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# Temporal and spatial distribution of the meiobenthic community in Daya Bay, South China Sea

L. Tang<sup>1,2</sup>, H. X. Li<sup>1</sup>, and Y. Yan<sup>1</sup>

<sup>1</sup>CAS Key Laboratory of Marine Bio-resources Sustainable Utilization,  
Guangzhou 510301, China

<sup>2</sup>Graduate University of Chinese Academy of Sciences, Beijing 10039, China

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Correspondence to: Y. Yan (yany@scsio.ac.cn)

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## Abstract

Spatial and temporal biodiversity patterns of the meiobenthos were studied for the first time in Daya Bay, which is a tropical semi-enclosed basin located in the South China Sea. The abundance, biomass, and composition of the meiobenthos and the basic environmental factors in the bay were investigated. The following 19 taxonomic groups were represented in the meiofauna: Nematoda, Copepoda, Polychaeta, Oligochaeta, Kinorhyncha, Gastrotricha, Ostracoda, Bivalvia, Turbellaria, Nemertinea, Sipuncula, Hydroida, Amphipoda, Cumacea, Halacaroida, Priapulida, Echinodermata, Tanaidacea, and Rotifera. Total abundance and biomass of the meiobenthos showed great spatial and temporal variation, with mean values of  $993.57 \pm 455.36 \text{ ind cm}^{-2}$  and  $690.51 \pm 210.64 \mu\text{g } 10 \text{ cm}^{-2}$ , respectively. Nematodes constituted 95.60% of the total abundance and thus had the greatest effect on meiofauna quantity and distribution, followed by copepods (1.55%) and polychaetes (1.39%). Meiobenthos abundance was significantly negatively correlated with water depth at stations ( $r = -0.747$ ,  $P < 0.05$ ) and significantly negatively correlated with silt-clay content ( $r = -0.516$ ,  $P < 0.01$ ) and medium diameter ( $r = -0.499$ ,  $P < 0.01$ ) of the sediment. Similar results were found for correlations of biomass and abundance of nematodes with environmental parameters. Polychaete abundance was positively correlated with the bottom water temperature ( $r = 0.456$ ,  $P < 0.01$ ). Meiobenthos abundance differed significantly among seasons ( $P < 0.05$ ), although no significant difference among stations and the interaction of station  $\times$  season was detected by two-way ANOVA. In terms of vertical distribution, most of the meiobenthos was found in the surface layer of sediment. This pattern was apparent for nematodes and copepods, but a vertical distribution pattern for polychaetes was not as obvious. Based on the biotic indices and analyses of their correlations and variance, the diversity of this community was likely to be influenced by environmental variations.

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# 1 Introduction

Meiofauna are defined as metazoan invertebrates that are smaller than 500  $\mu\text{m}$  in size and are retained on sieve meshes of 31–64  $\mu\text{m}$  (Coull and Chandler, 2001). Meiofauna are effective decomposers of organic material (Giere, 1993) and active consumers of benthic bacteria and microalgae, thus they are intimately associated with the benthic marine environment (Heip et al., 1985). Meiofauna exhibit high abundance, diversity, and productivity in most sedimentary habitats, and they play an important role in marine benthic food chains due to their small size, high abundance, fast turnover rates, and rapid generation times (Heip et al., 1985; Gee, 1989; Coull, 1999). Because of their biological characteristics, meiofaunal communities can be used to monitor marine environments, evaluate the effects of human activity on marine environments, and verify hypotheses about different ecosystems (Schratzberger et al., 2004; Veit-Kahler et al., 2008).

Daya Bay is a semi-enclosed bay located in the Northern South China Sea (from 113°29'42" to 114°49'42" E, 23°31'12" to 24°50'00" N). Its coastal line is 92 km, and it has an area of 600 km<sup>2</sup> (Wang et al., 2006, 2008). The water depth ranges from 6 to 16 m, with an average of 10 m (Xu, 1989; Wang et al., 2006, 2008; Wu and Wang, 2007). It is a typical subtropical bay in the South China Sea, and it is important because of its natural resources and habitat diversity (e.g., mangroves and coral communities). Daya Bay and its adjacent areas have become an important economic development district, in which molluscs and fish are being cultivated (Wang et al., 2008). Wang et al. (2008) reported that the ecological environment of Daya Bay changed substantially between 1982 and 2004, with an increased average N/P ratio (from 1.377 to 49.09), decreased numbers of phytoplankton species (from 159 species, 46 genera to 126 species, 44 genera) and zooplankton species (from 46 species to 36 species), and reduced megabenthos biomass. However, meiofauna were not included in that investigation, and the biomass and distribution of meiobenthic organisms in Daya Bay were unknown.

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In this study, the composition, abundance, biomass, diversity, and distribution of the meiofauna were investigated in Daya Bay from October 2009 to August 2010. The relationships among these parameters and basic environmental factors were analyzed to assess the biodiversity patterns of the meiobenthos in this ecosystem and to evaluate the potential response of the meiofauna to changes in the ecological environment of Daya Bay.

