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The Pelagic In situ Observation System (PELAGIOS) to reveal biodiversity, behavior and ecology of elusive oceanic fauna

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1. Abstract

There is a need for cost-efficient tools to explore deep ocean ecosystems to collect baseline biological observations on pelagic fauna (zooplankton and nekton) and establish the vertical ecological zonation in the deep sea. The Pelagic In situ Observation System (PELAGIOS) is a 3000 m-rated slowly (0.5 m/s) towed camera system with LED illumination, an integrated oceanographic sensor set (CTD-O_2) and telemetry allowing for online data acquisition and video inspection (Low Definition). The High Definition video is stored on the camera and later annotated using the VARS annotation software and related to concomitantly recorded environmental data. The PELAGIOS is particularly suitable for open ocean observations of gelatinous fauna, which is notoriously undersampled by nets and/or destroyed by fixatives. In addition to counts, diversity and distribution data as a function of depth and environmental conditions (T, S, O_2), in situ observations of behavior, orientation and species interactions are collected. Here we present an overview of the technical setup of the PELAGIOS as well as example observations and analyses from the eastern tropical North Atlantic. Comparisons to MOCNESS net sampling and data from the Underwater Vision Profiler are provided and discussed.

2. Introduction

The open ocean pelagic zones include the largest, yet least explored habitats on the planet (Robison, 2004; Webb et al., 2010; Ramirez-Llodra et al., 2010). Since the first oceanographic expeditions, oceanic communities of macrozooplankton and micronekton have been sampled using nets. Such sampling has revealed a community typically consisting of crustaceans, cephalopods, fishes and some sturdy and commonly found gelatinous fauna. Underwater observations in the open ocean via SCUBA diving (Hamner et al., 1975) and later via submersibles

(Robison, 1983; Robison and Wishner, 1990) and in situ camera systems (Biard et al., 2016, Picheral et al. 2010) revealed that a variety of organisms are much more abundant in the open ocean than previously estimated from net sampling (Robison, 2004, Haddock, 2004; Biard et al. 2016, Christiansen et al. 2018). This was particularly true for fragile gelatinous zooplankton, a diverse taxonomic group of different phyla, including the ctenophores, medusae, siphonophorae, thaliaceans, polychaetes, rhizaria and larvaceans, which often are too delicate to be quantified using nets as they are damaged beyond identification, or they are easily destroyed by the use of common fixatives.

Underwater (*in situ*) observations in the pelagic ocean not only revealed a previously unknown community, they also allowed the collection of fine scale distribution patterns, information on posture, interactions, and behavior and a better understanding of the ecological context and role of pelagic organisms (Hamner and Robison, 1992; Robison, 2004; Robison, 1999; Biard et al., 2016; Hoving et al., 2017). Submersibles have proven to be valuable instruments to study deep-sea pelagic biology (e.g. Robison, 1987; Bush et al., 2007; Hoving et al., 2013; 2016). Using video transecting methodology, pelagic ROV surveys have been applied to study inter and intra-annual variation in mesopelagic zooplankton communities (Robison et al., 1998) and to explore deep pelagic communities in different oceans (Youngbluth et al., 2008; Hosia et al., 2017; Robison et al., 2010). However, due to high costs as well as technological and logistical challenges, regular submersible operations are still restricted to very few institutes and geographical locations. Hence, there is a need for the development of additional more cost-effective methodologies to explore and document deep-sea communities via in situ observations.

In the last decades, a variety of optical instruments has been developed to image and quantify plankton *in situ*. The factors that typically differentiate the available plankton imaging technologies are the size fraction of the observed organisms, illumination type, resolution of collected images/video, depth rating, deployment mode (e.g., autonomous, towed, CTD-mounted) and towing speed (Benfield et al., 2007). Examples of instruments include the autonomous Underwater Vision Profiler (UVP) (Picheral et al., 2010), the Lightframe On-sight Key species Investigations (LOKI; Schulz et al 2009) and towed plankton recorders (ISiiS; Cowen and Guigand 2008; for review see Benfield et al 2007). These instruments can be deployed from ships of opportunity and collect detailed information on fine scale distribution and diversity patterns of particles and plankton. The data reveal biological patterns on a global scale (Kiko et al., 2017) and of previously underappreciated plankton species (Biard et al., 2016).

Various towed camera platforms have been developed that can obtain video transect observations above the deep sea floor. Examples are the TowCam (WHOI), the DTIS (Deep Towed Imaging system, NIWA), the WASP vehicle (Wide Angle Seafloor Photography), OFOS (Ocean Floor Observation System, GEOMAR), and the more recent version OFOBS (Ocean Floor Observation and Bathymetry System) (Purser et al., 2018). All these instruments are used for video or photo transects of the seafloor, with a downward looking camera, and typically a set of lasers for size reference. However, published descriptions of optical systems, other than ROVs and submersibles, that visualize macrozooplankton and micronekton (>1 cm) in the water column are, to the best of our knowledge, restricted to one (Madin et al., 2006). The Large Area Plankton Imaging System (LAPIS) is the only towed system that was developed for the documentation of larger organisms in the water column (Madin et al., 2006). LAPIS visualizes organisms between 1 and 100 cm, it combines a low light camera with red illumination, and it is towed at 1 knot via a conducting wire. Deployments in the Southern Ocean enabled the reconstruction of depth distributions of the pelagic fauna (salps, medusae) but also allowed behavior observations, e.g. the moulting of krill (Madin et al., 2006).

To establish a baseline in abundance, distribution and diversity of the pelagic fauna in its natural environment, we developed an ocean observation platform for pelagic video transects. The functional requirements for the instrument were the ability to: (1) visualize organisms > 1 cm in waters down to 1000 m, (2) deploy the instrument from ships of opportunity in an autonomous or transmitting mode, (3) make it light and practical so it can be deployed easily and safe with 2 deck persons and a crane operator, (4) enable correlation of observations with environmental parameters (S, T, O_2) and other sensor data, and (5) make observations comparable to ROV video transects in other reference areas. We present a description of the Pelagic In situ Observation System (PELAGIOS), examples of the kind of biological information it may gather, as well as biological discoveries that have resulted from deployments on research cruises in the eastern tropical Atlantic.

