

Blue: response to the comments

Red: Changes in the revised manuscript

L223-226 I suggest to change this to:

The error of these measurements (in triplicate) was in the range of 8-25%. As the measuring scheme is possibly not optimal yet, we may repeat the measurements with purge flow rates between 350 and 450 mL per minute, especially around 400 mL/min. It might be possible to obtain even better results with fine tuning.

Corrected as suggested.

L251 minutes (not: min)

Corrected as suggested.

L266 used (not: design)

Corrected as suggested.

L279-284 I think this paragraph would fit better at the beginning of this sub-section, or even in the Introduction.

According to the editor's suggestion, we have moved this paragraph to the beginning of this sub-section. (L221-226 in the marked manuscript)

L351 I suggest to change to: During the photolysis of NO_2^- , mainly NO and OH are produced.

Corrected as suggested.

L355- 356 I do not understand what is meant with this: "thus, the authentic NO resulted from NO_2^- -photolysis was underestimated." Please modify.

The concentration of NO after exposure to sunlight is a balancing of this production against consumption by radical recombination. The study area has high concentrations of DOC and is rich in CDOM (Liu et al., 2010; Yang et al., 2011). Light might also induce NO losses by forming NO-reactive radicals from CDOM in irradiated waters during the irradiation experiments. Thus, the authentic NO resulted from NO_2^- photolysis was underestimated. In order to make it clearer, the sentence has also been rewritten as follows:

"The study area has high concentrations of DOC and is rich in CDOM (Liu et al., 2010; Yang et al., 2011). Light might also induce NO losses by forming NO-reactive radicals from CDOM in irradiated waters during the irradiation experiments. Thus, the authentic NO resulted from NO_2^- photolysis was underestimated." (L354-357 in the marked manuscript)

L462 Fehsenfeld

In Table 1: Zafiriou

Corrected as suggested.

1 **Determination of dissolved nitric oxide in coastal waters of the**
2 **Yellow Sea off Qingdao**

3 Chun-Ying Liu^{1,2,3}, Wei-Hua Feng^{1,4}, Ye Tian¹, Gui-Peng Yang^{1,2,3*}, Pei-Feng Li¹, Hermann W.
4 Bange⁵

5 ¹ College of Chemistry and Chemical Engineering, Ocean University of China, Qingdao, 266100, China

6 ² Laboratory for Marine Ecology and Environmental Science, Qingdao National Laboratory for Marine Science and
7 Technology, Qingdao, 266071, China

8 ³ Key Laboratory of Marine Chemistry Theory and Technology, Ministry of Education, Qingdao, 266100, China

9 ⁴ Key Laboratory of Engineering Oceanography, Second Institute of Oceanography, SOA, Hangzhou, 310012, China

10 ⁵ GEOMAR Helmholtz-Zentrum für Ozeanforschung Kiel, Kiel, 24105, Germany

11

12

13 * Corresponding author:

14 Prof. Gui-Peng Yang

15 College of Chemistry and Chemical Engineering

16 Ocean University of China

17 238 Songling Road, Qingdao 266100, China.

18

19 Tel.: +86 532 66782657

20 Fax: +86 532 66782540

21 *E-mail address:* gpyang@ouc.edu.cn

22

23

24 **Determination of dissolved nitric oxide in coastal waters of the** 25 **Yellow Sea off Qingdao**

26 Chun-Ying Liu^{1,2,3}, Wei-Hua Feng^{1,4}, Ye Tian¹, Gui-Peng Yang^{1,2,3*}, Pei-Feng Li¹, Hermann W.
27 Bange⁵

28 ¹ College of Chemistry and Chemical Engineering, Ocean University of China, Qingdao, 266100, China

29 ² Laboratory for Marine Ecology and Environmental Science, Qingdao National Laboratory for Marine Science and
30 Technology, Qingdao, 266071, China

31 ³ Key Laboratory of Marine Chemistry Theory and Technology, Ministry of Education, Qingdao, 266100, China

32 ⁴ Key Laboratory of Engineering Oceanography, Second Institute of Oceanography, SOA, Hangzhou, 310012, China

33 ⁵ GEOMAR Helmholtz-Zentrum für Ozeanforschung Kiel, Kiel, 24105, Germany

34 *Correspondence to:* Gui-Peng Yang (gpyang@ouc.edu.cn)

35

36 **Abstract** We developed a new method for the determination of dissolved nitric oxide (NO) in discrete seawater
37 samples based on a combination of a purge-and-trap set-up and fluorometric detection of NO.
38 2,3-diaminonaphthalene (DAN) reacts with NO in seawater to form the highly fluorescent 2,3-naphthotriazole
39 (NAT). The fluorescence intensity was linear for NO concentrations in the range from 0.14 nmol L⁻¹ to 19 nmol
40 L⁻¹. We determined a detection limit of 0.068 nmol L⁻¹, an average recovery coefficient of 83.8% (80.2-90.0%),
41 and a relative standard deviation of ±7.2%. With our method we determined for the first time the temporal and
42 spatial distributions of NO surface concentrations in coastal waters of the Yellow Sea off Qingdao and in Jiaozhou
43 Bay during a cruise in November 2009. The concentrations of NO varied from below the detection limit to 0.50
44 nmol L⁻¹ with an average of 0.26 ± 0.14 nmol L⁻¹. NO surface concentrations were generally enhanced
45 significantly during daytime implying that NO formation processes such as NO₂⁻ photolysis are much higher
46 during daytime than chemical NO consumption which, in turn, lead to a significant decrease of NO
47 concentrations during nighttime. In general, NO surface concentrations and measured NO production rates were
48 higher compared to previously reported measurements. This might be caused by the high NO₂⁻ surface
49 concentrations encountered during the cruise. Moreover, additional measurements of NO production rates
50 implied that the occurrence of particles and a temperature increase can enhance NO production rates. With the

51 method introduced here we have a reliable and comparably easy to use method at hand to measure oceanic NO
52 surface concentrations which can be used to decipher both its temporal and spatial distributions as well as its
53 biogeochemical pathways in the oceans.

54 **Keywords:** Nitric oxide (NO), determination method, coastal waters of the Yellow Sea, distribution, production
55 rate

56

57 **1 Introduction**

58 As a reactive atmospheric trace gas, nitric oxide (NO) plays important roles in tropospheric
59 chemistry: It is a key player in the formation of acid rain and ozone (Williams et al., 1992; Lee et al.,
60 1997; Mazzeo, et al., 2005). NO is an intermediate of both the terrestrial and marine nitrogen cycle
61 (Ward and Zafiriou, 1988; Williams et al., 1992; Canfield et al., 2010; Chen et al., 2010; Thamdrup,
62 2012; Voss et al., 2013). It has a variety of sources in seawater, including nitrite photolysis and various
63 microbial processes such as denitrification, anammox and dissimilatory nitrate reduction to ammonia
64 (Law, 2001; Schreiber et al., 2012; Martens-Habbena et al., 2015). Because of its chemical reactivity,
65 NO usually does not accumulate in large amounts in seawater and the ocean as a source of atmospheric
66 NO is, therefore, negligible in a global context (Zehr and Ward, 2002; Bange, 2008). Moreover, NO
67 was found to have significant effects on the growth of marine algae (Zhang et al., 2005; Liu et al., 2004;
68 2005; 2006; 2014). To this end, the determination of the spatial and temporal distributions of NO in the
69 ocean as well as deciphering its oceanic production processes and their major influencing factors are
70 essential to improve our understanding of the biogeochemical cycling NO in the ocean.

71 Because of its low concentrations in seawater caused by its fast diffusion and high chemical
72 reactivity, measurements of NO in seawater are very difficult. Therefore, there are only a few methods
73 available to determine NO (Hetrick and Schoenfish, 2009), see Tab. 1. The electrochemical method
74 using sensors in seawater medium achieved a detection limit of 42 nmol L⁻¹ (Xing et al., 2005; Zhang
75 et al., 2005). Olasehinde et al. (2009) developed a method for the determination of photochemically

76 generated NO in natural waters adopting 4,5-diaminofluorescein as a probe compound and a
77 measurement of reversed-phase high performance liquid chromatography (HPLC) with fluorescence
78 detector. The NO concentrations and signal intensities exhibited a good linearity correlation over the
79 range of 0.025-10 nmol L⁻¹ triazolofluorescein. Zafiriou and McFarland (1980) determined the NO
80 concentration of seawater by using a flow system to equilibrate the seawater samples with a gas stream
81 coupled to a chemiluminescence detector. They report an analytical precision of ±3% and an accuracy
82 of ±20%. More recently, Lutterbeck and Bange (2015) developed an improved method of a
83 chemiluminescence NO analyser connected to a stripping unit, and the limit of detection was 0.25
84 nmol L⁻¹ using a 20 mL seawater sample volume. Until now only these two chemiluminescence
85 methods were applied successfully to determine NO concentrations in seawater samples. The
86 N-nitrosation of 2,3-diaminonaphthalene (DAN) results in the highly fluorescent 2,3-naphthotriazole
87 (NAT), which could be used to detect NO concentrations as low as 10 nmol L⁻¹ (Miles et al., 1995). We
88 adopted this method for seawater medium instead of NaOH medium and the calibration curve
89 exhibited linearity over the concentration range of 1.4 - 1400 nmol L⁻¹ NO (Liu et al., 2009). However,
90 this assay cannot be used to detect trace levels of NO in seawater samples directly.