## 2 Materials and methods

### 2.1 Field studies

Field sampling of sediment was conducted at quarterly intervals (October 2009 (autumn), January 2010 (winter), April 2010 (spring), and August 2010 (summer) at nine stations in Daya Bay (S4, S6, S7, S8, S10, S11, S12, S14, and S15), which were located using a global positioning system (Fig. 1). At each station, three replicate sediment samples were collected using a 0.1 m<sup>2</sup> Gray-Ohara box grab. When the box grab was onboard the ship, one subsample from each grab was collected for the meiofauna analysis using a cut-off syringe (2.9 cm diameter, 6.6 cm<sup>2</sup> surface area) pushed to 10 cm depth in the sediment. The sample was divided into three vertical layers (0–2, 2–5, and 5–10 cm) and then was fixed with 4% formalin separately. A core was taken from each of the three box grabs at a given site at each time point to promote randomness in the sampling process except in January 2010. Another subsample of the surface sediment from each box grab was taken for analysis of water content and particle size, and three subsamples were taken for analysis of water content, chlorophyll-*a* (Chl-*a*), total organic carbon and granularity of sediment. Sediment samples for environmental analysis were sorted into PVC valve bags, placed in an ice box on board, and later transferred to the laboratory for storage at –20 °C until analysis. Temperature, salinity of bottom water, and the depth of each station were measured at three regions within each station at each sampling time point using a CTD.

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## 2.2 Laboratory procedures

Meiofauna were extracted from the sediment using the Ludox™ centrifugation technique (Heip et al., 1985). All biological core samples were stained with Rose Bengal for at least 24 h and then washed with tap water under a set of sieves with mesh sizes of 38 μm to 500 μm. Meiofauna were extracted from the remaining sand particles retained on the smallest mesh size by centrifugation with colloidal silica polymer (Ludox™-50, 1.15 g cm<sup>-3</sup>, Sigma-Aldrich, St. Louis, Mo, USA). Centrifugation was repeated three times at 1800 r for 10 min each time. After each centrifugation, the floating matter was washed and decanted into a lined Petri dish, and the meiofauna were sorted, identified, and counted under a stereoscopic microscope (Widbom, 1984; Higgins and Thiel, 1988; Giere, 1993).

The sediment samples, except those for water content and particle size analysis, were frozen at -20 °C for 24 h, followed by freeze-drying. The freeze-dried sediment was homogenized manually and stored in desiccators for further analyses. The samples for Chl-*a* was placed in a 50 ml polyethylene bottle with 30 ml of 90 % acetone for the extraction of pigments. Chl-*a* concentrations were measured using a 10- AU Turner Designs fluorometer (Turner Designs, Sunnyvale CA, USA) fitted with the narrow-band, non acidification system of Welschmeyer (1994) (Nozais et al., 2001). The total organic carbon (TOC) in these samples was estimated using the wet oxidation method (El-Wakeel and Riley, 1957), and then TOC content was then converted to organic matter (OM) content (Wiseman and Bennette, 1960). Fresh wet sediment samples with a known weight were dried at 105 °C for 24 h and the final dry weight was measured. The water content was calculated as the sediment weight loss before and after drying. The particle size of sediments was determined in the Granularity Laboratory of the South China Sea Institute of Oceanology, Chinese Academy of Sciences using the Masterizer2000 (Malvern, Worcestershire, UK, 0.02–2000 μm).

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## 2.3 Statistics

All meiofaunal abundance data were expressed as individuals per 10 cm<sup>2</sup> for each sample, and the biomass values of different taxonomic groups were calculated based on their dry weight following Gerlach (1971) and Widbom (1984).

The Shannon-Wiener diversity index ( $H'$ ), the Margalef's index ( $D$ ) and the Pielou's evenness ( $J'$ ) were used in characterization of meiofaunal community structure:

$$H' = - \sum n_i / N \log n_i / N$$

$$D = (S - 1) / \ln RN$$

$$J' = H' / H'_{\max}$$

Where  $n_i$  – is community density of each taxonomic groups,  $N$  – total density of communities,  $S$  – the number of taxonomic group.

Biological data were analyzed using univariate and multivariate methods. Pearson's correlation analysis was performed to examine the relationship among environmental factors and the relationship between meiofauna and environmental variables. Statistical differences among stations and seasons were tested by two-way analysis of variance (ANOVA) using the quantitative information about the meiofaunal assemblages (i.e., number of taxonomic groups, abundance, biomass, and biotic indices). These analyses were performed using the SPSS 17.0 statistical software package. The multivariate analysis followed the standard methods described in Clarke (1993) and used the PRIMER software 5.0 statistical package (Plymouth Routines in Multivariate Ecological Research). Environmental variables were first subjected to principal components analysis (PCA), to allow the identification of those most responsible for temporal and spatial variations. The Bray-Curtis similarity measure was used, after square root transformation of the data (Clarke and Gorley, 2006), for CLUSTER analysis and two-dimensional multidimensional scaling (MDS) ordination of environmental and meiofaunal parameters at the nine stations at four different time points (seasons). The map of the stations,

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following five groups showed a similarity level of 95 %: (1) S6 for all four seasons, S10 in autumn, spring, and summer, and S11 and S8 in autumn, winter, and summer constituted group 1; (2) S12 for all four seasons, S14 in autumn, winter, and spring, and S4, S7, and S10 in winter made up group 2; (3) S15 for all four seasons and S14 in summer represented group 3; (4) S4, S8, and S11 in spring made up group 4; and (5) S7 in autumn, spring, and summer and S4 in autumn and summer composed group 5 (Fig. 6).