3. Pelagic In Situ Observation System

3.1 Technical Specifications

The PELAGIOS consists of an aluminum frame (length = 2 m) that carries the oceanographic equipment (Figure 1). White light LED arrays (4 LEDs produced at GEOMAR, 2 LED arrays type LightSphere of Deep-Sea Power and Light ©) which illuminate the water in front of the system are mounted on an aluminum ring (diameter = 1.2 m). Power is provided by two lithium batteries (24V; 32 Ah) in a deep-sea housing. High-definition video is collected continuously by a forward viewing deep-sea camera (type 1Cam Alpha, SubC Imaging ©) which is mounted in the center of the ring. We used the maximum frame rate of 50 frames s^{-1} but a lower frame rate is possible. A

CTD (SBE 19 SeaCAT, Sea-Bird Scientific) with an oxygen sensor (SBE 43, Sea-Bird Scientific) records environmental data. A deep-sea telemetry (DST-6, Sea and Sun Technology ©; Linke et al., 2015) transmits video and CTD data to a deck unit on board allowing a low resolution preview (600x480 lines) of the high definition video that is stored locally on the SD card (256 GB) of the camera. The power from the batteries is distributed to the LEDs via the camera. The 1Cam Alpha camera is programmable in such a way that there is a delay between providing power to the camera (by connecting to the battery) and the start of recording and switching on the LEDs. This enables the illumination to be turned on only underwater, and prevents overheating of the LED arrays while out of the water. During a cruise with the German research vessel MARIA S. MERIAN (MSM 49) we mounted a steel scale bar in front of the camera at a distance of 1 m. The distance between the centers of the white marks on the bar measured 5 cm.

3.2 Video transects

The PELAGIOS is towed horizontally at specified depths of 20-1000 m. The standard towing speed over ground is 1 knot (0.5 m/s), and the speed is monitored via the ship's navigational system. A video transect at a particular depth can take as long as desired and is terminated by lowering the PELAGIOS to the next desired depth. Maximum deployment time with full batteries is approximately 6 hours. The typical transect duration is 10-30 min. The depth of the PELAGIOS can be monitored via online CTD data. Figure 2 shows the trajectories of the PELAGIOS at different depths in the water column during a video transect down to 700 m. The deployment from deck into the water and the reverse is fast and typically takes only about 5 min (see video clip: https://www.wissenschaftsjahr.de/2016-17/das-wissenschaftsjahr/die-forschungsflotte/forschungsschiff-blogs/unerforschte-meeresgebiete.html). It is possible to deploy PELAGIOS in 'blind mode', where only the depth is monitored using an online depth sensor (e.g., Hydrobios ©) and the video (without transmitted preview) is recorded locally on the camera. The system can be operated completely blind (i.e., with no communication between deck and underwater unit) where the target depth is estimated from the length and angle of the wire put out, and the actual depth is recorded on the system by an offline pressure sensor e.g. SBE Microcat ©.

3.3 Video analysis and curation

After a deployment, the video (consisting of individual clips of one hour) is downloaded from the camera. Synchronisation between video and CTD data is done by setting all instruments to UTC prior to deployment, which allows the data and video to be linked during analysis. The video is annotated using the Video Annotation and Reference System VARS developed by the Monterey Bay Aquarium Research Institute (Schlining and Jacobsen, 2006). This annotation program allows for frame grabs from the video including time code. A Knowledge Base allows for inserting taxonomic names and hierarchy, and a Query allows for searching the created database. While many kinds of annotation software are available (for review see Gomes-Pereira et al., 2016), we consider VARS the most suitable for our purposes since it combines the features of high resolution video play with a user friendly annotation-interface and the automatic creation of an annotation database which can easily be accessed through the various search-functions and tools of the Query. The taxonomic hierarchy and phylogenetic trees in the database are directly applicable to our video transects. Since this software was developed by MBARI, which also maintains the most extensive databases of deep pelagic observations, it makes communication about and comparison of observations and data practical. Videos are transported on hard drives after an expedition. At GEOMAR, videos are transferred for long term storage on servers maintained by the central data and computing centre at GEOMAR, providing instant access to videos and images with metadata description via the media server ProxSys.

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3.4 Sample volume

To estimate the sample volume of the PELAGIOS we compared video counts from the PELAGIOS with concomitantly obtained abundance data from an Underwater Vision Profiler (UVP5; Picheral et al., 2010). Four deployments from the R/V Maria S. Merian cruise MSM 49 (28.11. - 21.12.2015, Las Palmas de Gran Canaria/Spain – Mindelo/Cape Verde) were used for the comparison where a UVP5 was mounted underneath the PELAGIOS (Figure 1). The UVP5 takes between 6-11 images per second of a defined volume (1.03 L) and thus enables a quantitative assessment of particle and zooplankton abundances. Objects with an equivalent spherical diameter (ESD) >0.5 mm are saved as images, which can be classified into different zooplankton, phytoplankton and particle categories. For the comparison between PELAGIOS and the UVP5, we used the pelagic polychaete *Poeobius* sp., as 1) this organism could be observed well on both instruments, 2) *Poeobius* sp. is not an active swimmer and 3) it was locally very abundant, thus providing a good basis for the direct instrument comparison.