91 In this paper, we describe a modified spectrofluorometric method using a purge-and-trap technique
92 which can be used to quantify NO in seawater samples. This method was applied in a first field study on
93 the distribution and production rates of dissolved NO in coastal waters of the Yellow Sea off Qingdao
94 and Jiaozhou Bay.

95

96 2 Materials and methods

97 2.1 Instrumental set-up

98 The analytical system consists of a degassing column to purge NO from seawater samples, a
99 reaction chamber where NO reacts to form a fluorescent compound (Fig. 1), and a fluorescence
100 spectrophotometer (F-4500, Hitachi Co., Japan). The 800 mL degassing column has a sodium silicate

101 bonded sand core at the bottom to disperse the nitrogen (N₂) purge gas stream. There are four ports on
102 the column: (1) a gas port at the bottom of the degassing column where the high purity N₂ purge gas
103 (99.999%, Qingdao Heli Industry Gas Center, China) or a NO standard gas mixture (5.4 ppmv, NO/N₂)
104 (Beijing Sida Standard Substance Co., China) are introduced, (2) a drain port as outlet for water
105 samples, (3) an inlet port where water samples are pushed into the degassing column with N₂, and (4) an
106 outlet port on the top of the degassing column connected with the reaction chamber.

107 The NO standard gas cylinder is linked to the degassing column via a gas-tight syringe (Shanghai
108 Anting Injector Co., China). The N₂ gas cylinder is connected to the degassing column via a deoxygen
109 tube (Agilent Technologies, USA) to remove traces of O₂ and a glass rotameter to monitor the gas flow
110 (0.1-1 L min⁻¹, Jiangyin, China). These two gas streams enter the degassing column via the port at the
111 bottom of the flask, controlled by a three-port valve. The tubing used is made of polytetrafluorethylyene
112 (PTFE, 1/8-inch tubing outer diameter [o.d.]). Moreover, an Ultraviolet-Visible spectrophotometer
113 (UV-2550, Shimadzu Co., Japan) and an Automatic Analytical Balance (Beijing Sartorius Co., China)
114 were used in this work.

115 The degassing column, reaction chamber and the syringe were degreased with organic solvents
116 and rinsed several times with methanol and distilled water in order to minimize potential
117 contamination and adsorption effects. The degassing column was cleaned initially with detergent,
118 rinsed with water, acetone, methanol, and distilled water, and then treated for 30 min with 10% (v/v)
119 HCl in an ultrasonic bath, followed by rinsing with distilled water. Subsequently, those parts of the
120 set-up which comes into contact with the sample solutions were rinsed with methanol, water, HCl
121 solution, and dilute NaOH solution. No significant difference was found from the test of the set up
122 loaded with a water sample and without a water sample (dry run).

123

124 **2.2 Preparation of DAN and NO solutions**

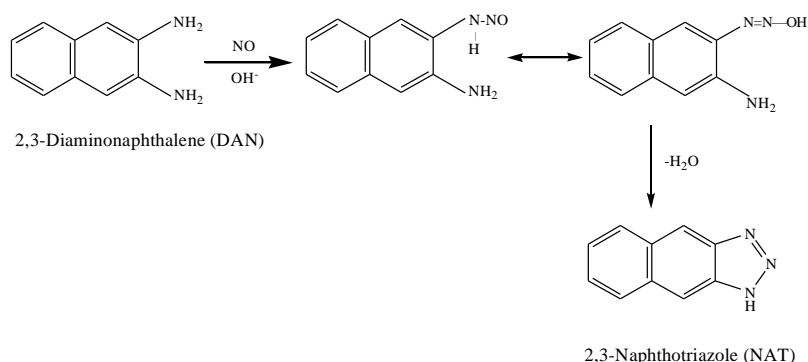
125 A 2,3-diaminonaphthalene (DAN, $\geq 95\%$, GC, Sigma-Aldrich Chemical Co., USA) stock solution
126 was prepared fresh with a concentration of 10 mmol L^{-1} in dimethylformamide (Sigma-Aldrich Chemical
127 Co., USA) and kept in the dark at $-21 \text{ }^\circ\text{C}$ until used. DAN solutions of $40 \text{ } \mu\text{mol L}^{-1}$ were prepared from
128 the stock solution in Milli-Q water, 10 mmol L^{-1} NaOH aqueous solution and filtered natural seawater,
129 respectively. Natural seawater was sampled from the coastal waters off Qingdao and was filtered through
130 a $0.45 \text{ } \mu\text{m}$ acetate cellulose membrane (Millipore, USA). The DAN solutions were purged with N_2 gas
131 for 30 min to remove oxygen (O_2), then stored on ice and transferred to a refrigerator at $4 \text{ }^\circ\text{C}$ before use.

132 An aliquot of 10 mL Milli-Q water was bubbled with N_2 gas at a flow of 10 mL min^{-1} for 1h to
133 remove O_2 after 10 min of ultrasonic degassing. The solution was then bubbled with high purity NO gas
134 (99.9% , Dalian Date Gas Ltd, China) for 30 min. The concentration of the saturated NO stock solution
135 was 1.4 mmol L^{-1} , which could be used within 3h (Lantoine et al., 1995). A series of diluted NO
136 solutions were prepared in N_2 -purged water from the NO stock solution using a syringe (Xing et al.,
137 2005).

138

139 2.3 Fluorometric detection of NO

140 DAN reacts with NO_x ($= \text{NO} + \text{NO}_2$) in alkaline medium, thus forms the highly fluorescent
141 2,3-naphthotriazole (NAT) as follows:



142

143 The reaction of NO and O_2 with 2,3 diaminonaphthalene (DAN) produced a fluorescent triazole.
144 Although the mechanism of this fluorescence has not yet been established in detail, the fluorescence was
145 reported to increase dose-dependently by NO addition (Nakatsubo et al., 1998). In seawater samples, the

146 concentration of O₂ (10⁻⁴ M order of magnitude) was far higher than that of NO (10⁻¹⁰ M order of
147 magnitude). Both of them were stripped out and reached the DAN solution finally, thus, the NO in
148 samples could almost quantitatively transform into NAT. Based on this reaction, a fluorometric method
149 was originally developed for the detection of NO in oxygenated media (Misko et al., 1993; Miles et al.,
150 1995) and has been adapted to detect NO in seawater medium instead of aqueous NaOH medium. Our
151 experiments showed that the DAN solution was stable in 12 h and the NAT solution did not change
152 within 4 h. The wavelength for NAT excitement is 383 nm and the NAT emission is monitored at a
153 wavelength of 410 nm (Liu et al., 2009).

154

155 **2.4 The influence of nitrite in seawater on the reaction of DAN and NO**

156 NO can be formed from nitrite (NO₂⁻) in seawater (Zafiriou and McFarland, 1981). Therefore, we
157 tested a potential interference of dissolved NO₂⁻ by adding different concentrations of NO₂⁻ to seawater
158 samples. The tests were conducted in the dark or with ultraviolet B (UV-B) radiation (HR 1×18 w,
159 Xinghui Electric Instrument Factory, China). The final concentrations of NO₂⁻ in the seawater samples
160 were set to 40, 80, 120, 160, and 200 μmol L⁻¹, respectively, and the reaction time was 1 h or 12 h.

161

162 **2.5 Sampling and analysis**

163 Sampling was conducted aboard the R/V ‘*Dong Fang Hong 2*’ on a cruise to the coastal waters off
164 Qingdao and Jiaozhou Bay from 4 to 6 November 2009. The locations of sampling stations are shown in
165 Fig. 2. The surface seawater samples were collected from 1 m depth at 11 stations using 8 L Niskin
166 bottles mounted on a Seabird CTD Rosette (Sea-Bird Electronics, Inc., USA). A time-course
167 observation of 24 h was carried out at station 10 near the mouth of Jiaozhou Bay.

168 A 500 mL Wheaton glass serum bottle was rinsed with the seawater three times before it was filled
169 with seawater quickly through a siphon. When the overflowed sample reached the half volume of the
170 bottle, the siphon was withdrawn rapidly and 0.5 mL saturated HgCl₂ solution was added to stop

171 biological activities and the bottle was sealed quickly. All glass bottles were covered with aluminum
172 foil to prevent NO_2^- photolysis during sampling.

173 Because NO reacts with O_2 both in the gas phase and in aqueous solution, we purged our set-up
174 for 1h with N_2 gas and sealed it before the measurements. In a first step, a certain amount of standard
175 NO gas was transferred to the reaction chamber via the degassing column by injecting it from a gas
176 tight syringe into the N_2 carrier gas stream. In the reaction chamber NO reacts with the DAN solution.
177 After the measurement of the NO gas standard, a 500 mL seawater sample was injected into the
178 degassing column and purged with N_2 gas and immediately transferred into the reaction chamber where
179 it reacts with 10 mL DAN solution. The gas flow is controlled to ensure that the reaction of NO with
180 DAN solution was completed. Finally, the fluorescence intensity of the resulting NAT solution was
181 measured with the F-4500 fluorescence spectrophotometer.

182 In order to prevent NO photochemical generation, the entire glass parts were wrapped with
183 aluminum foil. The purge-and-trap procedure was conducted at room temperature of 20 °C.