### 3.2 Taxonomic composition, abundance, and biomass of meiofauna

The following 19 taxonomic groups were represented in the meiofauna collected at the nine stations during the four seasonal sampling events: Nematoda, Copepoda, Polychaeta, Oligochaeta, Kinorhyncha, Gastrotricha, Ostracoda, Bivalvia, Turbellaria, Nemertinea, Sipuncula, Hydroida, Amphipoda, Cumacea, Halacaroidea, Priapulida, Echinodermata, Tanaidacea, and Rotifera (Appendice). The average abundance and biomass of meiofauna collected from the nine stations during the four seasons were  $993.57 \pm 455.36 \text{ ind cm}^{-2}$  and  $690.51 \pm 205.0 \mu\text{g } 10 \text{ cm}^{-2}$ , respectively. Nematodes accounted for 95.60% of the abundance of meiofauna. Copepods and polychaetes ranked second in abundance (1.55% and 1.39%, respectively). Nematodes constituted 55.02% of biomass, followed by polychaetes at 28.01%. The number of taxonomic group, abundance, and biomass of the meiobenthos from Daya Bay all differed significantly among stations and seasons (two-way ANOVA,  $P < 0.001$ ).

### 3.3 Temporal distribution of meiofauna

In autumn, the highest and lowest abundance of meiobenthos occurred at S4 and S8, with  $939.7 \text{ ind } 10 \text{ cm}^{-2}$  and  $233.2 \text{ ind } 10 \text{ cm}^{-2}$ , respectively. Nematodes were dominant at 92.28% (range, 224.7–821.5 ind  $10 \text{ cm}^{-2}$ ), followed by polychaetes at 2.05% (range, 2.5–39.4 ind  $10 \text{ cm}^{-2}$ ). In winter, the highest and lowest abundance of meiobenthos was found at S15 and S10, with  $1088.08 \text{ ind } 10 \text{ cm}^{-2}$  and  $424.63 \text{ ind } 10 \text{ cm}^{-2}$ , respectively.

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Nematodes were dominant at 94.98 % (range, 393.66–1021.60 ind 10 cm<sup>-2</sup>), followed by copepods at 2.26 % (range, 0.67–39.21 ind 10 cm<sup>-2</sup>). In spring, the highest and lowest abundance of meiobenthos occurred at S4 and S7, with 2301.88 ind 10 cm<sup>-2</sup> and 650.83 ind 10 cm<sup>-2</sup>, respectively. Nematodes were dominant at 97.63 % (range, 623.06–2240.79 ind 10 cm<sup>-2</sup>), followed by copepods at 0.91 % (range, 3.03–26.26 ind 10 cm<sup>-2</sup>). In summer, the highest and lowest abundance of meiobenthos was found at S11 and S15, with 1895.43 ind 10 cm<sup>-2</sup> and 313.80 ind 10 cm<sup>-2</sup>, respectively. Nematodes were dominant at 94.87 % (range, 294.11–1844.94 ind 10 cm<sup>-2</sup>), followed by polychaetes at 2.47 % (range, 4.54–111.08 ind 10 cm<sup>-2</sup>). The abundance of total meiobenthos, nematodes, and polychaetes differed significantly among stations and months; however, this was not true for copepod abundance. The interaction of station × season was significant difference for polychaete abundance from ANOVA results; however for abundance of total meiobenthos, nematodes, and copepod showed different trends.

### 3.4 Spatial distribution of meiofauna

#### 3.4.1 Horizontal distribution

The highest mean abundance of meiobenthos of the four seasons occurred at S4 (1397.81 ± 725.76 ind 10 cm<sup>-2</sup>), followed by S11 (1294.12 ± 850.32 ind 10 cm<sup>-2</sup>). The lowest mean abundance was found at S7 (623.09 ± 135.04 ind 10 cm<sup>-2</sup>) (Fig. 7a). The mean abundance of meiofauna of seasons differed significantly among stations. The horizontal distribution of nematodes showed the same trend as that of the total meiofauna, with the highest abundance at S4 (1325.73 ± 738.76 ind 10 cm<sup>-2</sup>) and the lowest at S7 (577.86 ± 146.07 ind 10 cm<sup>-2</sup>) (Fig. 7b). The highest abundance of copepods occurred at S4 (20.38 ind 10 cm<sup>-2</sup>), followed by value for S15, and the lowest abundance was found value at S8 (Fig. 7c). The highest abundance of polychaetes occurred at S8 (29.49 ind 10 cm<sup>-2</sup>) and the lowest was value at S6 (Fig. 7d).