The UVP5 images were classified as described in Christiansen et al. (2018). *Poeobiu*s sp. abundance (ind m^{-3}) was calculated for 20 s time bins and all bins of one distinct depth step averaged. These mean abundances were compared to the PELAGIOS counts (ind s^{-1}) of the same depth step. A linear model between the PELAGIOS counts as a function of UVP5 abundance provided a highly significant relationship (linear regression: $p < 0.001$, *adjusted* $r^2 = 0.69$; Figure 3). The linear regression slope b (0.116 $m^3\ s^{-1}$, standard error 0.01 $m^3\ s^{-1}$) between the PELAGIOS-based count (C_{PELAGIOS}, ind s^{-1}) and mean UVP-based abundance (A_{UVP}, ind m^{-3}):

$$C_{PELAGIOS} = b * A_{UVP} + a \quad \text{(Equation 1)}$$

was used to estimate the volume recorded per time in m^3 s^{-1} (*b*) and the field of view in m^2 (*b*/towing speed) recorded by PELAGIOS.

From this calculation it can be derived that PELAGIOS recorded an average volume of 0.116 m^3 s^{-1} at a towing speed of 1 knot (= 0.5144 m s^{-1}). A cross-sectional view field of approximately 0.23 m^2 of PELAGIOS can be expected.

3.5 Abundance, size and diversity at an example station "Senghor NW"

To provide an example of the type of data that can be obtained with the PELAGIOS, we report here on day and night video transects down to 950 m in the Eastern Tropical North Atlantic, on the northwestern slope of Senghor Seamount (17°14.2'N, 22°00.7'W; bottom depth of approximately 1000 m). The results from the video annotations show that faunal abundances depend on the depth of deployment, and time of the day. During two transects of 11 minutes at 400 m, 232 individuals were encountered during the day (the three dominant organism groups are fish, euphausiids and appendicularians) compared to 208 individuals during the night (the four dominant organism groups are fishes, chaetognaths, medusae and ctenophores). Overall abundance of chaetognaths, decapods and mysids, and somewhat for fishes was higher during the night. The peak of euphausiids' abundance at 400 m shifts to the surface at night (Figure 4). The higher abundance of decapods, mysids and chaetognaths at night may indicate lateral migration or daytime avoidance. The vertical migration that was observed for fishes and crustaceans was much less clear for the gelatinous zooplankton groups including the medusae and appendicularians (Figure 4). Ctenophores and siphonophores were abundant in the surface at night and the thaliaceans migrated vertically and were most abundant in shallow waters at night. The total number of annotated organisms for the daytime transects (total transect time 182 minutes; max. depth 950 m) was 835 compared to 1865 organisms for the nighttime transects. Remarkable is the enormous abundance of gelatinous zooplankton (129 annotated organisms belonging to the three dominant groups of Ctenophora (53), Siphonophorae (32) and Thaliacea (44) in the topmost layer (20 m) at night. Below this layer, the depth profile shows a minimum in numbers of annotated individuals at 100, 200, and 300 m water depth with a smaller peak of 56 gelatinous organisms in 450 m. Compared to this, the depth distribution at day time shows a more regular, almost Gaussian shape with a maximum of 47 and 54 gelatinous organisms at 200 and 400 m water depth, respectively.

The faunal observations at station Senghor NW include a wide variety of taxa (Table 1). The smallest annotated specimens belonged to the radiolarians. Chaetognaths were the dominant faunal group. Large larvaceans tentatively identified to belong to the genus *Bathochordaeus* and *Mesochordaeus* were also observed. Pelagic polychaetes of the genus *Poeobius* can be easily distinguished and are up to 23 mm long (Christiansen et al., 2018). Other pelagic worms are tomopterid and alciopid worms, the latter can reach 1 m in length. The faunal group with the largest specimens, attaining up to several metres in length, are the siphonophores, including *Praya dubia* and *Apolemia*. Siphonophores of the genus *Bargmannia* and *Lilyopsis* were also observed. *Lilyopsis* can be easily distinguished by their fluorescent body parts. Observed medusae belonged to the genera *Periphylla*, *Halitrephes*, *Haliscera*, *Crossota,* *Colobonaema*, *Solmissus* and *Solmundella*. Lobate ctenophores such as *Thalassocalyce inconstans*, *Leucothea*, *Bathyceroe* are typical examples of organisms that cannot be captured by nets but which can be properly quantified by PELAGIOS. Venus girdles (*Cestum* spp.), *Beroe* and cydippids are other ctenophores that were encountered at Senghor NW. Cephalopod observations are rare but small individual cranchid squids were observed in the upper 50 m at night. Mastigoteuthid squids were observed with their mantle in a vertical orientation and with extended tentacles in waters below 500 m. One *Taningia danae* was observed during a transit between transecting depths. Other pelagic molluscs include the nudibranch *Phylliroe* and different pteropod species. Observed fishes are snipe eels, hatchet fishes, lantern fishes and *Cyclothone*. Fishes are among the dominant organisms encountered during PELAGIOS transects but it is often impossible to identify fishes to species level from the video.

We compared PELAGIOS video transects with MOCNESS net (opening 1 m^2) abundance data by integrating the PELAGIOS counts over the respective depth strata of the MOCNESS. The diversity of the gelatinous zooplankton in the total MOCNESS catch is much lower (8 different taxa) than in the pooled video transects (53 annotated taxa) on the same station. The ctenophore *Beroe* is captured in MOCNESS hauls and also observed on PELAGIOS transects. Normalization and subsequent standardization of the encountered *Beroe* in MOCNESS and PELAGIOS transects shows that on the same station and the same depths, PELAGIOS observes 3.3-4.7 times more *Beroe* at the three depths where they were encountered by both instruments. Additionally, the PELAGIOS also repeatedly observed *Beroe* at depths where they were not captured by MOCNESS at all (although there were also depths where PELAGIOS did not observe any *Beroe*).