184

185 **2.6 O_2 , nutrients, DOC and chlorophyll *a* measurements**

186 Dissolved O_2 (DO) concentrations were determined according to the Winkler method. The
187 concentrations of dissolved nitrate, nitrite, and ammonia were measured by using an AutoAnalyzer 3
188 (SEAL Analytical, USA). The detection limits of the method were 0.003, 0.015, and 0.040 $\mu\text{mol L}^{-1}$ for
189 nitrate, nitrite, and ammonia, with the precision less than 1%. The intensity of sunlight was monitored
190 by the use of a TES-1322A actinometer (Taishi Co. Taiwan). Dissolved organic carbon (DOC) was
191 determined by a high-temperature combustion method using a Shimadzu TOC-5000 Analyzer with an
192 Al-Pt catalyst (Shimadzu Co., Japan). The precision of the DOC measurements was less than 2%.
193 Concentrations of chlorophyll *a* were measured with a bbe Cuvette Fluorometer (bbe-Moldaenke
194 GmbH, Kiel, Germany).

195

196 2.7 NO production rates

197 Experiments for NO production by NO_2^- photolysis were conducted at station 10 as follows:
198 Aliquots of 10 mL untreated seawater samples from 0.2 m depth or Millipore membrane (0.45 μm)
199 filtered samples were distributed into three 14 mL glass vials. The initial concentrations of NO_2^- and
200 DOC in seawater were 0.75 $\mu\text{mol L}^{-1}$ and 439 $\mu\text{mol L}^{-1}$ C, respectively. Then 200 μL of 20% NaN_3
201 solutions (instead of saturated HgCl_2 solution to avoid contamination by the photosensitive Hg) and 20
202 μL of 1 mmol L^{-1} DAN solutions were added. The vials were sealed with rubber septa and aluminum
203 crimp tops, and were exposed to sunlight on the deck at ambient temperatures (17 $^\circ\text{C}$) or at 13 ± 2 $^\circ\text{C}$
204 in a water bath supplied with the ambient seawater. For “dark” controls vials were wrapped in
205 aluminum foil. The intensity of sunlight ranged from 67565 lux to 71500 lux (average: 69430 lux).
206 After irradiation by sunlight for 30 min, the NO concentrations were measured with the method
207 described above. The NO photolysis production rates were computed as the increase of the NO
208 concentrations during the incubation time.

209 We also measured NO production rates in natural seawater at station 10. Three transparent
210 polyethylene buckets (3.5 L) were filled with the surface seawater from 0.2 m depth. The buckets were
211 exposed to sunlight in the water bath on deck. The experiment began at 8:30h (local time) and the NO
212 production rates and chlorophyll *a* concentrations were concurrently measured in 2 h intervals. An
213 aliquot of 10 mL sample was collected from each bucket using a glass syringe, distributed and sealed in
214 a 14 mL glass vial, and then incubated under the same conditions as the bucket samples. Three vials
215 per sample were used in the experiments. After 30 min of incubation, solutions of 20 μL DAN (1 mmol
216 L^{-1}) were injected into the vials, respectively. Concentrations of NO were detected and NO production
217 rates were calculated.

218

219 3 Results and Discussion

220 3.1 Method evaluation

221 NO is a conceptually important intermediate in N-cycle biogeochemistry, product of ocean
222 photochemistry, and putative inter-cellular signal. Unfortunately, our knowledge about the oceanic NO
223 distribution and the major pathways of NO is very poor. There are only a few published NO
224 concentration measurements available because a reliable and easy to use method to determine dissolved
225 NO at in-situ concentrations in seawater samples is missing. We try to find a solid method both
226 convenient for many labs and sensitive enough, which seems to have promise.

227 Both the purge time and flow of the purge gas (N₂) significantly influence the yield of the NO +
228 DAN reaction and thus, the overall purge efficiency (see Tab. 2). The optimal (i.e. maximum) reaction
229 yield was 85% after 30 min of purging at a flow of 400 mL min⁻¹. The error of these measurements (in
230 triplicate) was in the range of 8-25%. As the measuring scheme is possibly not optimal yet, we may
231 repeat the measurements with purge flow rates between 350 and 450 mL per minute, especially around
232 400 mL/min. It might be possible to obtain even better results with fine tuning.

233 The set-up was tested for internal NO production or loss by comparing the fluorescence intensity
234 from NO-free gas or NO calibration gas passing through the degassing column with the fluorescence
235 intensity from the same gas bypassing the degassing column. This procedure was repeated with both a
236 dry degassing column and a moistening degassing column (by a minimum amount of filtered seawater).
237 Neither NO production nor NO loss by adsorption was observed in the set-up in all test runs.

238 Seawater samples from coastal waters off Qingdao were analyzed in the lab up to of 7 times and
239 gave a relative standard deviation of ±7.2%. The detection limit of our method was determined to be
240 0.068 nmol L⁻¹ (S/N = 3), which is lower than most of the reported detection limits for NO
241 measurements in seawater (see Tab. 1)

242 The NO recovery coefficient of our purge-and-trap system was estimated by the addition of the
243 same volume of a NO standard solution to (i) 500 mL NO-free seawater in the degassing column and (ii)
244 to 10 mL DAN solution (with a DAN concentration of 40 μmol L⁻¹) in the reaction chamber. The
245 recovery coefficient (*RC*) of NO was calculated according to:

246
$$RC(\%) = NO(\text{sw}) / NO(\text{DAN}) \times 100\%.$$

247 Where $NO(\text{DAN})$ stands for the NO directly injected to the DAN solution and $NO(\text{sw})$ stands for the
248 NO measured from the sample in degassing column according to the method described above. The rate
249 law obtained from the oxidation of NO is

250
$$-d[\text{NO}]/dt = 4k[\text{NO}]^2[\text{O}_2]$$

251 with $k = 2 \times 10^6 \text{ M}^{-2} \text{ s}^{-1}$. The reaction of NO with O_2 could consume NO in the stripping period. The NO
252 recovery coefficients of the purge-and-trap system were evaluated and ranged from 80.2% to 90.0%,
253 with an average of 83.8%. Furthermore, three replicates of in-situ seawater were measured using our
254 system and method, the aqueous NO solution did not change within one hour, which was also
255 demonstrated by Lutterbeck and Bange (2015).

256 In order to check the linearity of our method, a solution of 10 mL $40 \mu\text{mol L}^{-1}$ DAN was injected
257 into the reaction chamber and purged with N_2 gas at a rate of 10 mL min^{-1} for 5 minutes prior to the
258 actual measurements. A series of NO-free seawater samples placed in the degassing column were
259 spiked with different volumes of the NO standard gas (mixing ratio 5.4 ppmv NO/ N_2) and analyzed
260 according to the procedure described above. The resulting fluorescence intensity was linear with the NO
261 concentrations in the range from 0.14 to 19.0 nmol L^{-1} ($y = 7.4286x + 0.6188$, $R = 0.9976$, $P < 0.0001$)
262 (Fig. 3).

263 The results of the samples spiked with varying concentrations of dissolved NO_2^- are given in Fig.
264 4. The blank had no signal when measured with the fluorescence spectrophotometer after UV-B radiation
265 or in the dark. That is, the NO concentrations were zero. Nitrite did not cause significant problems with
266 natural samples during the measurement process. In general, samples with the same NO_2^- concentration
267 showed higher fluorescence when UV-irradiated or kept in dark for 12 h compared to samples under
268 short term (i.e. 1 h) UV irradiation or kept in dark. This points a significant NO production under UV
269 irradiation ($n=5$, $F=76.13$, $p=2.32 \times 10^{-5}$) and (albeit weaker) NO dark production from NO_2^- . Higher NO_2^-
270 concentrations resulted in a slight increase of fluorescence when irradiated. Therefore we conclude that

271 the measurements of NO should be done in the dark as soon as possible after sampling when high NO_2^-
272 concentrations occur. Here these high NO_2^- concentrations were used to demonstrate no obvious effect
273 caused by NO_2^- on the detection method, thus, low concentrations of NO_2^- also do not affect this method.
274 On the other hand, the fluorescence intensity could not be detected with low concentrations of NO_2^- .

275 To assess the influence of the interferences of dissolved organic matter, trace metals, nutrients, and
276 other substances in seawater, the NO/fluorescence intensity relationship should be determined when
277 the method is applied in different oceanic regions.

278 With our method we are able to detect $> 0.068 \text{ nmol L}^{-1}$ NO in discrete seawater samples with a
279 volume of 500 mL. With a larger degassing column, even lower concentrations of NO might be
280 determined.

281 A U-shaped tube and cold bath (i.e. a water trap) was initially placed between the degassing
282 column and the reaction chamber in order to eliminate small amounts of water carried by the N_2 gas
283 stream. However, we found that the fluorescence intensities did not show significant differences when
284 the water trap was removed.