### 3.4.2 Vertical distribution

The vertical distribution of meiobenthos at the nine stations was studied in all seasons except winter (Fig. 8). The greatest abundance of meiobenthos (50.77%) occurred in the upper layer (0–2 cm), and the number of organisms gradually declined with depth (31.63% in the 2–5 cm layer and 17.60% in the 5–10 cm layer). As the dominant group, the distribution of nematodes was highly similar to that of the total meiobenthos: 50.77%, 30.55%, and 18.68% were found in the 0–2 cm, 2–5 cm, and 5–10 cm layers, respectively. A large proportion of the copepods occurred in the upper layer (74.27%), and the percentages in the deeper two layers were similar (12.56% and 13.16%, respectively). Almost half of the polychaetes (50.89%) were found in the upper layer, with 22.17% in the 2–5 cm layer and 26.94% in the 5–10 cm layer. The taxonomic composition and abundance of the meiobenthos and the abundance of nematodes and copepods were significantly different among the vertical layers ( $P < 0.001$ ). Moreover, the vertical distribution of the taxa, the abundance of meiobenthos, and the abundance of nematodes differed significantly among seasons ( $P < 0.001$ ).

### 3.5 Distribution of meiobenthos biomass

The temporal and spatial distribution of biomass was similar to that of abundance. The highest mean biomass of all stations occurred in spring ( $764.73 \pm 596.03 \mu\text{g } 10 \text{ cm}^{-2}$ ), and the lowest value occurred in winter ( $486.64 \pm 157.04 \mu\text{g } 10 \text{ cm}^{-2}$ ). The highest mean biomass of the meiobenthos of four seasons was  $1121.22 \pm 358.64 \mu\text{g } 10 \text{ cm}^{-2}$  at S4, and the lowest value was  $395.10 \pm 102.44 \mu\text{g } 10 \text{ cm}^{-2}$  at S6. The biomass of meiobenthos differed significantly among stations, seasons, and interaction of station  $\times$  season. In terms of vertical distribution, more than half (51.95%) of the biomass was found in the 0–2 cm layer, with 26.99% and 21.64% in the 2–5 cm and 5–10 cm layers, respectively. Thus, the biomass differed significantly among layers. Pearson correlation analysis showed that the biomass in the 0–2 cm layer was significantly positively correlated with the abundance of nematodes, copepods, polychaetes, and total meiobenthos in

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the 0–2 cm layer. The biomass in the 2–5 cm layer also was significantly positively correlated with the abundance of nematodes, copepods, polychaetes, and total meiobenthos in the 2–5 cm layer. The biomass in the 5–10 cm layer was significantly positively correlated with the abundance of nematodes, copepods, and total meiobenthos in the 5–10 cm layer, but not with polychaete abundance.

### 3.6 Correlations between the meiobenthos and environmental parameters

The number of taxonomic group was significantly negatively correlated with the bottom water temperature. Meiobenthos abundance was significantly negatively correlated with water depth at stations and significantly negatively correlated with silt-clay content and Md of the sediment. Similar results were found for correlations of biomass and abundance of nematodes with environmental parameters. Polychaete abundance was positively correlated with the bottom water temperature. There were no significant correlations between the other biotic variables and environmental parameters. Pearson correlation analyses of diversity indices and environmental parameters indicated that  $H'$  and  $D$  at the stations were significantly negatively correlated with the water content of the sediment, and  $J'$  was significantly negatively correlated bottom water temperature (Table 2).

Ordination of the samples using the MDS technique showed meiobenthos assemblages was similar by season than by station (Fig. 9). Some samples belonging to the same season were clustered together (e.g., April), but samples from different stations were characterized by higher variability. Because nematodes constituted the largest proportion of the total meiobenthos abundance, two-way ANOVA showed the same results that biotic variations include the taxonomical groups, abundance and biomass among seasons were significant difference, but not different on stations and interaction station  $\times$  month likewise.

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## 4 Discussion

This study was the first to explore statistically the ways in which community composition, abundance, biomass, and distribution of the meiobenthos differ throughout Daya Bay over the course of a year. This approach enabled exploration of which environmental variables were mainly responsible for influencing the spatial and temporal distributions of the meiobenthos. Integration of the results provided a quantitative description of the biodiversity patterns of the meiobenthos on temporal and spatial (horizontal and vertical) scales as well as a relatively good characterization of the environment.

Nineteen taxonomic groups of meiofauna were found in Daya Bay, which was higher than in Ha Long Bay, South China Sea (11 groups) (Olga et al., 2008) and lower than in Nha Trang Bay, South China Sea (26 groups) (Olga et al., 2006). Free-living marine nematodes constituted as much as 95.60% of the total individuals for the four seasons studied herein, followed by copepods (1.55%). This finding is in agreement with results of previous studies conducted in the South China Sea (Olga et al., 2006, 2008; Liu, 2009) and other areas (Olafsson, 1997; Armenteros et al., 2009). The mean abundance of meiofauna ( $993.57 \pm 455.36 \text{ ind cm}^{-2}$ ) in Daya Bay was similar to that at Cienfuegos in the Caribbean ( $780.02 \pm 772 \text{ ind } 10 \text{ cm}^{-2}$ ) (Armenteros et al., 2009) and in Nha Trang Bay, South China Sea ( $944.3 \pm 303.7 - 1034.6 \pm 435.8 \text{ ind } 10 \text{ cm}^{-2}$ ) (Olga et al., 2006) and higher than that reported for Victoria Bay in Hong Kong ( $182.7-290.2 \text{ ind } 10 \text{ cm}^{-2}$ ) (Liu, 2010) and for Ha Long Bay, South China Sea ( $180.8 \pm 13.1 \text{ ind } 10 \text{ cm}^{-2}$ ) (Olga et al., 2008). The abundance of meiobenthos found in the present study was lower than the  $3275 \pm 701 \text{ ind } 10 \text{ cm}^{-2}$  reported for the southwest lagoon of New Caledonia (Boucher, 1997), the  $1524-6094 \text{ ind } 10 \text{ cm}^{-2}$  found in the Yellow Sea (Kim et al., 2000), and the  $810 \pm 410 \text{ ind } 10 \text{ cm}^{-2}$  collected from the East China Sea (Zhang et al., 2001, 2002), but there were no distinct differences among the taxonomic groups identified.

