br>2016                 | 395, 396, 397 | 3                                    | —                             | Continuous              | Factory                              |
| OARC — <i>Tank</i>           | Jan. 26 – Feb. 1,<br>2017                 | 395, 396, 397 | 1                                    | 10                            | 180                     | Factory                              |
| APSH<br><i>Field*</i>        | Mar. 5th – Jun. 6th,<br>2017              | 397           | 10                                   | 30                            | 180                     | Factory,<br>SP and MP <i>in situ</i> |
| Kachemak Bay<br><i>Field</i> | Mar. 18th – Jun. 4th,<br>2017             | 395, 396      | 10                                   | 30                            | 180                     | Factory, SP <i>in situ</i>           |
| Sentry Shoal<br><i>Field</i> | Jul. 7th – Aug. 24,<br>Aug. 28 – Nov. 29, | 268           | 10                                   | 30                            | 30                      | Factory, SP <i>in situ</i>           |

925  
 926  
 927  
 928  
 929  
 930  
 931  
 932  
 933  
 934  
 935  
 936  
 937  
 938  
 939  
 940  
 941  
 942  
 943  
 944  
 945  
 946  
 947

Factory: factory calibration; SP: *in situ* single-point calibration; MP: *in situ* multi-point calibration.

948 **Table 2.** Terms and definitions used to describe the evaluation of the Sea-Bird SeaFET™ based  
 949 on observations specific to this study.

950  
 951

| Terms                   | Definition   |
|-------------------------|--|
| Uncertainty             | One or multiple factors that result in a discrepancy between SeaFET™ pH - "True pH" that are non-correctable |
| Accuracy                | Difference between SeaFET™ pH - "True pH"  |
| Overall Accuracy        | Integrated uncertainties   |
| "True pH <sub>t</sub> " | pH on the total scale measured by robust bench top methods: either VINDTA 3C or the Burke-o-lator            |
| Variability             | Specific difference in pH <sub>t</sub> between the internal or external electrodes on SeaFETs™ 395 and 396   |
| Mean Anomaly            | Average difference between the internal and external electrode pH <sub>t</sub>                               |

952  
 953  
 954  
 955  
 956  
 957  
 958  
 959  
 960  
 961  
 962  
 963  
 964  
 965  
 966  
 967  
 968  
 969  
 970

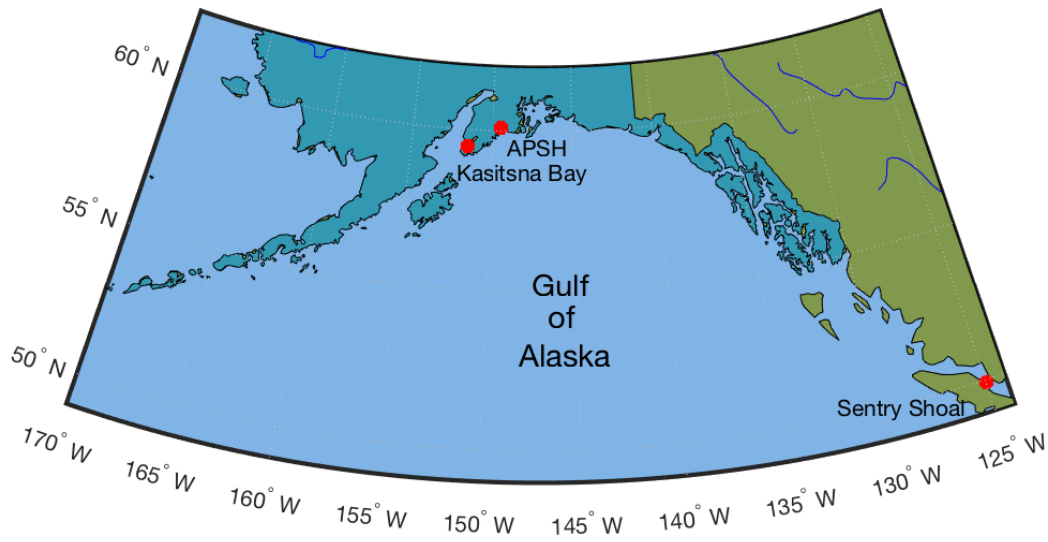
971 **Table 3.** One-way Analysis of variance comparing the  $\text{pH}_t$  error (SeaFET™  $\text{pH}_t$  – BoL  $\text{pH}_t$ )  
 972 across calibration methods for both the internal and external electrodes onboard SeaFETs™<sub>268</sub> at  
 973 Sentry Shoal (factory calibration and *in situ* single-point calibration) and SeaFET™<sub>397</sub> at the  
 974 Alutiiq Pride Shellfish Hatchery (factory calibration, *in situ* single-point calibration, and *in situ*  
 975 multi-point calibration). Bold type denotes statistical significance.  
 976