285

286 3.2 Distribution of dissolved NO in coastal waters of Qingdao

287 Fig. 5 shows the NO concentrations of surface seawater in coastal waters off Qingdao (stations
288 S01-S09) and in the Jiaozhou Bay (stations S10 and S11). The concentrations of NO ranged from
289 below the detection limit (stat. 02 and 03) up to $0.50 \pm 0.01 \text{ nmol L}^{-1}$ (stat. S08), with an overall mean
290 of $0.26 \pm 0.14 \text{ nmol L}^{-1}$. It is noteworthy that the higher NO concentrations seem to be related to the
291 time point of sampling (given in local time): Samples of stations 2 and 3 were collected at night time,
292 22:30h and 00:50h, respectively, while samples for stations 5, 6, 7 and 8 were collected during the day
293 time (08:58h - 15:38h). (Stations S09 and S10 have been measured in Jiaozhou Bay and, thus, their
294 NO concentrations are directly not comparable with the stations off Qingdao). Our results are
295 generally consistent with the findings in the aquatic ecosystem of Daya Bay in China (Zhang et al.,

296 2006) and the nitrite-rich surface waters of the central equatorial Pacific Ocean (Zafiriou et al., 1980),
297 indicating that sunlight could be a main factor affecting NO formation in seawater. The concentrations
298 of NO in coastal surface waters off Qingdao were found to be an order of magnitude higher than those
299 in surface waters during day time in the central equatorial Pacific Ocean (0.05 nmol L^{-1}) (Zafiriou et
300 al., 1980; Zafiriou and McFarland, 1981). This difference is probably related to the concentrations of
301 NO_2^- in seawater. Zafiriou et al. (1980) proposed that sunlight photolyzes NO_2^- in surface water by the
302 following reaction:



304 According to the reaction above, high concentrations of NO_2^- together with strong solar irradiation
305 could cause enhanced concentrations of NO in seawater. The sunlight intensity of the central
306 equatorial Pacific is generally higher than that of coastal waters of Qingdao (located at $36^\circ 05' \text{N}$);
307 however, the coastal waters off Qingdao at the time of our measurements exhibited an average NO_2^-
308 concentration of $0.49 \pm 0.25 \text{ } \mu\text{mol L}^{-1}$, which was much higher than that observed concentration in the
309 central equatorial Pacific Ocean ($\sim 0.1 \text{ } \mu\text{mol L}^{-1}$).

310 NO is a short-lived intermediate of various microbial processes of the nitrogen cycle, which is
311 involved in denitrification (Kampschreur et al., 2007), anammox (Kartal et al., 2011) and archaea
312 ammonia-oxidizing (Martens-Habbena et al., 2015) processes. Zafiriou and McFarland (1981) analyzed
313 NO in seawater samples at the sea surface of the central equatorial Pacific by stripping NO into an air
314 and N_2 stream by passing it through the chemiluminescence - type detector. Thus, the NO
315 concentrations were underestimated to some extent because seawater samples were suboxic or anoxic.
316 However, time-dependent losses from microbial processes were minimized. Lutterbeck and Bange
317 (2015) improved the method above to determine dissolved NO in discrete seawater samples of the
318 eastern tropical South Pacific Ocean. The contamination by O_2 diffusion into the continue samples
319 could be further minimized. This work was also designed to detect dissolved NO in discrete seawater

320 samples with a combination of a purge-and-trap set-up and fluorometric NO analyzer. The HgCl₂
321 solution was added to stop biological activities during the stripping. However, the disposal of these
322 Hg-contaminated solutions is a tough proposition. To improve the method the purge-and-trap set-up
323 could be modified and the stripping time could be reduced, then the addition of HgCl₂ solution may be
324 removed in the future.

325 The diurnal variation of NO concentrations and other parameters in surface seawater are shown in
326 Fig. 6. Concentrations of NO presented a significant diurnal variation within 24 h. The peak value
327 appeared at 15:00h (local time) with a concentration of 0.81 nmol L⁻¹. After that the concentration of
328 NO decreased with time gradually until a minimum value occurred at 03:00. Obviously, the
329 concentration of dissolved NO at this station was influenced by the in-situ sunlight intensity. However,
330 the maximum NO concentration appeared not at 12:00h but at 15:00h, which suggesting that there
331 were other influencing factors besides sunlight irradiation.

332

333 3.3 NO production rates in coastal waters

334 The results of the NO irradiation experiments are given in Fig. 7. The production rate of NO
335 through seawater irradiation was $1.52 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$ which is slightly higher than that NO production
336 rate of the 0.45 μm Millipore filtered samples ($1.46 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$). The difference may indicate that
337 particles in seawater could increase the NO production rate. The non-filtered samples incubated in the
338 water bath had a lower NO production rate ($1.44 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$) compared to the other non-filtered
339 treatment, which could be ascribed to the difference of the temperature. The ambient temperature and
340 water bath were 17 °C and 13 °C, respectively, thus the higher temperature may resulted in a higher
341 photolysis rate. The photochemical production rates of NO in Qingdao coastal waters during the
342 daytime were generally higher than that reported from the central equatorial Pacific Ocean
343 ($0.4\text{-}1.2 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$) (Zafiriou and McFarland, 1981).

344 Previous experiments about NO₂⁻ photolysis were also carried out in our laboratory (Li et al.,

2011): The production of NO was observed after 3 h illumination of 10-100 $\mu\text{mol L}^{-1}$ NO_2^- in Milli-Q water. There was an increasing trend of NO concentrations with the NO_2^- concentrations. For natural seawater, it was observed to have an increasing trend of NO concentration with the illumination time (Li et al., 2011). The process of sunlight photolysis of NO_2^- in surface water was demonstrated, which was consistent with the results of Zafiriou et al. (1980) and Olasehinde et al. (2009).

The production and consumption of NO occur synchronously when sunlight photolyze natural seawater. During the photolysis of NO_2^- , mainly NO and OH are produced. On the other hand, the loss of NO happens by forming NO-reactive radicals from CDOM (Zafiriou et al., 1990; Zafiriou and Dister, 1991; Olasehinde et al., 2009). The concentration of NO after exposure to sunlight is a balancing of this production against consumption by radical recombination. The study area has high concentrations of DOC and is rich in CDOM (Liu et al., 2010; Yang et al., 2011). Light might also induce NO losses by forming NO-reactive radicals from CDOM in irradiated waters during the irradiation experiments. Thus, the authentic NO resulted from NO_2^- photolysis was underestimated. The photochemical production rates of NO were only a total value of production and consumption in this study.

The on-deck incubation experiments for the production rates of NO in Qingdao coastal waters, together with chlorophyll *a* concentrations and sunlight intensities, are shown in Fig. 8. The production rates of NO exhibited a clear variation during the course of the day with a maximum value appearing at 14:30h (local time). The maximum value of $2.52 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$ was about seven-fold higher than the minimum value at 08:30h. The production rates of NO kept an increasing trend from 08:30h to 14:30h. The mean production rate in Qingdao coastal waters was $1.51 \times 10^{-12} \text{ mol L}^{-1} \text{ s}^{-1}$ during the day. The variation of the production rates of NO did not follow the trends in chlorophyll *a* concentrations and solar radiation. Therefore, the production pattern of NO in marine environments deserves further research.

4 Summary

370 For the determination of NO concentrations in discrete seawater samples we developed a new
371 method by combining a purge-and-trap set-up with fluorometric detection of NO. The method showed
372 a linear fluorescence intensity for NO concentrations ranging from 0.14 nmol L⁻¹ to 19 nmol L⁻¹. The
373 detection limit is 0.068 nmol L⁻¹ (S/N =3), the average recovery coefficient is 83.8% (80.2~90.0%),
374 and the relative standard deviation is ±7.2%. Our method was applied to measure concentrations of
375 NO in surface layer of the coastal waters off Qingdao and Jiaozhou Bay. NO concentrations varied
376 from below the detection limit to 0.50 nmol L⁻¹, with an average of 0.26 ± 0.14 nmol L⁻¹. The
377 concentrations of NO in coastal waters off Qingdao were an order of magnitude higher than those in
378 surface waters of the central equatorial Pacific. NO surface concentrations were generally enhanced
379 significantly during daytime implying that NO formation processes such as NO₂⁻ photolysis are much
380 higher during daytime than chemical NO consumption which, in turn leads to the observed significant
381 decrease of the NO concentrations during nighttime. The measurements of NO production rates
382 showed that the occurrence of particles and an increase in temperature can enhance NO production.

383 We conclude that our method can be applied to measure (i) NO concentrations in the ocean
384 surface, (ii) NO production and consumption pathways in oceanic waters and (ii) NO production rates
385 in culture experiments.

386

387 **Acknowledgments**

388 We thank Li Tie and Zhu Chenjian for their assistance during sample collection. This research was
389 supported by the National Natural Science Foundation of China (No. 41676065), the National Key
390 Research and Development Program of China (Grant No. 2016YFA0601301), the Fundamental Research
391 Funds for the Central Universities (No. 201564015), and Aoshan Talents Program Supported by Qingdao
392 National Laboratory for Marine Science and Technology (No. 2015ASTP).