Our results demonstrated that the community composition and abundance of the meiofauna in Daya Bay are influenced markedly by both station and season. In terms of

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by trawling cannot be ignored, as it leads to release of suspended sediments (Churchill, 1989; Palanques et al., 2001), resuspension and burial of biologically recyclable organic materials (Mayer et al., 1991), and release of nutrients to the overlying water (Pilskaln et al., 1998; de Madron et al., 2005). Second, in Aotou Bay (near S4 and S15), red tides usually appear in spring and summer every year (Song et al., 2004), and the affected area often is limited to the northern waters of the bay around the cage culture area. Many environmental factors can cause red tides; even in Daya Bay the cause of each red tide may not be exactly the same (Li et al., 1993; Xu et al., 2001a, b). The abrupt change in primary production that occurs during an algal bloom would affect the quantity and structure of the meiobenthos (Armenteros et al., 2009).

Distribution of the biomass of the meiobenthos basically followed that of abundance, and the observed differences between the two to a large extent were due to the proportion of polychaetes. Polychaetes exhibited seasonal variation, as they were the second large taxonomic group in autumn and summer but not in winter and spring. This result differs from most other research. In this study, polychaetes were sensitive to water temperature and the granularity of sediment (i.e., positively correlated with temperature and significantly negatively correlated with the percentage of clay-silt and Md of the sediment), and they exhibited significant differences among stations and seasons and the interaction of station  $\times$  season. The seasonal change in algae mentioned above would impact the quantity of copepods, which mainly feed on phytoplankton (Blanchard, 1991), and copepods are more affected than other meiofaunal by pollution in the environment (Raffaelli and Mason, 1981; Findlay, 1982). The dry weights of copepods (DW = 1.86) and polychaetes (DW = 14) differ greatly (Widbom, 1984), which lead to different distributions of biomass and abundance.

Diversity indices can be used as an indication of the diversity of a biological community, and the diversity of a community reflects its stability to some extent. The meiofaunal diversity indices of Daya Bay were low because of the large percentage of nematodes present (95.60%). The Shannon-Wiener index, Simpson domination index, and Pielou's evenness index were significantly correlated with sediment granularity

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and water content of the sediment, and they were different in different seasons. This result suggests that the diversity of the community likely is influenced by environmental variations.

## 5 Conclusions

This was the first study of the abundance, biomass, and temporal and spatial distribution of the meiobenthos in Daya Bay, South China Sea. The key results were as follows:

- (1) Depth did not differ significantly among stations, and water temperature did differ significantly among seasons. Silt-clay and silt-sand were the main sediment types in Daya Bay. Water content of the sediment was influenced by granularity and was significantly negatively correlated with water depth. The content of Chl-*a* and OM did not differ significantly among stations and seasons;
- (2) The absolute dominant taxonomic group was Nematoda, followed by Copepoda and Polychaeta. Community structure and abundance were significantly correlated with granularity of the sediment among stations, and had correlation with bottom water temperature among seasons;
- (3) The biomass of the meiobenthos depended on the proportion of copepods/polychaetes, and it was significantly correlated with the percentage of clay-silt and Md of the sediment;
- (4) Most of the meiobenthos were found in the surface layer of the sediment, and as the largest group, nematodes had the main effect on quantity and proportion of the vertical distribution of the meiofauna;
- (5) These results suggest that the meiobenthos was influenced by anthropogenic activities (e.g., trawling, red tides), which were apparent as seasonal changes in environmental parameters.

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**Table 2.** Correlation between biotic variables and environmental parameters of Daya Bay.

	Depth	Temperature	Salinity	Water content	OM content	Silt-clay content	Md	Chl- <i>a</i>
NO. of taxonomic group	0.301	-0.747**	0.137	-0.120	-0.031	0.039	-0.006	-0.294
Abundance of meiofauna	-0.361*	0.224	-0.130	-0.016	-0.079	-0.516**	-0.499**	-0.186
Biomass of meiofauna	-0.333*	0.243	-0.095	-0.256	-0.047	-0.375*	-0.381*	-0.091
Abundance of nematodes	-0.359*	0.210	-0.129	0.001	-0.078	-0.518**	-0.500**	-0.194
Abundance of copepods	-0.457	-0.019	0.0	-0.157	-0.223	-0.196	-0.174	-0.121
Abundance of polychaetes	-0.239	0.456**	-0.135	-0.137	-0.002	0.035	0.022	0.184
H'	0.210	-0.003	0.114	-0.450**	0.063	0.085	0.047	0.147
D	0.306	-0.775**	0.126	-0.074	0.024	0.157	0.116	-0.222
J'	0.120	0.142	0.047	-0.430**	0.085	0.129	0.094	0.239

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**Table A1.** Average abundance and biomass of major meiobenthic groups collected at four different seasons.