| Site | Electrode | Source                       | SS    | df  | MS    | F        | p-value        |
|------|-----------|------------------------------|-------|-----|-------|----------|----------------|
| APSH | Internal  | Fac Cal. vs. Single-point    | 27.5  | 1   | 27.5  | 4.96E+04 | < <b>0.001</b> |
|      |           | Error                        | 0.225 | 406 | 0.001 |          |                |
|      |           | Total                        | 27.7  | 407 |       |          |                |
| APSH | External  | Fac Cal. vs. Single-point    | 0.681 | 1   | 0.681 | 536      | < <b>0.001</b> |
|      |           | Error                        | 0.516 | 406 | 0.001 |          |                |
|      |           | Total                        | 1.19  | 407 |       |          |                |
| APSH | Internal  | Factory Cal. vs. Multi-point | 28.3  | 1   | 28.3  | 6.19E+04 | < <b>0.001</b> |
|      |           | Error                        | 0.185 | 406 | 0.001 |          |                |
|      |           | Total                        | 28.5  | 407 |       |          |                |
| APSH | External  | Factory Cal. vs. Multi-point | 0.692 | 1   | 0.692 | 539      | < <b>0.001</b> |
|      |           | Error                        | 0.521 | 406 | 0.001 |          |                |
|      |           | Total                        | 1.21  | 407 |       |          |                |
| APSH | Internal  | Single-point vs. Multi-point | 0.005 | 1   | 0.005 | 15.0     | < <b>0.001</b> |
|      |           | Error                        | 0.143 | 406 | 0.000 |          |                |
|      |           | Total                        | 0.148 | 407 |       |          |                |
| APSH | External  | Single-point vs. Multi-point | 0.000 | 1   | 0.000 | 0.040    | 0.843          |
|      |           | Error                        | 0.415 | 406 | 0.001 |          |                |
|      |           | Total                        | 0.415 | 407 |       |          |                |

977  
 978  
 979  
 980  
 981  
 982  
 983  
 984  
 985  
 986  
 987  
 988  
 989  
 990  
 991  
 992  
 993  
 994

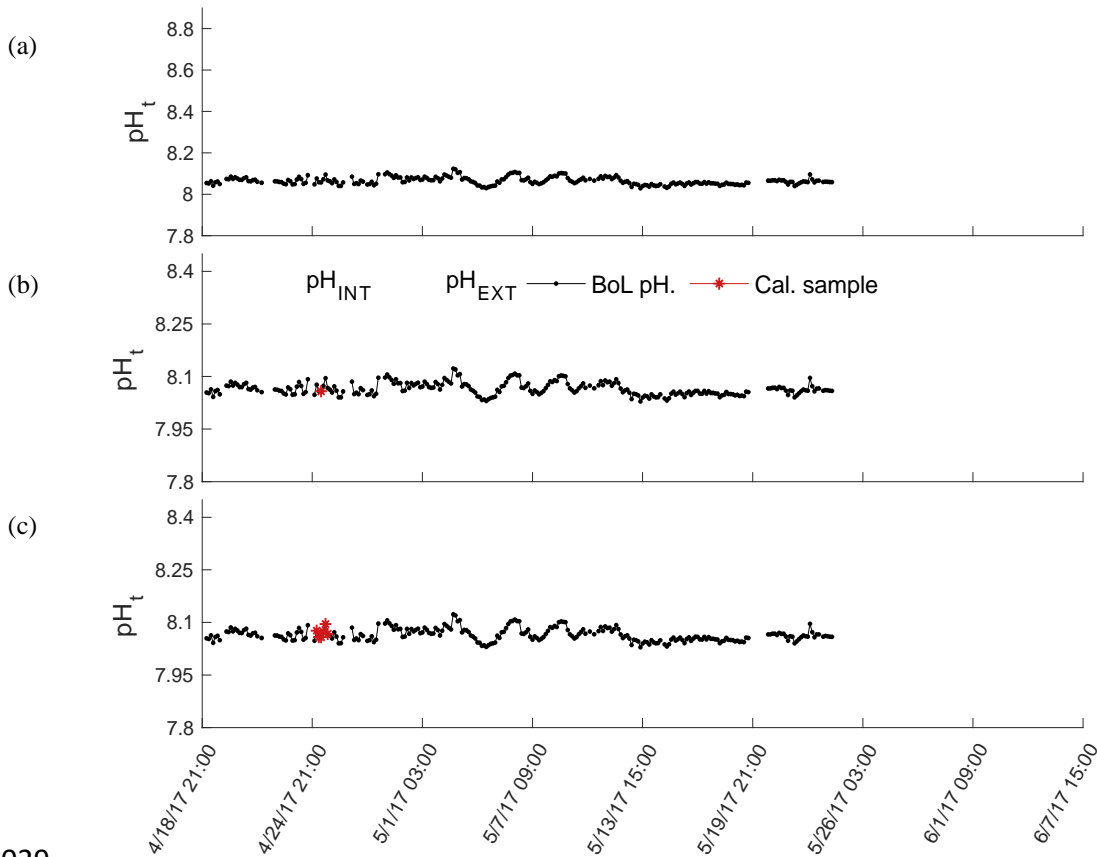


995 **Figure 1.**  
996  
997



998  
999  
1000 Geographical map with locations of SeaFET™ field deployments along Alaska's, USA, south-  
1001 central coast in Kasitsna Bay and at the Alutiiq Pride Shellfish Hatchery (APSH), and one  
1002 location at Sentry Shoal in the Strait of Georgia, British Columbia, Canada.  
1003  
1004  
1005  
1006  
1007  
1008  
1009  
1010  
1011  
1012  
1013  
1014  
1015  
1016  
1017  
1018  
1019  
1020  
1021  
1022  
1023  
1024  
1025  
1026