393

394 **References**

- 395 Bange, H. W.: Chapter 2 - Gaseous Nitrogen Compounds (NO, N₂O, N₂, NH₃) in the Ocean in: Nitrogen in the Marine
396 Environment (Second Edition), Elsevier, Amsterdam, Netherlands, 51-94, 2008.
- 397 Canfield, D. E., Glazer, A. N., and Falkowski, P. G.: The evolution and future of the Earth's nitrogen cycle, Science,
398 330,192-196, 2010.
- 399 Chen, J., Wu, F. H., Xiao, Q., Yang, Z. H., Huang, S. K., Wang, J., Wu, Y. G., Dong, X. J., Pei, Z. M., Zhen, H. L.:
400 Diurnal variation of nitric oxide emission flux from a mangrove wetland in Zhangjiang River Estuary, China,
401 Estuar. Coast. Shelf Sci., 90, 212-220, 2010.
- 402 Hetrick, E. M., and Schoenfish, M. H.: Analytical chemistry of nitric oxide, Annu. Rev. Anal. Chem., 2, 409-433,
403 2009.
- 404 Kampschreur, M. J., Picioreanu, C., Tan, N., Kleerebezem, R., Jetten, M. S. M., and van Loosdrecht, M. C. M.:
405 Unraveling the source of nitric oxide emission during nitrification, Water Environ. Res., 79, 2499–2509,
406 2007.
- 407 Kartal, B., Maalcke, W. J., de Almeida, N. M., Cirpus, I., Gloerich, J., Geert, S.W., den Camp, H., Harhangi, H. R.,
408 Janssen-Megens, E. M., Francoijs, K. J., Stunnenberg, H. G., Keltjens, J. T., Jetten, M. S. M., and Strous, M.:
409 Molecular mechanism of anaerobic ammonium oxidation, Nature, 479, 127–130, 2011.
- 410 Lantoiné, F., Trevin, S., Bedioui, F., and Devynck, J.: Selective and sensitive electrochemical measurement of nitric
411 oxide in aqueous solution: discussion and new results. J Electroanal. Chem., 392, 85-89, 1995.
- 412 Law, C. S.: Air-Sea Transfer: N₂O, NO, CH₄, CO. J.H. Steele, S.A. Thorpe, K.K. Turekian. (eds.) Encyclopedia of
413 Ocean Sciences. San Diego, USA, Academic Press, 137-143, 2001.
- 414 Lee, D. S., Köhler, I., Grobler, E., Rohrer, F., Sausen, R., Gallardo-Klenner, L., Olivier, J. G. J., Dentener, F. J., and
415 Bouwman, A. F.: Estimations of global no_x emissions and their uncertainties, Atmos. Environ., 31(12),
416 1735-1749. 1997.
- 417 Li, P. F., Li, W. S., Liu, C. Y., Zhu, X. C., and Zhang, Q.: The Photodecomposition of Nitrite in Water, Environ. Chem.,
418 30, 1883-1888, 2011. (in Chinese with English abstract).
- 419 Liu, C. Y., Kieber, D. J., Yang, G. P., Xue, C., Wang, L. L., and Liu, H. H.: Evidence for the mutual effects of
420 dimethylsulfoniopropionate and nitric oxide during the growth of marine microalgae, Nitric Oxide, 42, 54-61,
421 2014.
- 422 Liu, C. Y., Yang, X. M., Yang, G. P., Zhou, L. M., and Li, P. F.: Composition and characterization of colloidal organic

423 matter in the coastal surface waters of Qingdao, China, *Mar. Chem.*, 121:123-131, 2010.

424 Liu, C. Y., Zhang, Z. B., and Chen, X. R.: The mutual effects of nitric oxide and iron on the growth of marine algae,
425 *Acta Oceanol. Sin.*, 24(5), 100-109, 2005.

426 Liu, C. Y., Zhang, Z. B., Li, P. F., and Huang, H. W.: Growth effect of exogenous nitric oxide on *Platymonas*
427 *subcordiformis* and spectrum study, *Chinese J Environ. Sci.*, 27(6), 1062-1067, 2006. (in Chinese with English
428 abstract).

429 Liu, C. Y., Zhang, Z. B., Xing, L., Lin, C., and Wu, Z. Z.: The ocean biogeochemistry of nitric oxide, *Periodical of*
430 *Ocean University of China*, 34(sup), 16-22, 2004. (in Chinese with English abstract).

431 Liu, C. Y., Zhao, M., Ren, C.Y., Yang, G. P., Li, P. F., and Han, Y.: Direct measurement of nitric oxide in seawater
432 medium by fluorometric method. *Chinese J Anal. Chem.*, 37(10), 1463-1467, 2009.

433 Lutterbeck, H. E., and Bange, H. W.: An improved method for the determination of dissolved nitric oxide (NO) in
434 seawater samples, *Ocean Science*, 12, 959-981, 2015.

435 Martens-Habbena, W., Qin, W., Horak, R. E., Urakawa, H., Schauer, A. J., Moffett, J. W., Armbrust, E. V., Ingalls, A.
436 E., Devol, A. H., Stahl, D. A.: The production of nitric oxide by marine ammonia-oxidizing archaea and inhibition
437 of archaeal ammonia oxidation by a nitric oxide scavenger, *Environ. Microbiol.* 17(7), 2261-74, 2015. doi:
438 10.1111/1462-2920.12677.

439 Mazzeo, N. A., Venegas, L. E., and Choren, H.: Analysis of NO, NO₂, O₃ and NO_x concentrations measured at a
440 green area of Buenos Aires City during wintertime. *Atmos. Environ.*, 39(17), 3055-3068, 2005.

441 Miles, A. M., Chen, Y., Owens, M. W., and Grisham, M. B.: Fluorometric determination of nitric oxide. In *Methods:*
442 *A Companion to Methods in Enzymology*, 7, 40-47, 1995.

443 Misko, T. P., Schilling, R. J., Salvemini, D., Moore, W. M., and Currie, M. G.: A fluorometric assay for the
444 measurement of nitrite in biological samples. *Anal. Biochem.*, 214(1), 11-16, 1993.

445 Nakatsubo, N., Kojima, H., Sakurai, K., Kikuchi, K., Nagoshi, H., Hirata, Y., Akaike, T., Maeda, H., Urano,
446 Y., Higuchi, T., and Nagano, T.: Improved nitric oxide detection using 2,3-diaminonaphthalene and its
447 application to the evaluation of novel nitric oxide synthase inhibitors. *Biol. Pharm. Bull.* 21(12):1247-1250,
448 1998.

449 Olasehinde, E. F., Takeda, K., and Sakugawa, H.: Development of an analytical method for nitric oxide radical
450 determination in natural waters, *Anal. Chem.*, 81(16), 6843-6850, 2009.

- 451 Schreiber, F., Polerecky, L., and de Beer, D.: Nitric oxide microsensor for high spatial resolution measurements in
452 biofilms and sediments, *Anal. Chem.*, 80, 1152–1158, 2008. doi: 10.1021/ac071563x.
- 453 Schreiber, F., Wunderlich, P., Udert, K.M. and Wells, G.F.: Nitric oxide and nitrous oxide turnover in natural and
454 engineered microbial communities: biological pathways, chemical reactions, and novel technologies, *Front.*
455 *Microbiol.*, 2012. doi: 10.3389/fmicb.2012.0372.
- 456 Thamdrup, B.: New pathways and processes in the global nitrogen cycle, *Annual Review of Ecology Evolution and*
457 *Systematics*, 43, 407-428, 2012.
- 458 Voss, M., Bange, H. W., Joachim, W. D., Middelburg, J. J., Montoya, J. P., Ward, B.: The marine nitrogen cycle:
459 recent discoveries, uncertainties and the potential relevance of climate change, *Phil. Trans. R. Soc. B*, 368 (1621),
460 2013. doi: 10.1098/rstb.2013.0121.
- 461 Ward, B. B., and Zafiriou, O. C.: Nitrification and nitric oxide in the oxygen minimum of the eastern tropical North
462 Pacific, *Deep-Sea Res.*, 35(7), 1127-1142, 1988.
- 463 Williams, E. J., Hutchinson, G. L., and [Fehsenfeld](#), F. C.: NO_x and N₂O emissions from soil, *Global Biogeochem. Cy.*,
464 6, 351-388, 1992.
- 465 Xing, L., Zhang, Z. B., Liu, C. Y., Wu, Z. Z., and Lin, C.: Amperometric detection of nitric oxide with microsensor in
466 the medium of seawater and its applications, *Sensors*, 5(12), 537-545, 2005.
- 467 Yang, G. P., Ren, C. Y., Lu, X. L., Liu, C. Y., and Ding, H. B.: Distribution, flux and photoproduction of carbon
468 monoxide in the East China Sea and the Yellow Sea in spring, *J. Geophys. Res.* 116, C02001, 2011,
469 doi:10.1029/2010JC006300
- 470 Zafiriou, O. C., and McFarland, M.: Determination of trace levels of nitric oxide in aqueous solution, *Anal. Chem.*,
471 52(11), 1662-1667, 1980.
- 472 Zafiriou, O. C. and Dister, B.: Photochemical free radical production rates: Gulf of Maine and Woods Hole-Miami
473 transect, *J. Geophys. Res.* 96(C3), 4939–4945, 1991.
- 474 Zafiriou, O. C., and McFarland, M.: Nitric oxide from nitrite photolysis in the central equatorial Pacific, *J Geophys.*
475 *Res.*, 86(C4), 3173-3182, 1981.
- 476 Zafiriou, O. C., Blough, N. V., Dister, B., Kieber, D., and Moffett, J.: Molecular probe systems for reactive transients
477 in natural waters, *Mar. Chem.*, 30, 45–71, 1990.
- 478 Zafiriou, O. C., McFarland, M., and Bromund, R. H.: Nitric oxide in seawater, *Science*, 207, 637-639, 1980.