Group	Season	Abundance		Biomass		Group	Season	Abundance		Biomass	
		ind/10 cm <sup>2</sup>	%	μg/10 cm <sup>2</sup>	%			ind/10 cm <sup>2</sup>	%	μg/10 cm <sup>2</sup>	%
Nematoda	Autumn	546.7 ± 212.0	95.60	218.7 ± 84.8	55.02	Copepoda	Autumn	11.7 ± 10.1	1.55	21.8 ± 18.7	4.16
	Winter	653.2 ± 182.1		261.3 ± 72.9			Winter	15.6 ± 10.7		28.9 ± 19.9	
	Spring	1552.0 ± 564.1		620.8 ± 225.6			Spring	14.4 ± 9.0		26.8 ± 16.8	
	Summer	1047.6 ± 543.0		419.1 ± 217.2			Summer	20.0 ± 9.5		37.3 ± 17.6	
Polychaeta	Autumn	12.2 ± 11.5	1.39	170.3 ± 161.1	28.01	Oligochaeta	Autumn	5.61 ± 6.45	0.29	78.6 ± 90.3	5.92
	Winter	6.2 ± 4.2		86.1 ± 59.3			Winter	3.3 ± 3.2		46.3 ± 45.3	
	Spring	9.7 ± 5.6		135.1 ± 78.0			Spring	2.1 ± 1.9		29.1 ± 26.1	
	Summer	27.3 ± 34.1		382.1 ± 477.7			Summer	0.7 ± 1.7		9.4 ± 23.2	
Kinorhyncha	Autumn	2.1 ± 3.2	0.25	4.2 ± 6.3	0.70	Gastrotricha	Autumn	6.3 ± 9.2	0.20	21.6 ± 32.5	0.97
	Winter	2.9 ± 2.2		5.8 ± 4.5			Winter	0.3 ± 0.5		1.1 ± 1.7	
	Spring	2.92 ± 1.61		5.8 ± 3.2			Spring	0.3 ± 0.4		1.2 ± 1.3	
	Summer	1.9 ± 1.7		3.4 ± 3.4			Summer	0.8 ± 1.7		3.0 ± 3.6	
Ostracoda	Autumn	1.1 ± 1.5	0.18	28.9 ± 39.4	6.57	Bivalvia	Autumn	2.2 ± 2.0	0.17	9.4 ± 8.4	1.04
	Winter	1.0 ± 0.9		25.8 ± 23.5			Winter	0.8 ± 1.0		3.5 ± 4.4	
	Spring	4.2 ± 5.4		109.4 ± 140.3			Spring	0.7 ± 1.5		3.1 ± 6.2	
	Summer	0.7 ± 0.8		17.5 ± 20.5			Summer	3.1 ± 2.0		12.8 ± 8.6	
Turbellaria	Autumn	0.4 ± 1.2	0.10	1.6 ± 4.1	0.49	Nemertinea	Autumn	1.1 ± 0.8	0.09	3.9 ± 2.7	0.45
	Winter	2.2 ± 2.8		7.5 ± 9.8			Winter	0.7 ± 0.8		2.3 ± 2.7	
	Spring	1.2 ± 1.2		4.3 ± 4.3			Spring	1.0 ± 0.9		3.3 ± 3.2	
	Summer						Summer	0.8 ± 1.5		2.9 ± 5.2	
Sipuncula	Autumn	0.8 ± 0.8	0.06	2.3 ± 2.6	0.30	Hydroida	Autumn	0.7 ± 1.9	0.04	2.6 ± 6.5	0.19
	Winter	0.4 ± 0.3		1.2 ± 1.2			Winter	0.1 ± 0.1		0.5 ± 0.5	
	Spring	0.5 ± 0.8		1.6 ± 2.9			Spring	0.3 ± 0.4		1.0 ± 1.3	
	Summer	0.8 ± 1.8		2.8 ± 6.1			Summer	0.3 ± 0.7		1.1 ± 2.4	
Amphipoda	Autumn	0.3 ± 1.0	0.03	0.1 ± 0.1	0.11	Cumacea	Autumn	0.7 ± 2.0	0.02	2.4 ± 6.7	0.04
	Winter	2.2 ± 2.8		11.2 ± 5.8			Winter	0.0 ± 0.1		0.6 ± 1.0	
	Spring	0.1 ± 0.2		0.8 ± 2.5			Spring				
	Summer	0.1 ± 0.3		1.3 ± 3.8			Summer				
Halacaroida	Autumn	0.3 ± 0.7	0.03	0.5 ± 1.1	0.03	Others	Autumn	0.1 ± 0.2	0.01	0.2 ± 0.2	0.04
	Winter	0.0 ± 0.1		0.0 ± 0.1			Winter	0.07 ± 0.09		0.1 ± 0.09	
	Spring	0.1 ± 0.2		0.2 ± 0.4			Spring	0.1 ± 0.2		0.2 ± 0.2	
	Summer						Summer	0.1 ± 0.2		0.2 ± 0.6	
Total			Abundance					Biomass			
			993.57 ± 455.36 ind cm <sup>-2</sup>					690.51 ± 205.0 μg 10 cm <sup>-2</sup>			