3.6 Individual behaviour

In situ observations by PELAGIOS video may reveal direct observations on individual behavior. Decapod shrimps were observed to release a blue or green bioluminescent cloud after performing their tail flip as part of the escape response (Figure 5). Potential reproductive behavior was observed for two specimens of krill which were seen in a mating position, and salps were observed to reproduce asexually by the release of salp oozoids (Figure 5). Feeding behaviors were observed for large prayid siphonophores and calycophoran siphonophores which had their tentacles extended. *Poeobius* worms were observed with their mucus web deployed to capture particulate matter (Christiansen et al., 2018). Narcomedusae of the genus *Solmissus* were observed with their tentacles stretched up and down, which is a feeding posture (Figure 5). In situ observations by the PELAGIOS also showed the natural body position of pelagic organisms. Snipe eels were observed in a vertical position with their heads up, while dragonfishes and some myctophids were observed in an oblique body position with their head down.

4. Discussion

PELAGIOS is a cost-effective pelagic ocean exploration tool that fills a gap in the array of observation instruments that exist in biological oceanography. The instrument can be deployed with a small team and from vessels of opportunity, in transmission or blind mode. The relatively simple design limits technical failures and makes the PELAGIOS a reliable tool for oceanic expeditions. While thus far the system has only been deployed in the open ocean, it can be used in any pelagic environment with water that has reasonable clearance and visibility. The data obtained after annotation of the video can be uploaded into databases (e.g., Pangaea) after publication of the results allowing for efficient data sharing and curation.

The clear signal of the vertical migration in some animal groups (fishes, crustaceans) that we observed during the video transects confirms that established biological processes can be detected in PELAGIOS data, and that the distribution data that we observe for encountered organisms are representative for the natural situation. It has to be noted, though, that while the observed distribution patterns should be representative, care must be taken with regards to abundance estimates of especially actively- and fast-swimming organisms. Some fish and crustaceans react to the presence of underwater instrumentation (e.g. Stoner et al. 2008). Gear avoidance (e.g. Kaartvedt et al. 2012) can lead to an underestimation of abundance, whereas attraction to the camera lights (e.g. Utne-Palm et al. 2018, Wiebe et al. 2004) would result in an overestimation. The large bioluminescent squid *Taningia danae* seemed to be attracted to the lights of the PELAGIOS. Compared to day transects, the high abundance of gelatinous organisms close to the surface during night is likely to be partly an effect of the higher contrast in the videos of the night transects and better visibility of the gelatinous fauna than during day transects. Many of the observed gelatinous fauna might be as well be present at shallow depths during day-light but are not detectable at 'blue-water-conditions'. The large difference between encountered taxa during the day and night transect may also be explained by the lateral migration of animals towards Senghor seamount at night. However, from a methodological side it should be noted that while the ship's towing speed is typically 1 knot, the current speeds at the survey depths may differ, also between day and night. Currents may result in more or less sampled volume of water and hence a variation in plankton being visualized. Therefore it is recommended to perform future surveys with a current meter to measure the speed through water.

After annotation, the PELAGIOS video transects may be used to reconstruct species-specific distribution patterns, which can be related to environmental gradients. Such data is valuable for studies on overlap in distribution patterns of consumers and food items (e.g. *Poeobius* and particles, ctenophores and krill). The data can also be used in biological studies that aim to predict the consequences of a changing ocean with altering environmental gradients for species' distributions. One example of changing environmental gradients is the global trend of oxygen loss in the world oceans. Oxygen minimum zones (OMZ) are occurring naturally in the mesopelagic zone, and in different oceans they have been found to expand horizontally and vertically as a result of climate change (Stramma et al., 2008). Expansion of OMZs may result in a habitat reduction of the pelagic fauna (e.g., Stramma et al., 2012), or increase the habitat for species with hypoxia tolerance. To predict the potential consequences of OMZ expansion for pelagic invertebrates we investigated the abundance and distribution of distinct large gelatinous zooplankton species, including medusae, ctenophores, siphonophores and appendicularians, in the eastern tropical Atlantic using PELAGIOS video transects and correlated the biological patterns to the oxygen gradients (Neitzel, 2017; Hoving et al., in prep.).

Preliminary comparisons of the data obtained with PELAGIOS and with MOCNESS indicate substantial differences in the documented fauna. Many more gelatinous taxa were observed during PELAGIOS video transects than were captured in MOCNESS catches at the same station, with the exception of the small and robust calycophoran colonies of the families Diphyidae and Abylidae. This discrepancy is likely the result of the delicate nature of many ctenophores, medusae and siphonophores, preventing their intact capture by nets. Additionally, avoidence behavior of strong swimming jellyfish (e.g. *Atolla, Periphylla*), which escape from the relatively slow moving PELAGIOS, may explain their increased occurrence in nets compared to video recordings. While PELAGIOS is certainly suitable for visualizing delicate gelatinous fauna, it cannot replace net-sampling since complementary specimen collections are needed to validate the identity of organisms that were observed during PELAGIOS video observations. Therefore, it is desired that net tows with open and closing nets such as Multinet Maxi or MOCNESS are performed in the same areas, or that collections during submersible dives are made.

The potential of the PELAGIOS as an exploration tool is illustrated by the discovery of previously undocumented animals. An example is the ctenophore *Kiyohimea usagi* (Matsumoto and Robison,

1992) which was observed seven times by the PELAGIOS and once by the manned submersible JAGO during cruises in the eastern tropical Atlantic. This large (>40 cm wide) lobate ctenophore was previously unknown from the Atlantic Ocean and demonstrates how in situ observations in epipelagic waters can result in the discovery of relatively large fauna (Hoving et al., submitted). Since gelatinous organisms are increasingly recognized as vital players in the oceanic food web (Choy et al., 2017) and in the biological carbon pump (Robison et al., 2005), in situ observations with tools like the PELAGIOS can provide new important insights into the oceanic ecosystem and the carbon cycle. But small gelatinous organisms may also have a large biogeochemical impact on their environment. This was illustrated by the discovery of the pelagic polychaete *Poeobius* sp. during the PELAGIOS video transects in the eastern Atlantic (Christiansen et al., 2018). The observations of the PELAGIOS provided the first evidence for the occurrence of *Poeobius* sp. in the Atlantic Ocean. During the R/V Meteor cruise M119, *Poeobius* was found to be extremely abundant in a mesoscale eddy. Using an extensive database of the UVP5 (956 vertical CTD/UVP profiles) in the eastern tropical Atlantic it was possible to reconstruct the horizontal and vertical distribution of Atlantic *Poeobius* in great detail and to establish that the high local abundance of *Poeobius* was directly related to the presence of mesoscale eddies in which they possibly intercepted the entire particle flux that was on the way to the deep sea (Christiansen et al., 2018; Hauss et al., 2016).