- 479** Zehr, J. P., and Ward, B. B.: Nitrogen cycling in the ocean: New perspectives on processes and paradigms, *Appl.*
480 *Environ. Microb.*, 68, 1015-1024, 2002.
- 481** Zhang, Z. B., Lin, C., Liu, C. Y., Xing, L., Wu, Z. Z., and Sun, F.: Study on patterns and chemical features of NO effect
482 on marine phytoplankton growth, *Sci. China B: Chem.*, 48(3), 376-384, 2005.
- 483** Zhang, Z. B., Liu, C. Y., Wu, Z. Z., Xing, L., and Li, P. F.: Detection of nitric oxide in culture media and studies on
484 nitric oxide formation by marine microalgae, *Med. Sci. Monit.*, 12, 75-85, 2006.
- 485** Zhang, Z. B., Xing, L., Wu, Z. Z., Liu, C. Y., Lin, C., and Liu, L.S. Discovery of nitric oxide in marine ecological
486 system and the chemical characteristics of nitric oxide, *Sci. China B: Chem.*, 49(5), 475-480, 2006.
- 487**

Figure Captions

488

489

490 Fig. 1 The purge-and-trap system for the determination of dissolved nitric oxide in seawater

491 **Fig. 2** Locations of the sampling stations in the coastal waters off Qingdao and Jiaozhou Bay

492 **Fig. 3** Relationship between nitric oxide concentrations and fluorescence intensities

493 **Fig. 4** The fluorescence variations of NAT in seawater with different concentrations of nitrite in the

494 dark or under UV-B radiation

495 **Fig. 5** The concentrations of NO in the surface water off Qingdao and Jiaozhou Bay

496 **Fig. 6** The diurnal variations of NO concentrations and related parameters in the surface seawater at

497 station 10

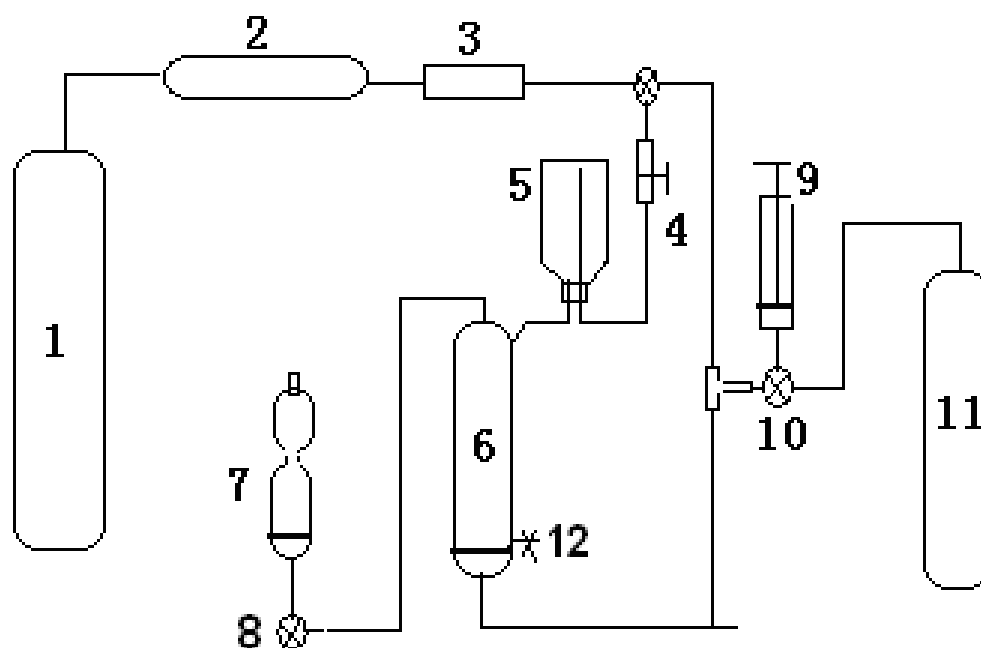
498 **Fig. 7** The production rates of NO by seawater irradiation under natural light after different treatments

499 **Fig. 8** The variations of NO production rates, chlorophyll *a* concentrations and sunlight intensities in the

500 incubation experiments with Qingdao coastal waters

501

502

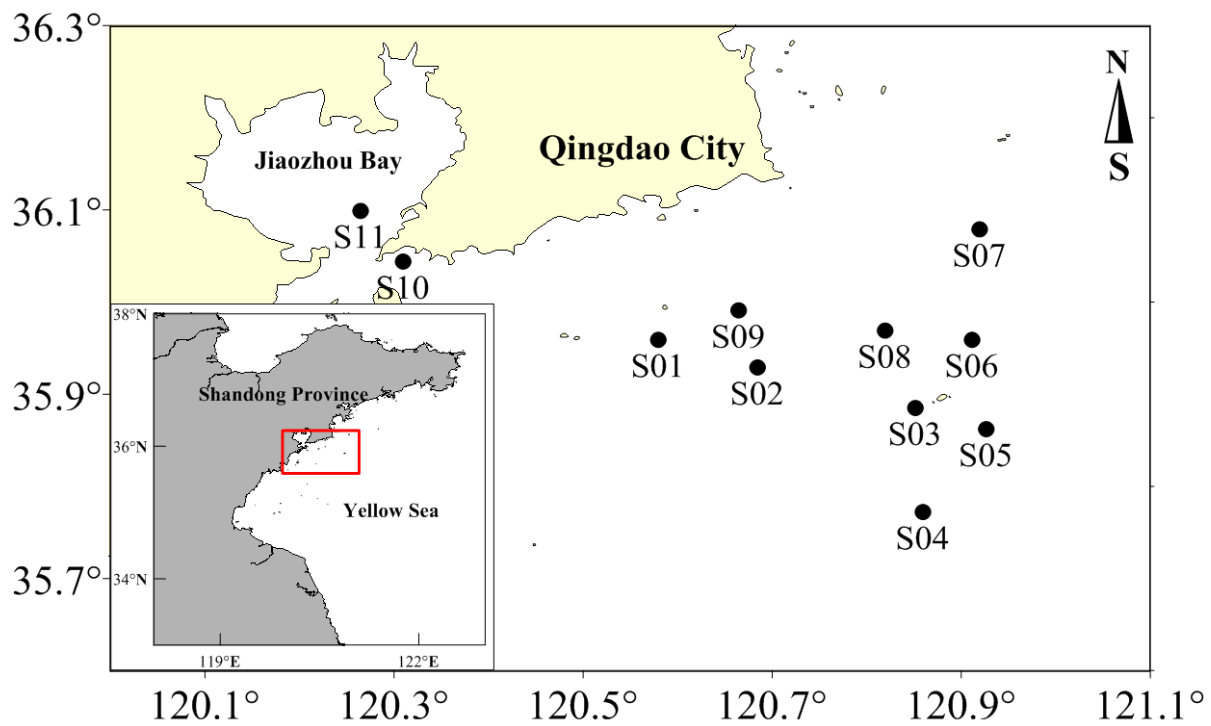


503

504 Fig. 1 The purge-and-trap system for the determination of dissolved nitric oxide in seawater

505 (1. N₂ gas; 2. Deoxygenation tube; 3. Glass rotameter; 4. 2-port valve; 5. Sample vial; 6. Degassing
506 column; 7. Reaction chamber; 8 and 10. 3-port valves; 9. Gas-tight syringe; 11. NO standard gas; 12.
507 Drain)