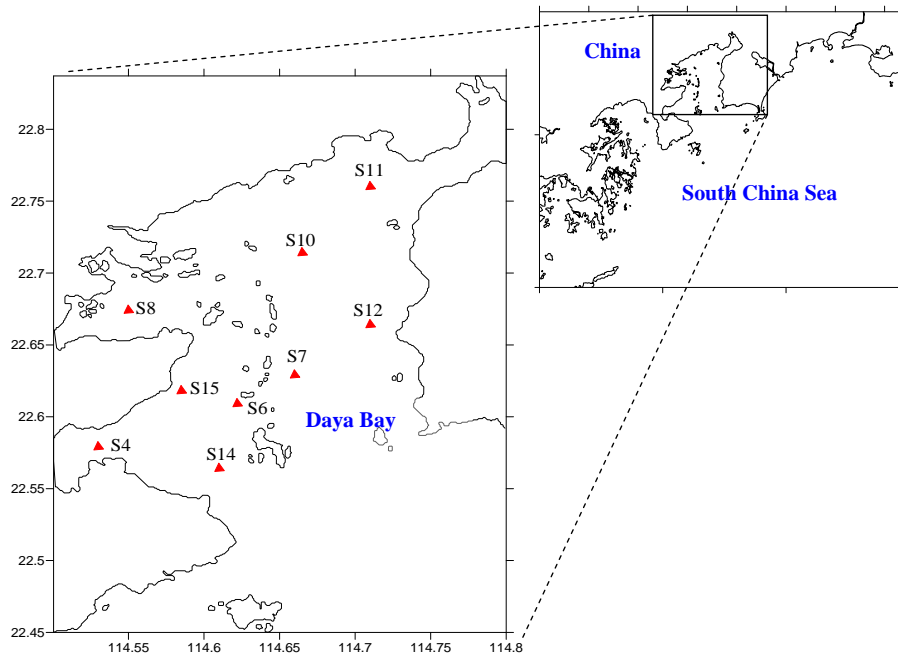
(Others include Priapulida, Echinodermata, Tanaidacea, and Rotifera, which were rarely found.)

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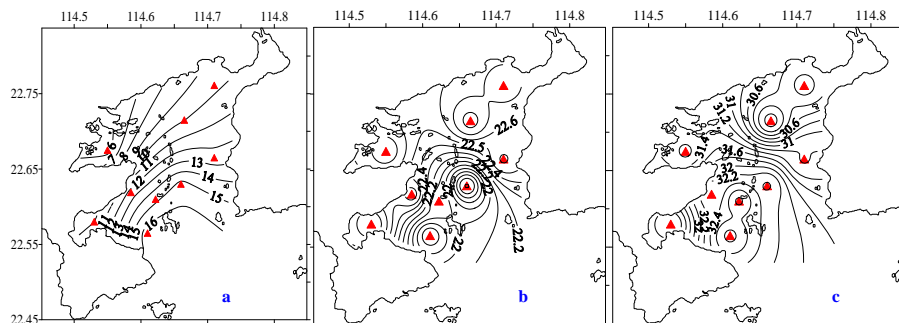


**Fig. 1.** Map of study area in Daya Bay with the locations of the nine sampling stations.

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**Fig. 2.** The horizontal distribution of **(a)** depth (m), **(b)** temperature ( $^{\circ}\text{C}$ ), and **(c)** salinity ( $\%$ ) in Daya Bay.

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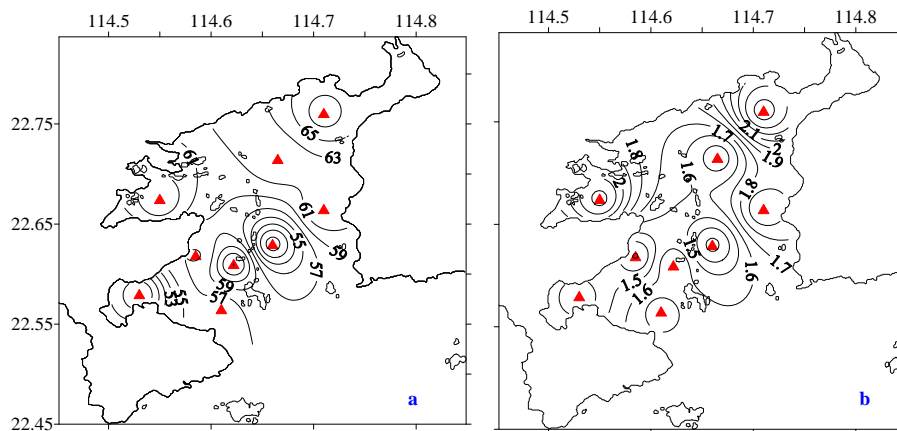
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**Fig. 3.** The horizontal distribution of **(a)** water content (%) and **(b)** OM (%) in sediment samples collected in Daya Bay.