During various cruises, the UVP 5 was mounted underneath the PELAGIOS providing concomitant data on macrozooplankton and nekton (PELAGIOS) as well as particles and mesozooplankton (UVP). The combination of the two instruments provides a great opportunity to assess both the mesopelagic fauna and particles during one sampling event. The joint deployment of the PELAGIOS and UVP also allowed a quantification of the sampled water volume of the

PELAGIOS as described above. The linear relationship between counts of the non-moving *Poeobius* sp. with UVP5 and the PELAGIOS indicates comparability of the two different methods and provides a correction factor to estimate organism abundance (ind m^{-3}) from PELAGIOS count (ind s^{-1}) data. The field of view (FOV) for the PELAGIOS was estimated to be 0.23 m^2. The angle of view of the PELAGIOS is 80° and therefore the field of view (FOV) is much smaller than the FOV of video transects with a wide-angle lens by ROV Tiburon (Robison et al., 2010). When comparing the FOV, it is important to take into account the object that is observed. We provided an estimate of the FOV using *Poeobius* sp., which is a small organism that can be detected only when it is close to the camera. Therefore, the area of the FOV for quantification of *Poeobius* sp. is smaller than when quantifying larger organisms, and the initial identification distance differs between species (Reisenbichler et al., 2017). Future effort should be focused on improving the assessment of the sample volume by integrating technology that can quantify it (e.g. current meters, a stereo-camera setup or a laser-based system). A stereo-camera set up would also allow for size measurements of the observed organisms, which could be beneficial to estimate the biomass of the observed organisms from published size-to-weight relationships. It might also be possible to obtain similar information based on structure-from-motion approaches that proved successful in benthic video imaging (Burns et al., 2015). The PELAGIOS system can also be a platform for other sensors. The PELAGIOS was used to mount and test the TuLUMIS multispectral camera (Liu et al., 2018). Future developments include the preparation of the system for deployments down to 6000 m. The integration of acoustic sensors would be valuable to measure target strength of camera observed organisms, to estimate gear avoidance or attraction and to estimate biomass and abundance of organisms outside the field of view of the camera.

Author contribution

This instrument was designed, tested and applied by Henk-Jan Hoving and Eduard Fabrizius. Rainer Kiko and Helena Hauss developed the idea of combining the PELAGIOS with the UVP5. Philipp Neitzel and Svenja Christiansen analyzed the data in this manuscript in consultation with Henk-Jan Hoving, Rainer Kiko and Helena Hauss. Arne Körtzinger, Uwe Piatkowski and Peter Linke added valuable input to the further development of the instrument and its application and/or the data interpretation. All authors contributed to writing the paper. All authors approved the final submitted manuscript.

Data availability

The datasets generated and/or analysed during the current study will be available in the Pangaea repository. A link will be provided when the paper is accepted.

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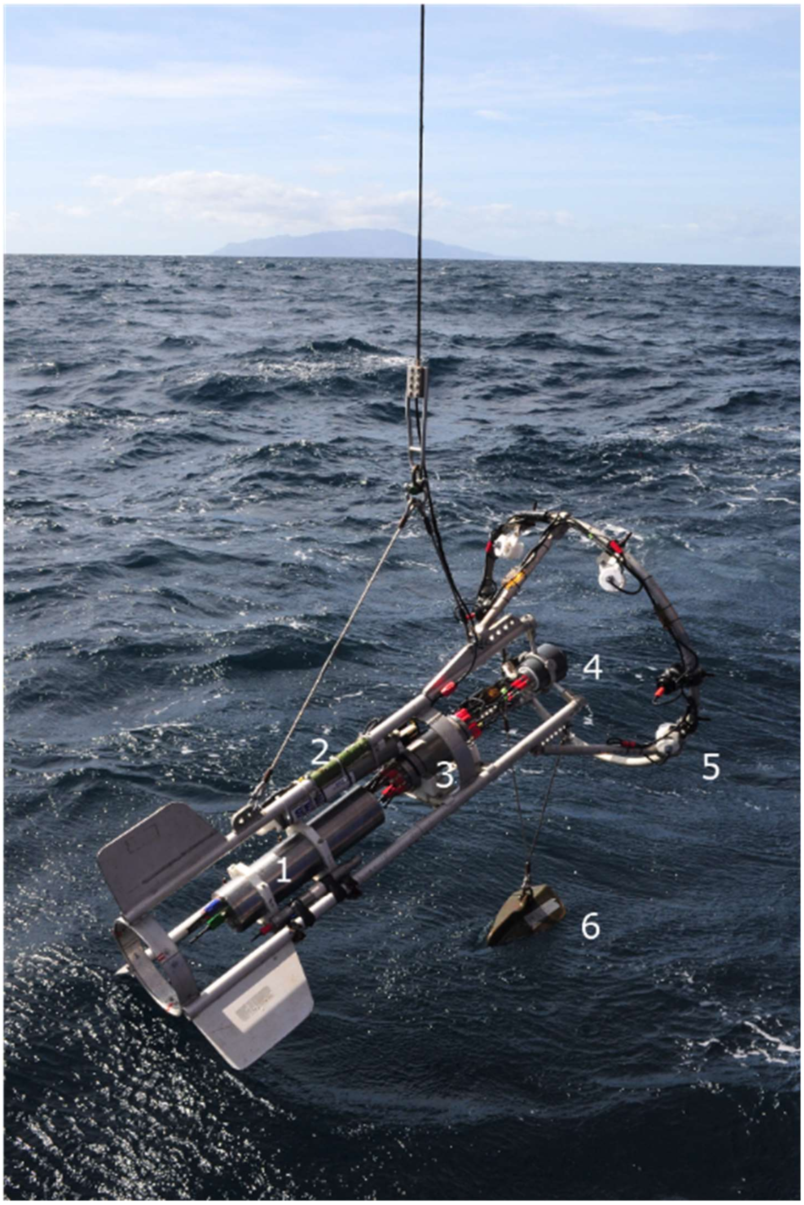