1027 **Figure 2.**  
1028

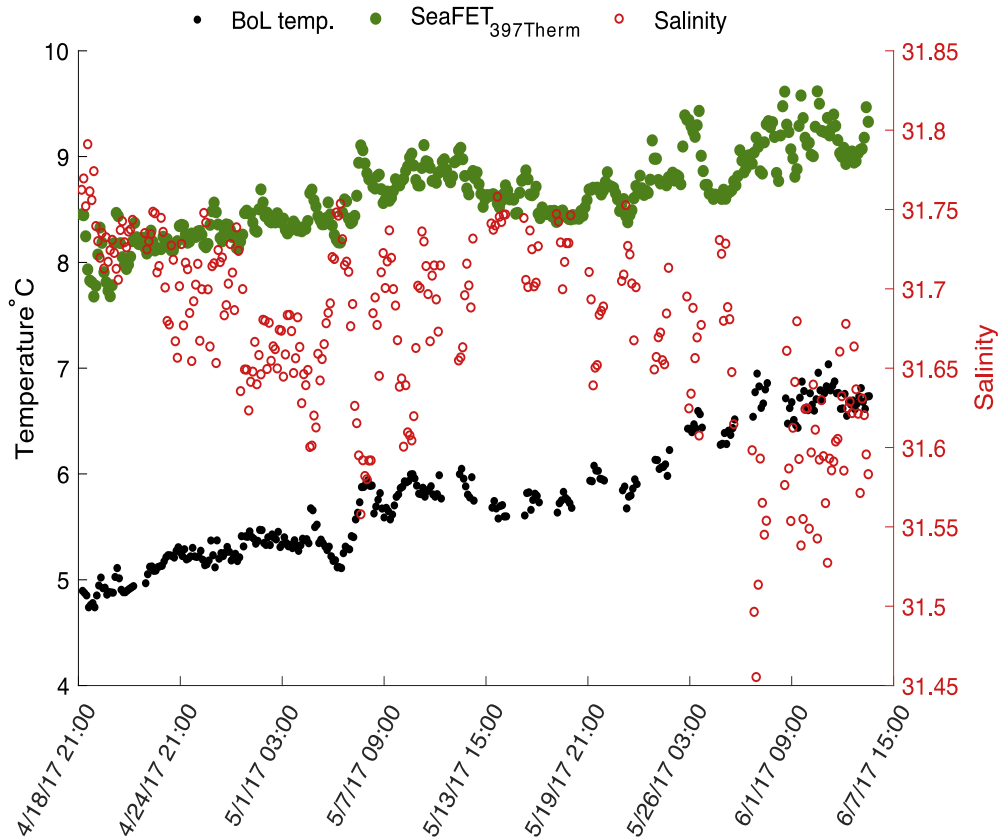


1029  
1030

1031  $pH_t$  recorded by the internal (solid) and external (dashed) electrodes on SeaFET<sup>TM</sup><sub>397</sub> deployed in  
1032 parallel with the BoL at the Alutiiq Pride Shellfish Hatchery.  $pH_t$  from both electrodes is shown  
1033 when derived using factory calibration (FC) coefficients (panel a), *in situ* single-point (SC)  
1034 calibration coefficients (panel b), and *in situ* multi-point (MC) calibration coefficients (panel c).  
1035 Black solid line is  $pH_t$  derived from continuous  $pCO_2$  measurements recorded by the BoL and  
1036 derived TA from the TA-S relationship (Evans et al. 2015). Red circles are the calibration points  
1037 from the BoL data.

1038  
1039  
1040  
1041  
1042  
1043  
1044  
1045  
1046  
1047  
1048  
1049  
1050