508

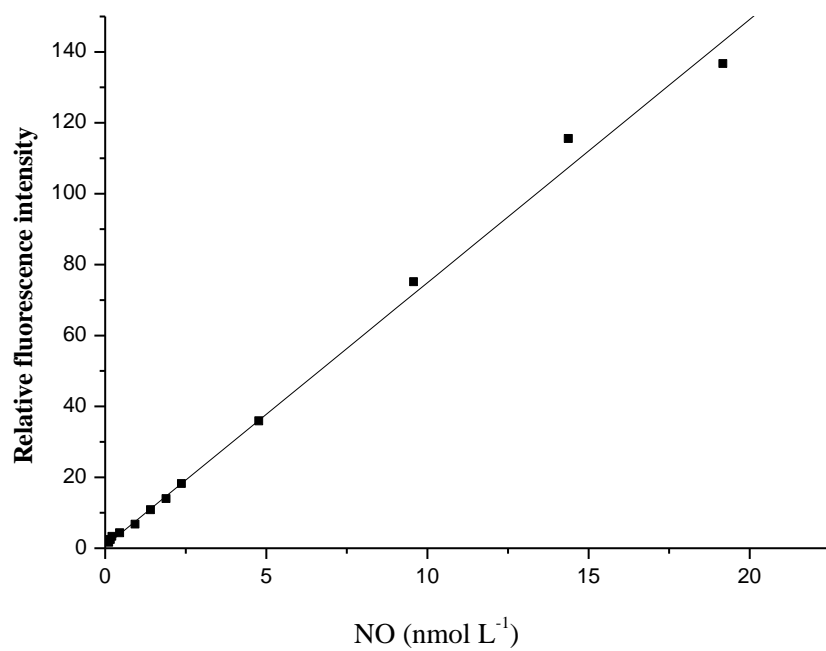


509

510

511

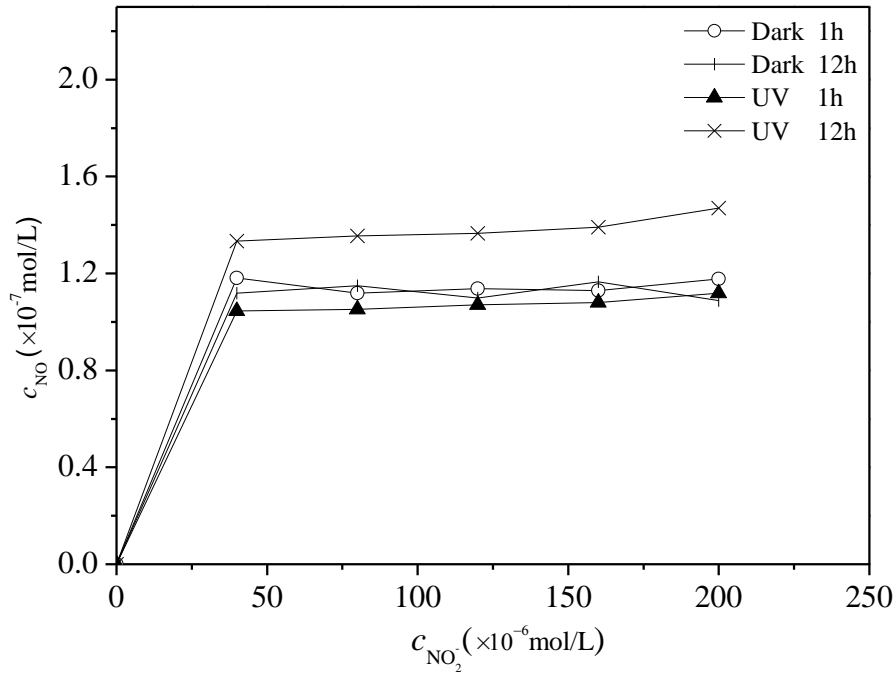
Fig. 2 Location of the sampling stations in the coastal waters off Qingdao and Jiaozhou Bay



512

513 **Fig. 3** Relationship between nitric oxide concentrations and fluorescence intensities

514

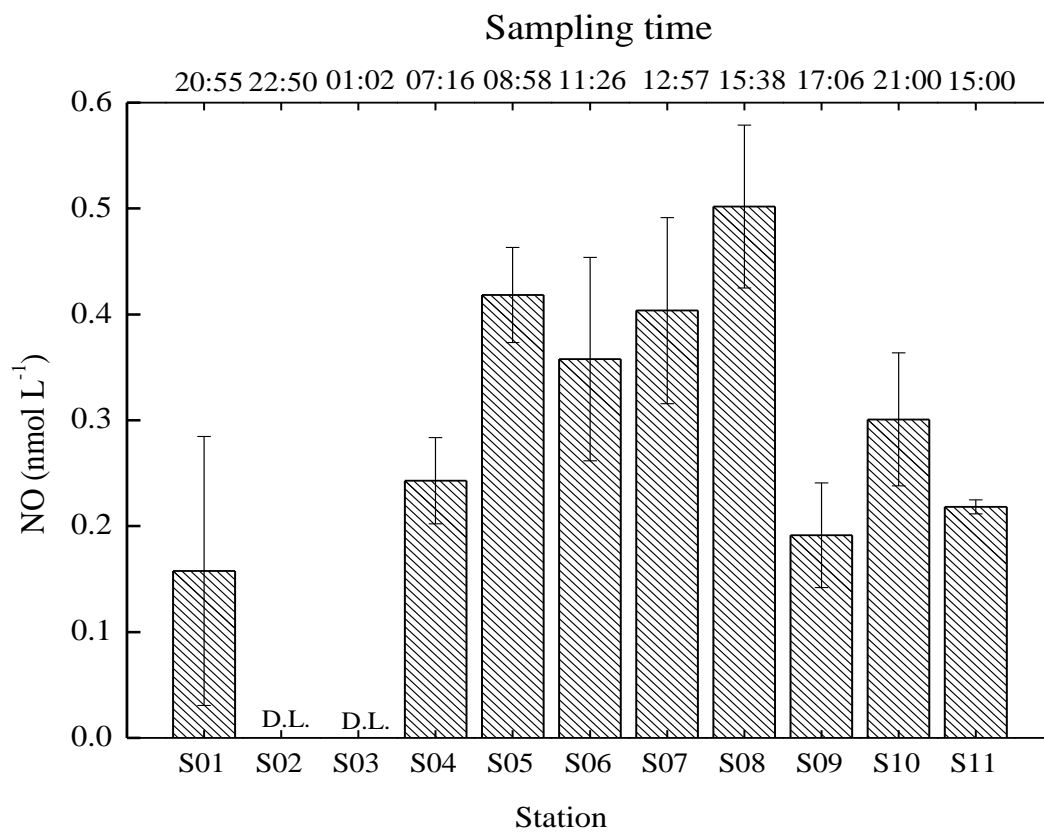


515

516 **Fig. 4** The variations of NO concentrations in seawater added with different concentrations of nitrite in

517 the dark or under UV-B radiation

518



519

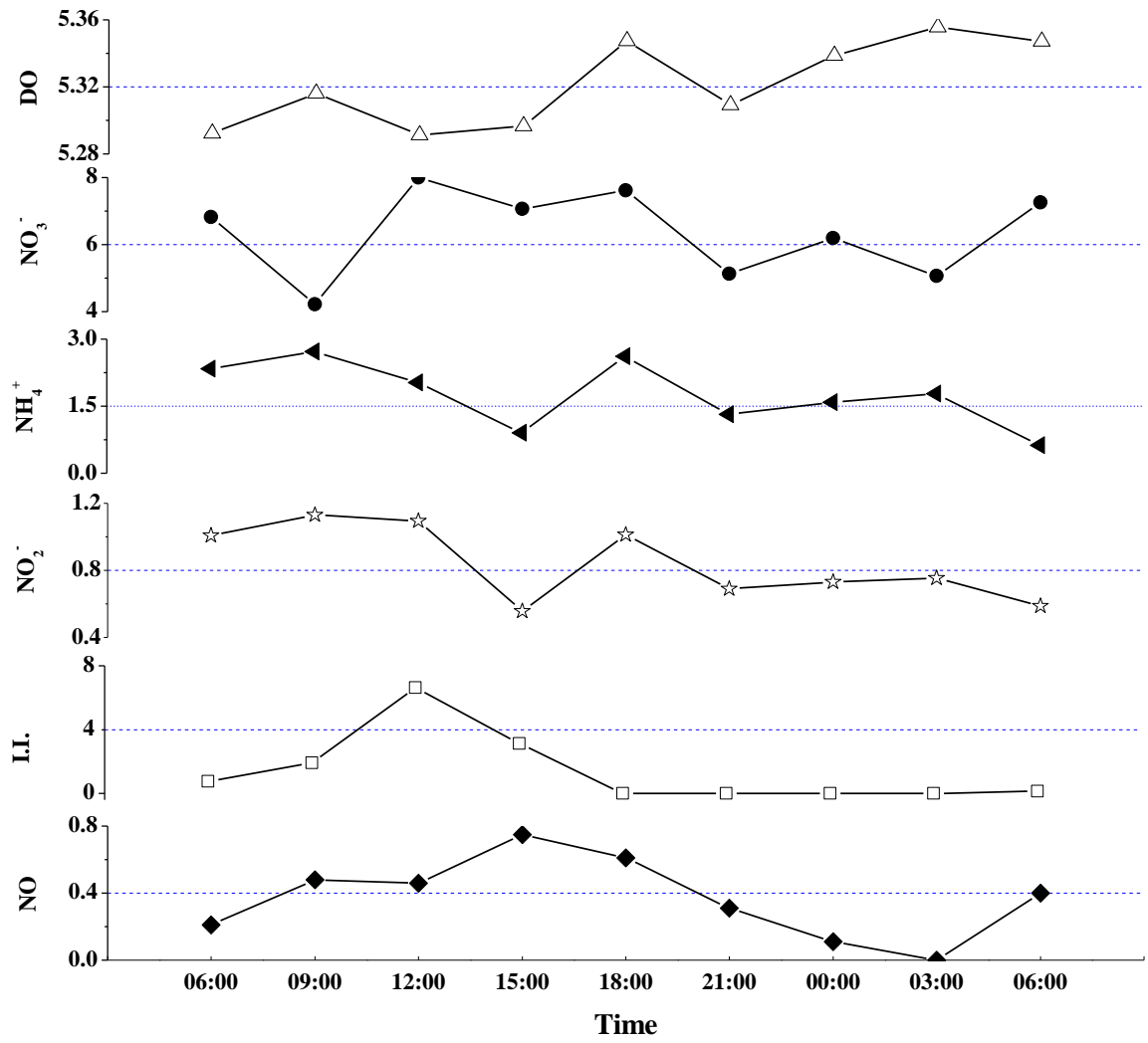
520

521 **Fig.5** The concentrations of NO in the surface waters off Qingdao (stations S01-S09) and Jiaozhou Bay

522 (stations S10 and S11)