Figure 1: a) The Pelagic in situ observations system with the battery (1), CTD (2), telemetry (3), camera (4), LEDs (5), depressor (6), during deployment from R/V POSEIDON in February 2018.

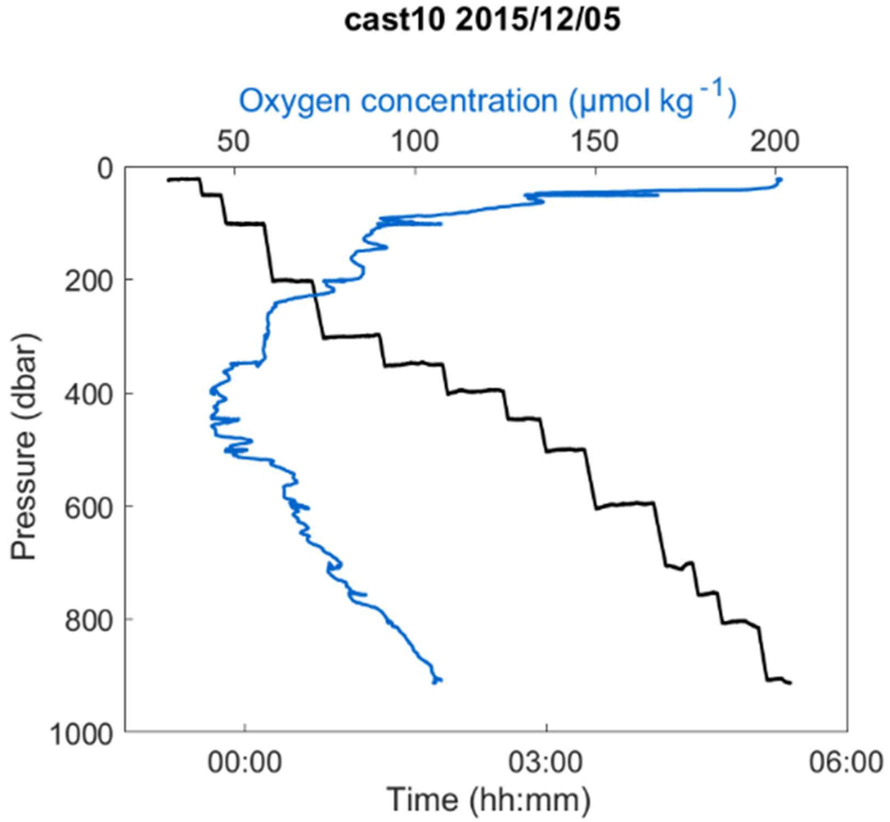

Figure 2: Stair wise trajectory of PELAGIOS through the water column, to the desired depth.

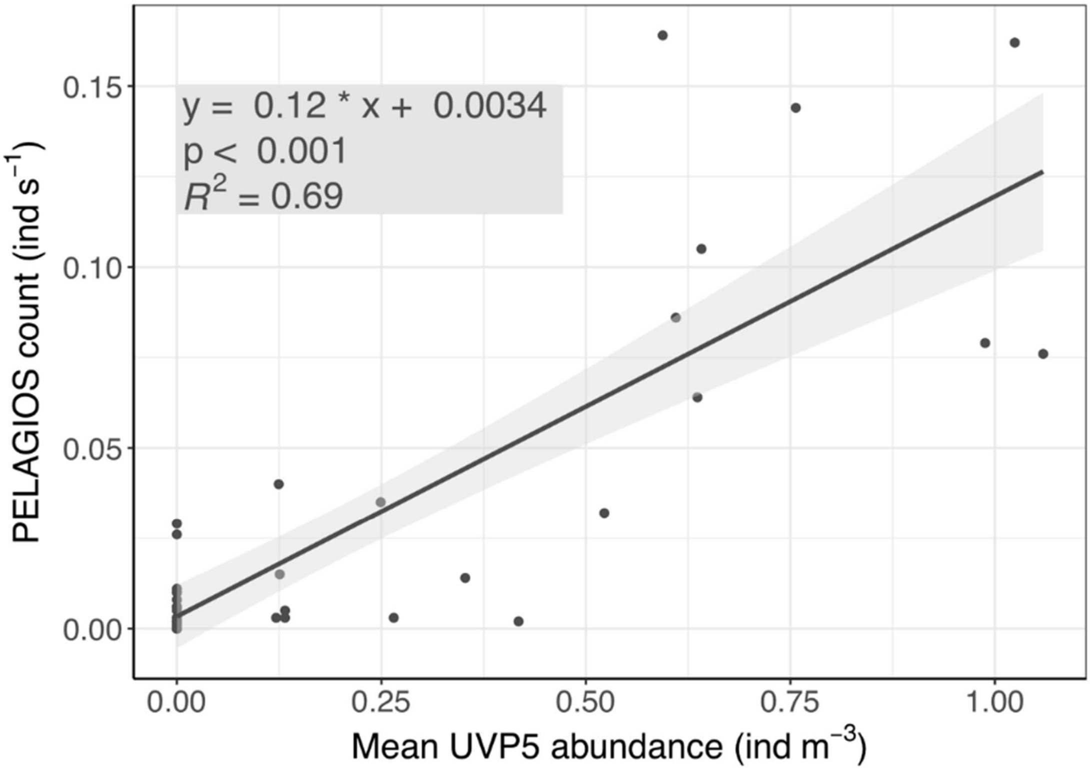

Figure 3: PELAGIOS video counts of *Poeobius* sp. as a function of UVP5-derived abundance on the same transects at two stations on cruise MSM 49 on RV MARIA S. MERIAN.