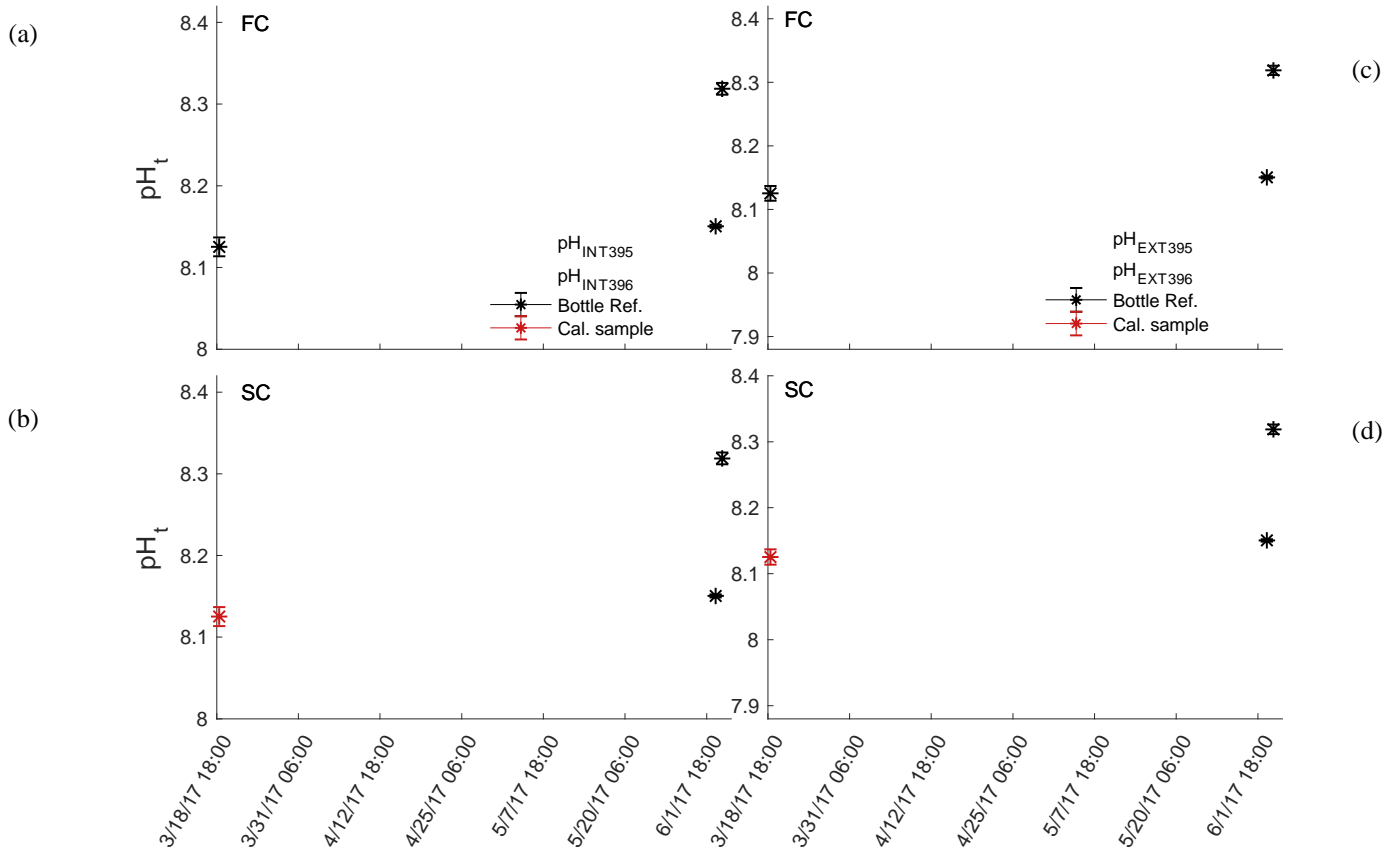
1051 **Figure 3.**  
1052



1053  
1054  
1055  
1056  
1057  
1058  
1059  
1060  
1061  
1062  
1063  
1064  
1065  
1066  
1067  
1068  
1069  
1070  
1071  
1072  
1073  
1074

Temperature derived from the internal thermistor on SeaFET<sup>TM</sup><sub>397</sub> (green circles) and the temperature recorded by the BoL (black circles) at the Alutiiq Pride Shellfish Hatchery from late winter through spring 2017. Salinity (red circles) recorded by the BoL on the right y-axis. SeaFET<sup>TM</sup><sub>397</sub> was only partially submerged resulting in the top half of the sensor exposed to air temperature fluctuations.