523 D.L. stands for concentration below the detection limit.

524

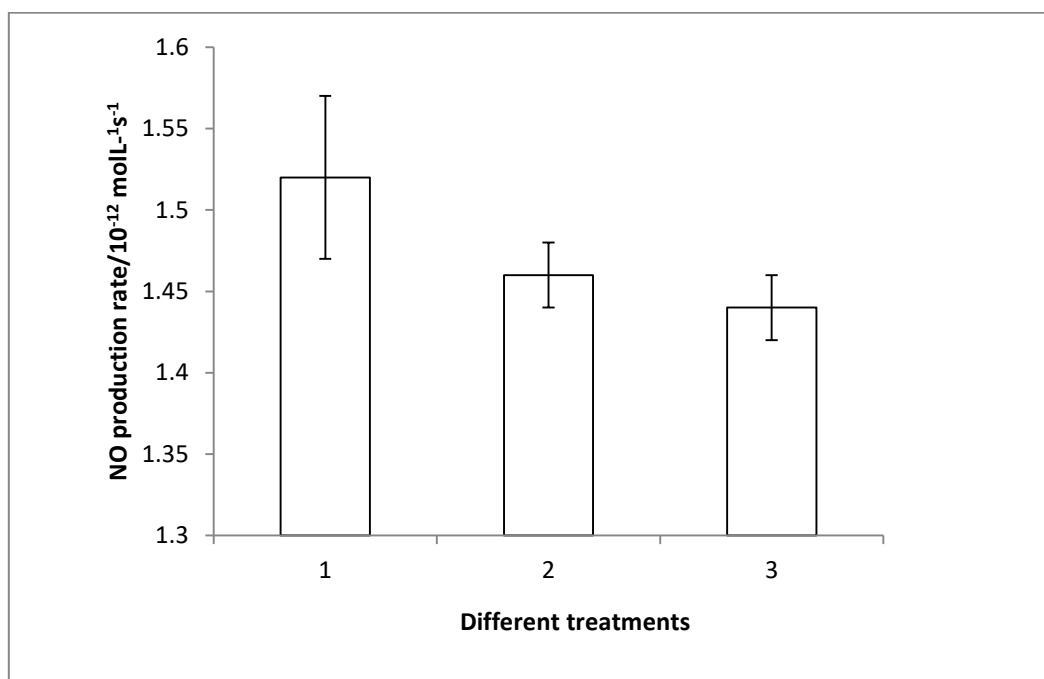


525

526

527 **Fig. 6** The diurnal variations of NO concentrations and related parameters in the surface seawater at
 528 station 10 (Units: DO (mL L⁻¹), NO₃⁻, NO₂⁻, NH₄⁺ (μmol L⁻¹), I.I.-illumination intensity (×10⁴ lux), NO
 529 (nmol L⁻¹))

530



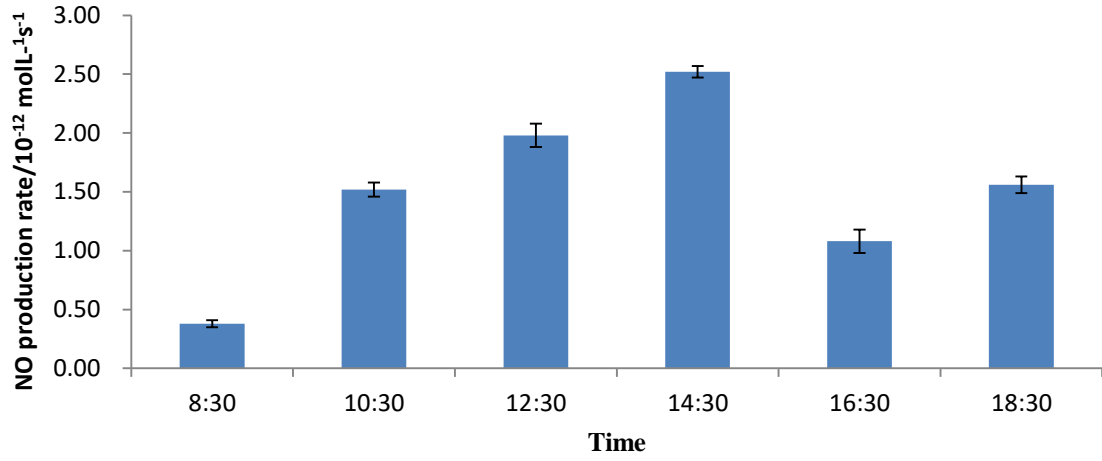
531

532 **Fig. 7** The production rates of NO by seawater irradiation under natural light after different treatments

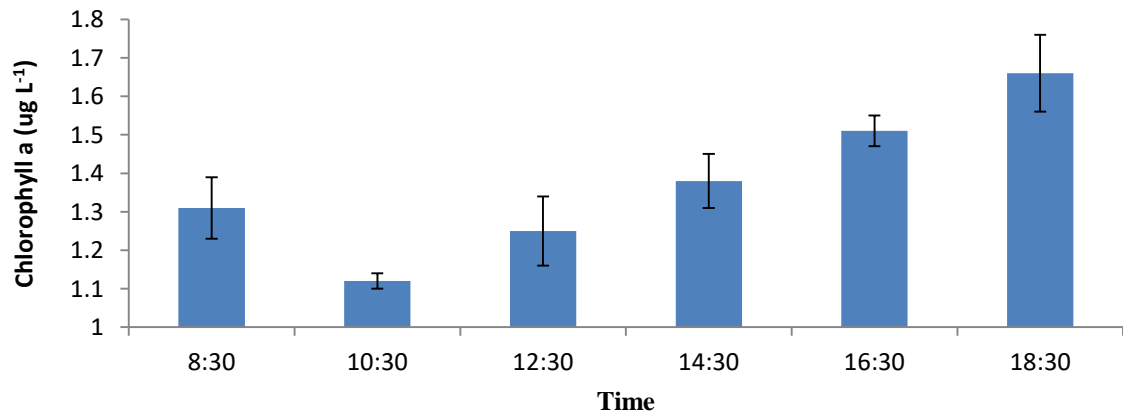
533 (1. Incubated on deck at ambient temperature, 2. 0.45 μm Millipore filtered at ambient temperature, 3.

534 Incubated in water bath supplied with surface seawater)

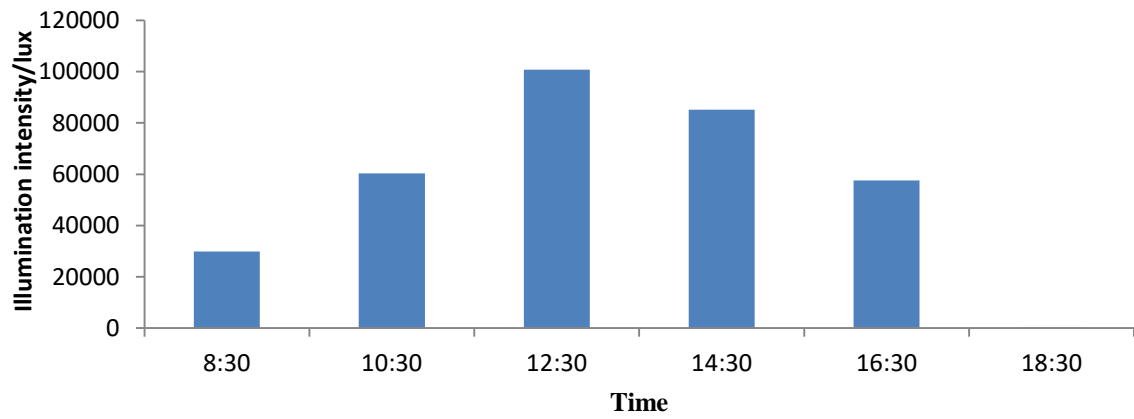
535



536



537



538

539 **Fig. 8** The variations of NO production rates, chlorophyll *a* concentrations and sunlight intensity in the
 540 incubation experiments with Qingdao coastal waters

541

542 Table 1 The methods for NO detection in seawater

| Method | Linearity range (nmol L ⁻¹) | Detection limit (nmol L ⁻¹) | Analytical precision | Reference |
|--|--|--|-------------------------|----------------------------------|
| Microelectrode | 140–9900 | 140 | 0.24% | Zhang et al. (2003) |
| Microelectrode | 1.4–1400 | 4.2×10 ⁻¹⁰ | 6.30% | Xing et al. (2005) |
| Microelectrode | 0.4–4000 | 30 | - | Schreiber et al.(2008) |
| Fluorescence | 1.4–1400 | 1.4 | 1.63% | Liu et al. (2009) |
| HPLC with fluorescence | 0.025–10 | 0.025 | 3-5% | Olasehinde et al. (2009) |
| Purge-and-trap with chemiluminescence | - | 0.0015 | 3% | Zafiriou and McFarland (1980) |
| Purge-and-trap with chemiluminescence | - | 0.25 | 3-25% | Bange and Lutterbeck (2015) |
| Purge-and-trap with fluorescence | 0.14–19 | 0.068 | 7.2% | This study |

543

544

545

Table 2 Reaction yields of the reaction of DAN with NO (in %)

| Purge flow rate /mL min ⁻¹ | Purge time /min | | | |
|---------------------------------------|-----------------|----|----|----|
| | 15 | 30 | 45 | 60 |
| 200 | — | — | — | — |
| 300 | — | — | 21 | 34 |
| 400 | 56 | 85 | 69 | 69 |
| 500 | — | — | 22 | 26 |
| 600 | — | — | 31 | 33 |

546