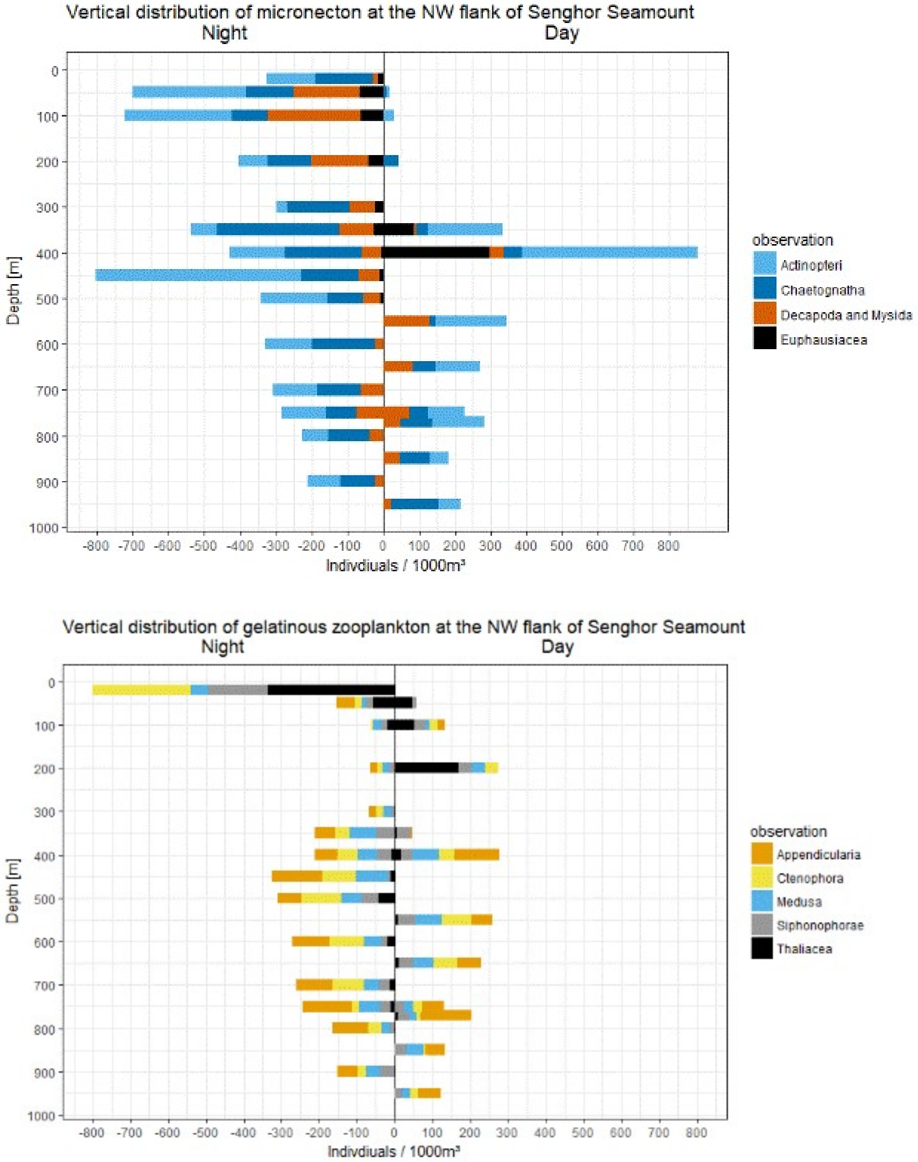

Figure 4: Day and night comparison of faunal observations obtained by PELAGIOS at the North West flank of Senghor seamount A: fishes, krill, chaetognaths and decapods B: gelatinous zooplankton groups