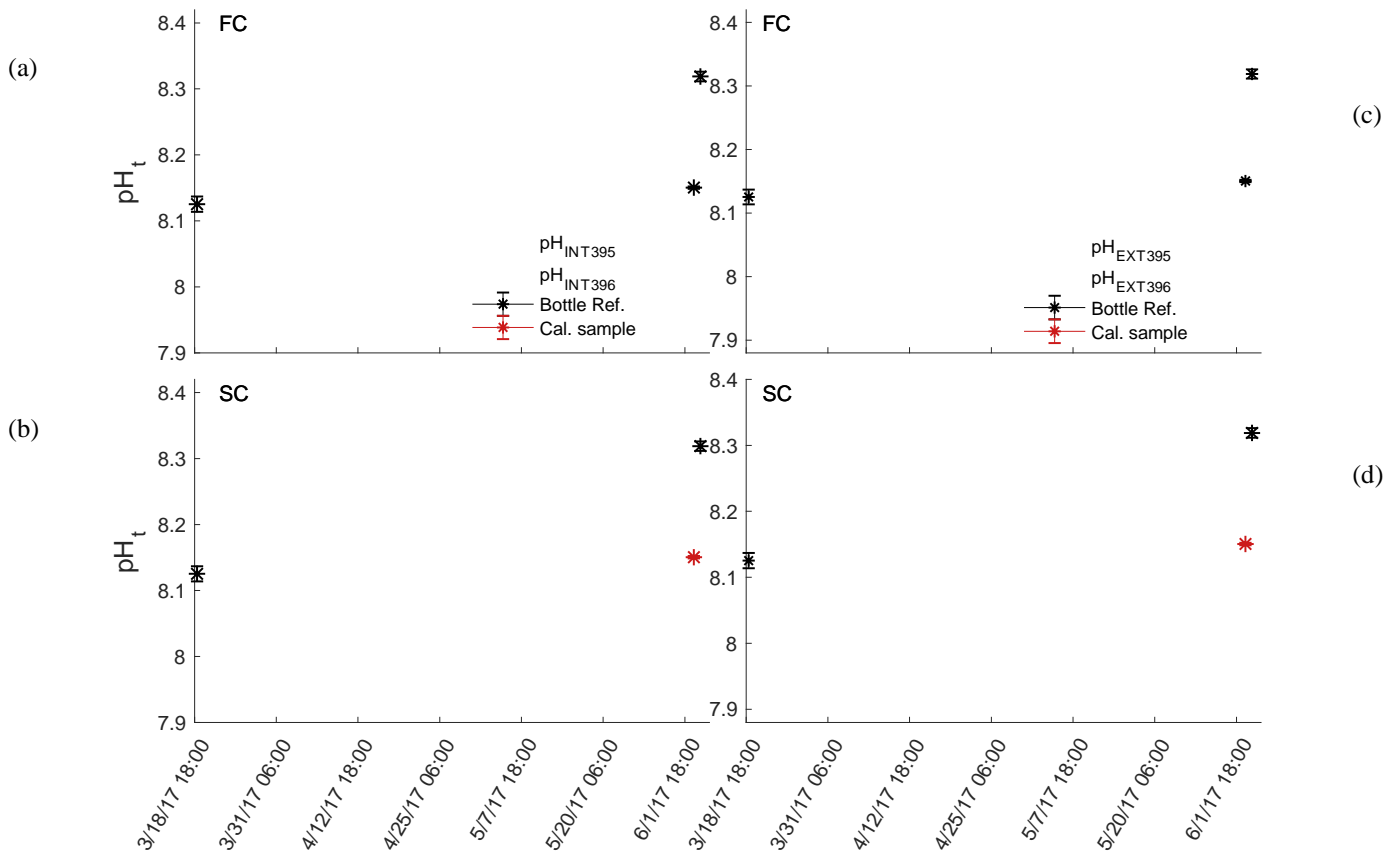
1075 **Figure 4.**  
 1076



1077  
 1078  
 1079 Comparison of pH<sub>t</sub> recorded by the internal (panel a and b) and external (panel c and d)  
 1080 electrodes on SeaFET<sup>TM</sup><sub>395</sub> (blue) and SeaFET<sup>TM</sup><sub>396</sub> (purple) before they were conditioned to the  
 1081 environment (non-conditioned) deployed in Kasitsna Bay, AK, based on calibration method:  
 1082 factory calibration (FC) and *in situ* single-point (SC) calibration. Discrete reference samples  
 1083 (black asterisks) and calibration sample (red asterisks) were collected 36 and 12 h pre-SeaFET<sup>TM</sup>  
 1084 recovery, and < 24 h post-deployment, respectively. Temperature and salinity measurements  
 1085 collected on reference and calibration samples were used to derive SeaFET<sup>TM</sup> pH<sub>t</sub> at those given  
 1086 time points. All other SeaFET<sup>TM</sup> pH<sub>t</sub> measurements use thermistor temperature and salinity  
 1087 logged by Kasitsna Bay data sonde.

1088  
 1089  
 1090  
 1091  
 1092  
 1093  
 1094  
 1095  
 1096  
 1097

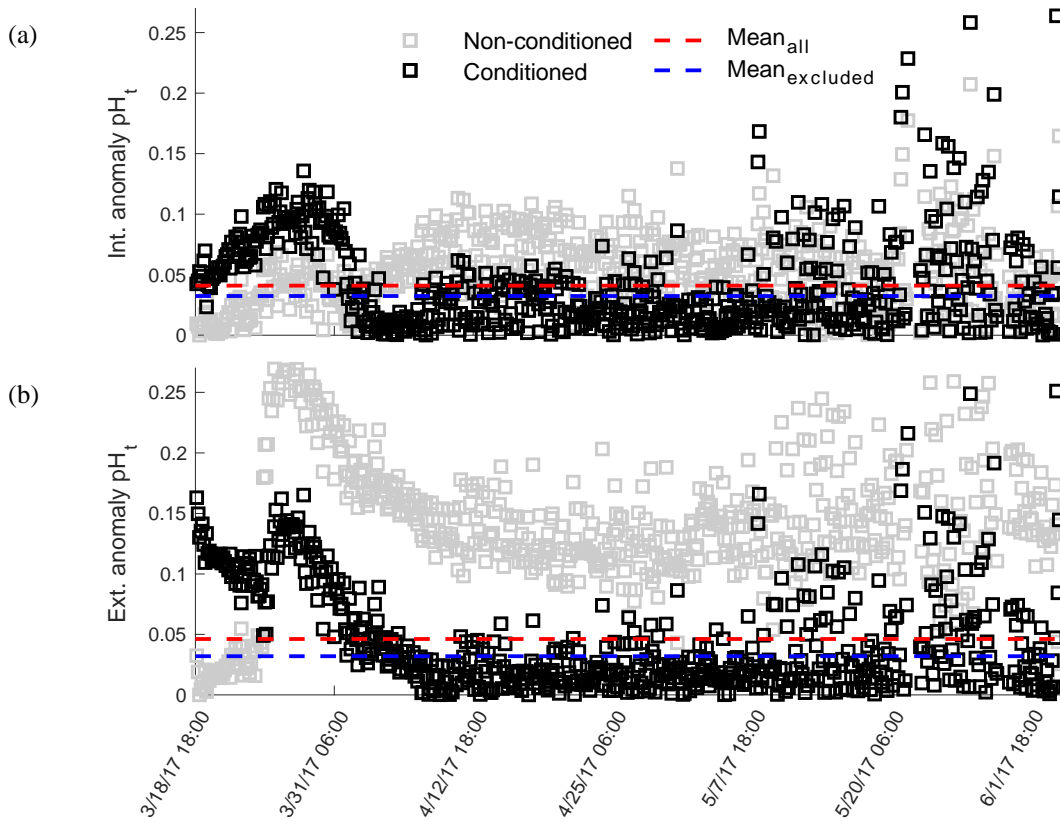
1098 **Figure 5.**  
 1099



1100  
 1101  
 1102 Comparison of pH<sub>t</sub> recorded by the internal (panel a and b) and external (panel c and d)  
 1103 electrodes on conditioned SeaFET<sup>TM</sup><sub>395</sub> (blue) and SeaFET<sup>TM</sup><sub>396</sub> (purple) deployed in Kasitsna  
 1104 Bay, AK, based on calibration method: factory calibration (FC) and *in situ* single-point (SC)  
 1105 calibration. The data set here is the same as figure 4, but timing of calibration method is  
 1106 different. Discrete reference samples (black asterisks) and calibration sample (red asterisks) were  
 1107 collected < 24 h post deployment and 12 h pre-SeaFET<sup>TM</sup> recovery, while calibration sample was  
 1108 collected 36 h pre-SeaFET<sup>TM</sup> recovery. Temperature and salinity measurements collected on  
 1109 reference and calibration samples were used to derive SeaFET<sup>TM</sup> pH<sub>t</sub> at those given time points.  
 1110 All other SeaFET<sup>TM</sup> pH<sub>t</sub> measurements use thermistor temperature and salinity logged by  
 1111 Kasitsna Bay data sonde.

1112  
 1113  
 1114  
 1115  
 1116  
 1117  
 1118  
 1119  
 1120

1121 **Figure 6.**  
1122



1123

1124

1125 Mean  $pH_t$  anomaly between *in situ* single-point calibrated SeaFET<sup>TM</sup><sub>395</sub> and SeaFET<sup>TM</sup><sub>396</sub>  
1126 internal (panel a) and external (panel b) electrodes during parallel deployment in Kasitsna Bay,  
1127 AK. Intra-anomaly comparison based on calibration sample taken at initial deployment (< 24 h  
1128 non-conditioned, gray squares) and end of deployment (36 h pre-recovery, black squares).  
1129 Shaded blue region indicates conditioning period. Data points in blue region omitted when mean  
1130 anomaly was calculated (non-conditioned: transparent blue-dashed line; conditioned: bold blue-  
1131 dashed line) compared to mean anomaly from entire data set (non-conditioned to environment:  
1132 red-dashed line; conditioned: red-dashed line).

1133

1134

1135

1136

1137

1138

1139

1140

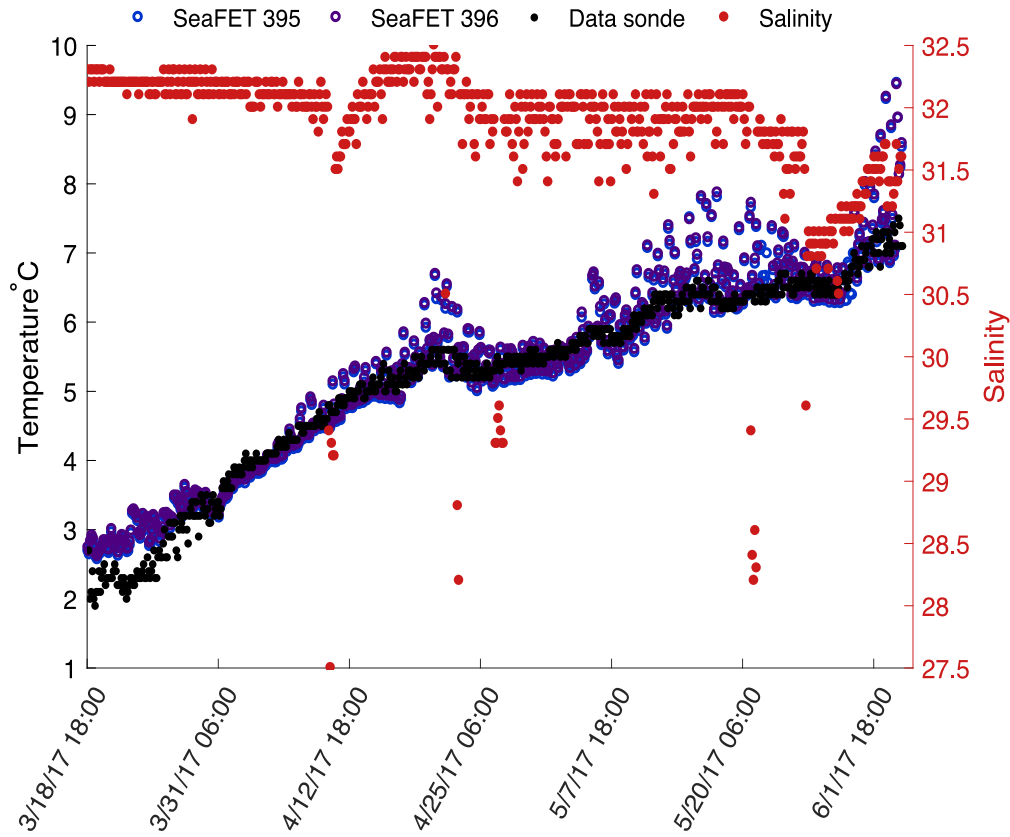
1141

1142

1143

1144

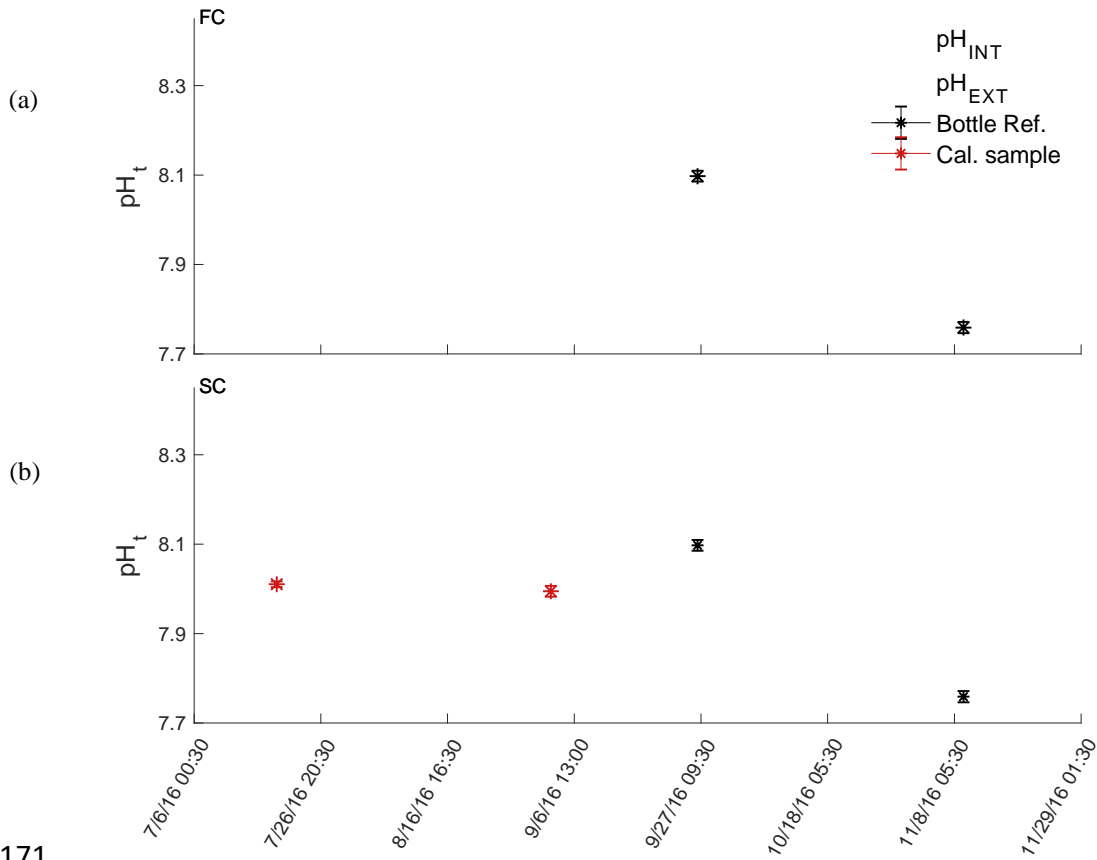
1145 **Figure 7.**  
1146



1147  
1148  
1149  
1150  
1151  
1152  
1153  
1154  