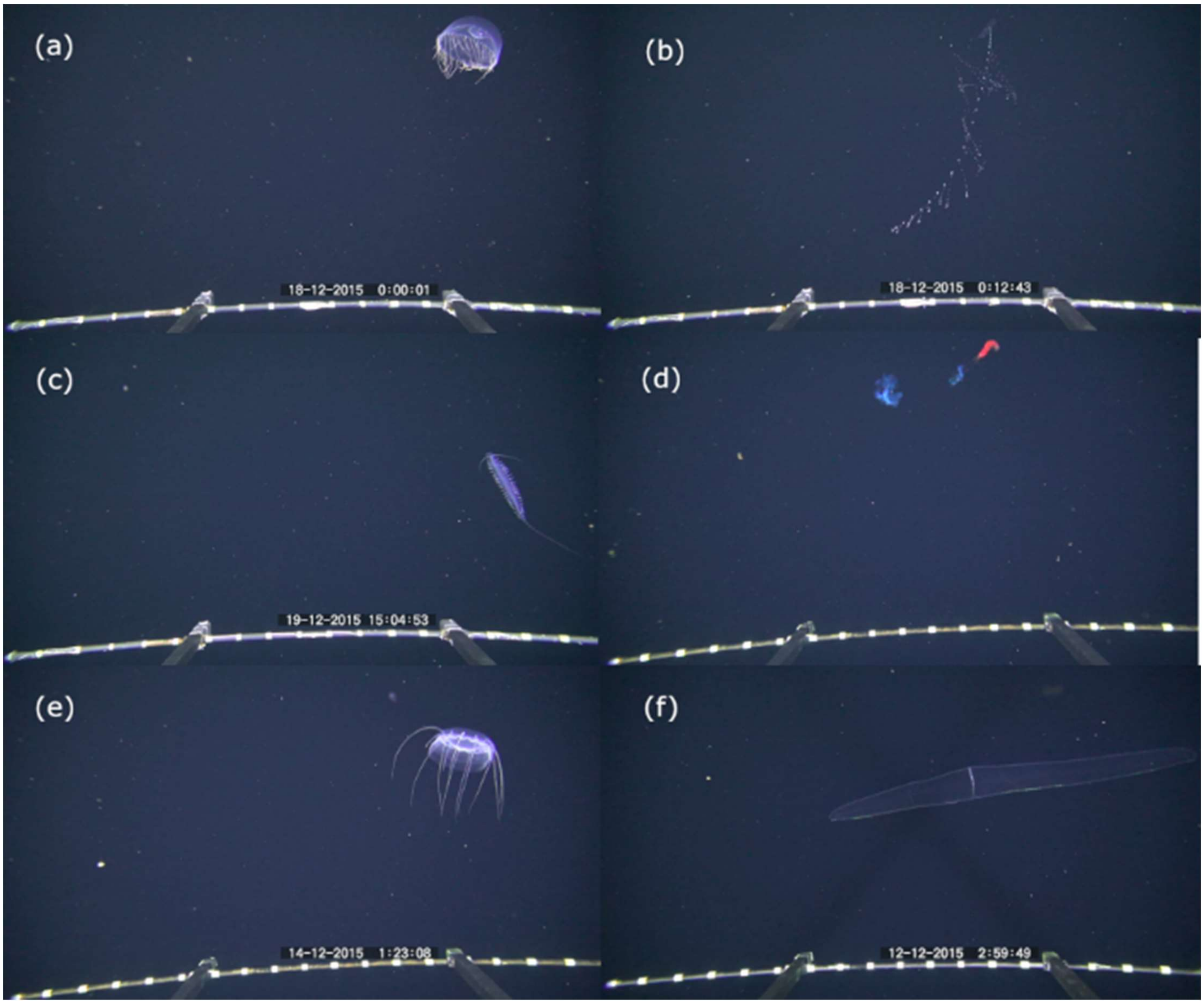

Figure 5: example of organisms encountered during pelagic video transects with PELAGIOS during cruise MSM49 in the eastern tropical Atlantic. (a) a medusa *Halitrephes* sp. (b) a calycophoran siphonophore in feeding position (c) a tomopterid worm (d) a crustacean releasing a bioluminescent cloud (e) the medusa *Solmissus* (f) the ctenophore *Cestum*

Table 1: Taxonomic groups which were encountered during pelagic video transects in the eastern tropical Atlantic.

Phylum	**Class**	**Order**	**Family**	**Genus**
Cercozoa	Thecofilosea			
Radiozoa				
Cnidaria	Hydrozoa	Narcomedusae	Solmundaeginidae	*Solmundella*
			Aeginidae	*Aegina*
				Aeginura
			Cuninidae	*Solmissus*
		Trachymedusae	Halicreatidae	*Halicreas*
				Haliscera
				Halitrephes
			Rhopalonematidae	*Colobonema*
				Crossota
				Rhopalonema
			Geryoniidae	*Geryonia*
				Liriope
		Siphonophorae	Agalmatidae	*Halistemma*
				Marrus
				Nanomia
			Apolemiidae	*Apolemia*
			Diphyidae	
			Forskaliidae	*Forskalia*
			Hippopodiidae	*Hippopodius*
				Vogtia
			Physophoridae	*Physophora*
			Prayidae	*Craseoa*
				Lilyopsis
				Praya
				Rosacea
			Pyrostephidae	*Bargmannia*
			Resomiidae	*Resomia*
	Scyphozoa	Coronatae	Atollidae	*Atolla*
			Nausithoidae	*Nausithoe*
			Peryphyllidae	*Periphylla*
Ctenophora	Nuda	Beroida	Beroidae	*Beroe*
	Tentaculata	Cestida	Cestidae	*Cestum*
				Velamen
		Cydippida	Aulacoctenidae	*Aulacoctena*
			Pleurobrachiidae	*Hormiphora*
		Lobata	Bathocyroidae	*Bathocyroe*
			Eurhamphaeidae	*Kiyohimea*
			Leucotheidae	*Leucothea*
			Ocryopsidae	*Ocyropsis*
		Thalassocalycida	Thalassocalycidae	*Thalassocalyce*

Chaeotognatha	Sagittoidea			
Annelida	Polychaeta	Phyllodocida	Tomopteridae	*Tomopteris*
		Canalipalpata	Flabelligeridae	*Poeobius*
Arthropoda	Malacostraca	Amphipoda		
		Decapoda		
		Euphausiacea		
		Isopoda	Munnopsidae	*Munnopsis*
Mollusca	Cephalopoda	Octopoda	Amphitretidae	*Bolitaena*
			Octopodidae	
		Teuthida	Cranchiidae	*Helicocranchia*
			Mastigoteuthidae	*Mastigoteuthis*
			Octopoteuthidae	*Octopoteuthis*
				Taningia
			Ommastrephidae	*Sthenoteuthis*
	Gastropoda	Nudibranchia	Phylliroidae	*Phylliroe*
		Pteropoda		
Chordata	Appendicularia	Copelata	Oikopleuridae	*Bathochordaeus*
				Mesochordaeus
	Thaliacea	Doliolida		
		Pyrosomatida	Pyrosomatidae	*Pyrostemma*
		Salpida	Salpidae	*Cyclosalpa*
	Actinopteri	Anguilliformes	Nemichthyidae	
		Myctophiformes	Myctophidae	
		Stomiiformes	Gonostomatidae	*Cyclothone*
			Sternoptychidae	