1155  
1156  
1157  
1158  
1159  
1160  
1161  
1162  
1163  
1164  
1165  
1166  
1167  
1168

Temperature derived from the internal thermistor on SeaFET<sup>TM</sup><sub>395</sub> (blue) and SeaFET<sup>TM</sup><sub>396</sub> (purple) compared against the temperature recorded by the Kachemak Bay National Estuarine Research Reserve data sonde. Salinity (Red circles) recorded by Kachemak Bay data sonde on the right y-axis.

1169 **Figure 8.**  
 1170



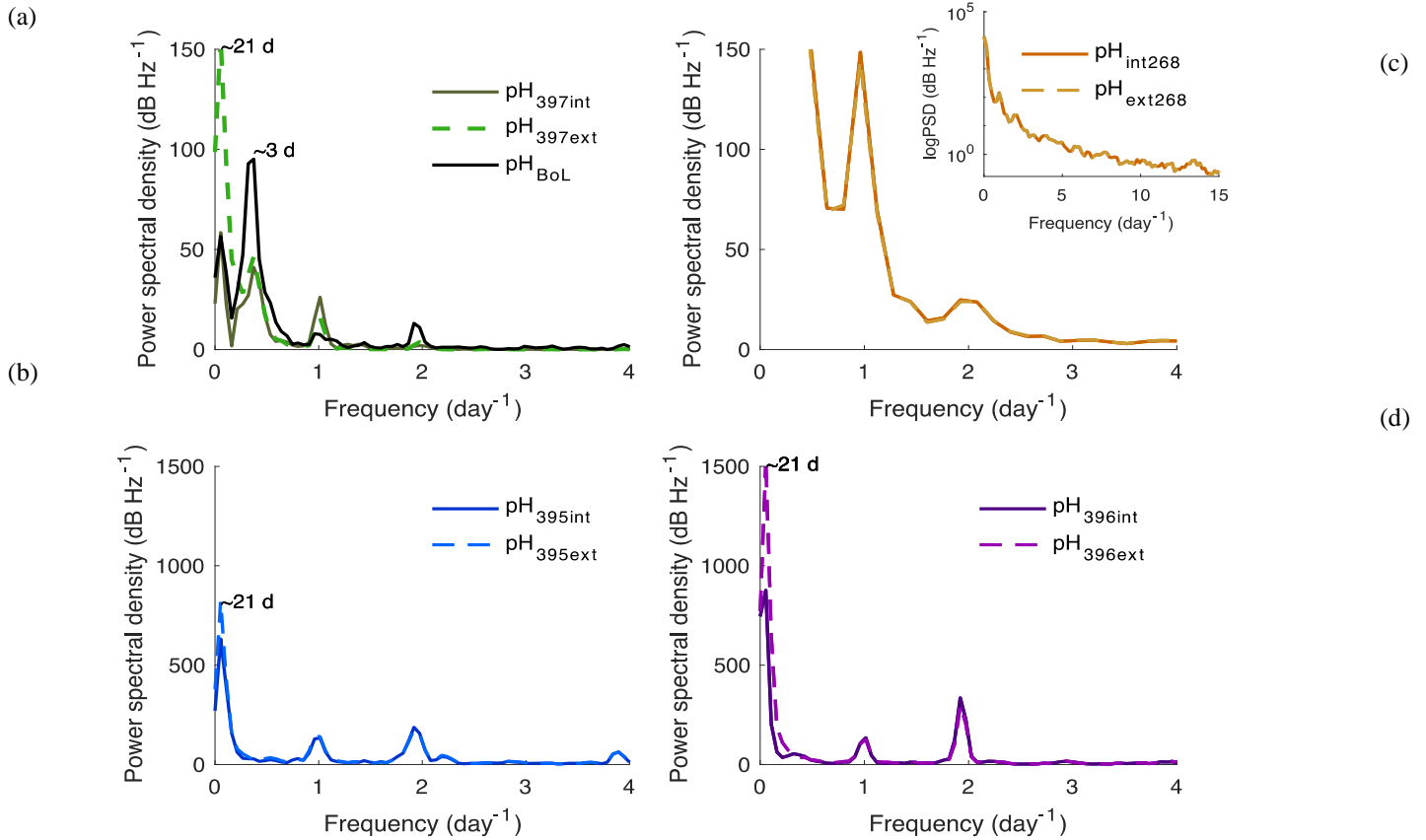
1171

1172  
 1173 pH<sub>t</sub> recorded by the internal (solid) and external (dashed) electrodes on SeaFET™<sub>268</sub> deployed at  
 1174 the Sentry Shoal mooring. pH<sub>t</sub> from both electrodes is shown when derived using factory  
 1175 calibration (FC) coefficients (panel a) and *in situ* single-point (SC) calibration coefficients (panel  
 1176 b). Black asterisks are references samples taken after initial calibration and recalibration (red  
 1177 asterisk), where pH<sub>t</sub> was derived from TCO<sub>2</sub> and pCO<sub>2</sub> measurements made on the BoL at the  
 1178 Hakai Institute's Quadra Island Field Station.

1179  
 1180  
 1181  
 1182  
 1183  
 1184  
 1185  
 1186  
 1187  
 1188  
 1189  
 1190  
 1191  
 1192



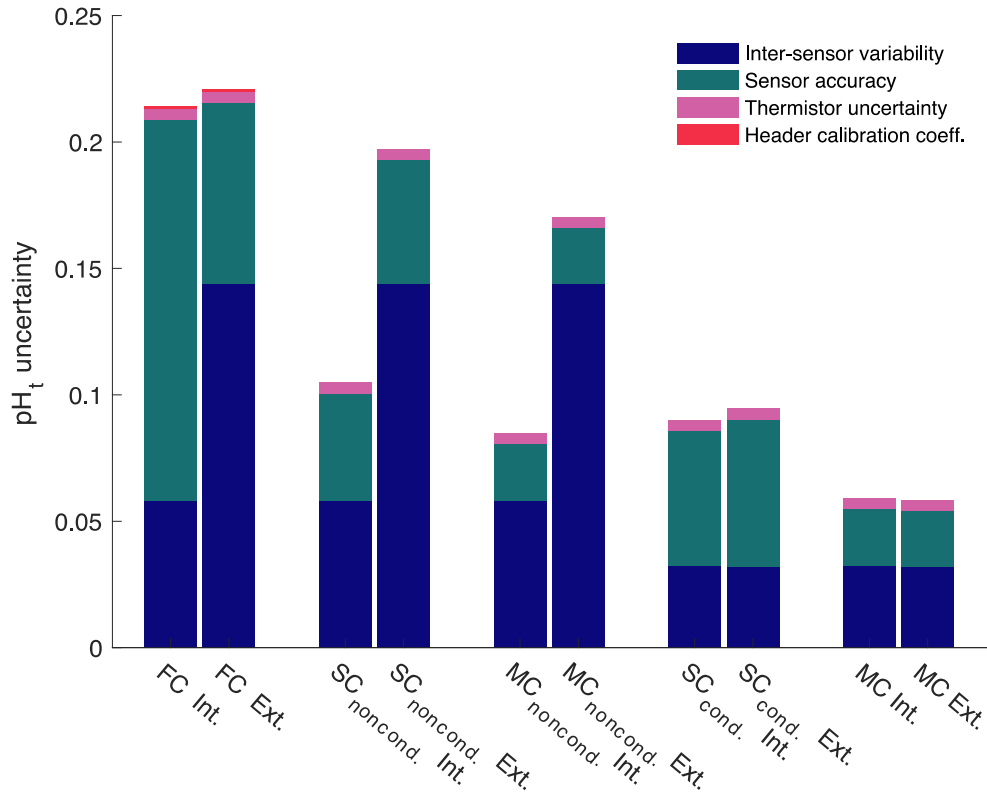
1193 **Figure 9.**  
 1194



1195  
 1196  
 1197  
 1198  
 1199  
 1200  
 1201  
 1202  
 1203  
 1204  
 1205  
 1206  
 1207  
 1208  
 1209  
 1210  
 1211  
 1212  
 1213  
 1214  
 1215

Power spectral density (PSD) analysis of  $\text{pH}_i$  in frequency per day for SeaFETs™ 397 (panel a), 268 (panel b), 395 (panel c), and 396 (panel c). Inset in panel b is log base 10 transformed PSD analysis of same data set. All internal electrodes marked as solid colored lines while external electrodes are colored dashed lines. BoL data set marked as solid black line (panel a).

1216 **Figure 10**  
 1217



1218  
 1219

1220 Quantified uncertainties based on field deployments of all Sea-Bird SeaFETs™ separated by  
 1221 electrode calibration method (FC: factory; SC: single-point; MC: multi-point), and calibration  
 1222 time for SeaFETs™ 395 and 396 (i.e., non-conditioned to environment and conditioned). pH<sub>t</sub>  
 1223 accuracy uncertainty calculated as the mean difference when comparing the absolute difference  
 1224 between reference samples and SeaFETs™ 395 (non-conditioned to environment and  
 1225 conditioned), 396 (non-conditioned to environment and conditioned), and 268 as well as the  
 1226 average absolute difference between SeaFET™ 397 and the BoL. Inter-sensor variability  
 1227 uncertainty determined by comparing SeaFETs™ 395 (non-conditioned to environment and  
 1228 conditioned) and 396 (non-conditioned to environment and conditioned), deployed side-by-side  
 1229 in Kasitsna Bay. Thermistor uncertainty is calculated pH<sub>t</sub> error when using thermistor derived  
 1230 temperature rather than external temperature sensor determined from SeaFETs™ 395 and 396.  
 1231 Header calibration coefficient uncertainty is the discrepancy in pH<sub>t</sub> when using SeaFETcom  
 1232 factory calibration coefficients from header file rather than disc file.

1233  
 1234