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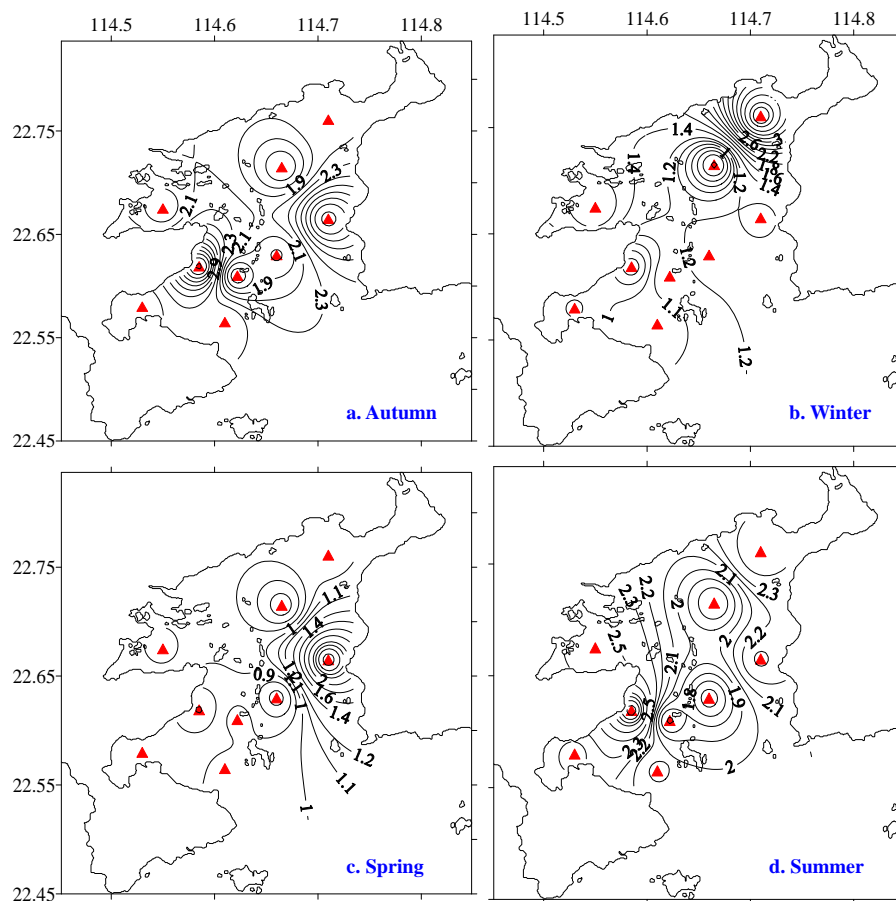
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**Fig. 4.** The horizontal distribution of Chl-a in sediments collected in Daya Bay at four different time points.

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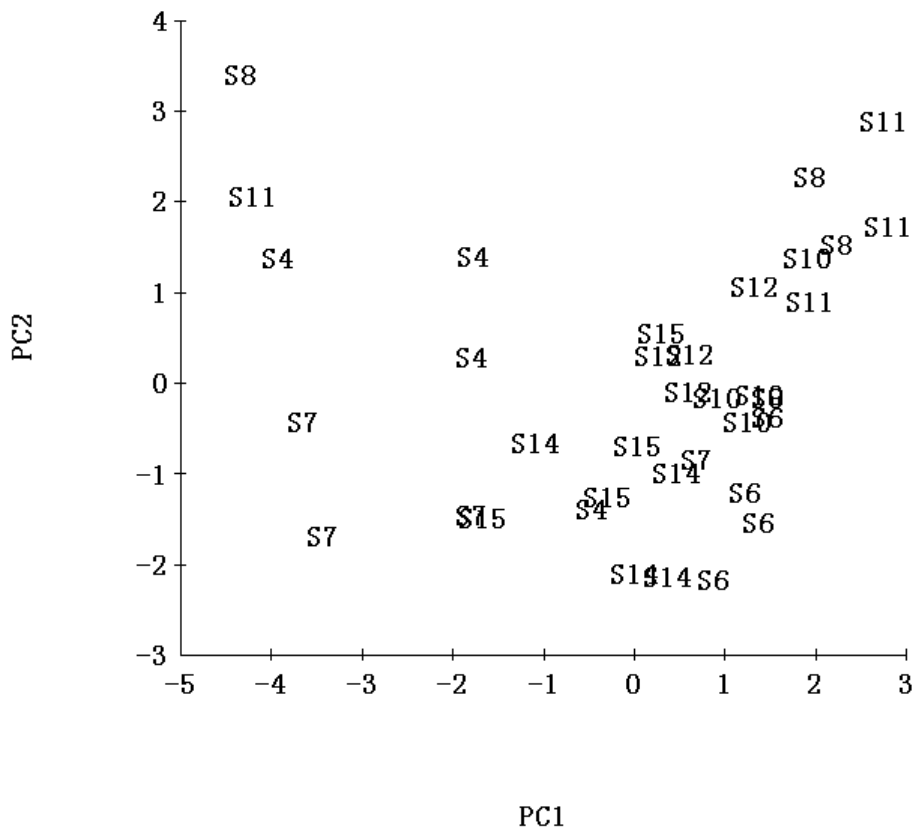
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**Fig. 5.** Ordination of nine stations for four seasons based on principal component analysis.

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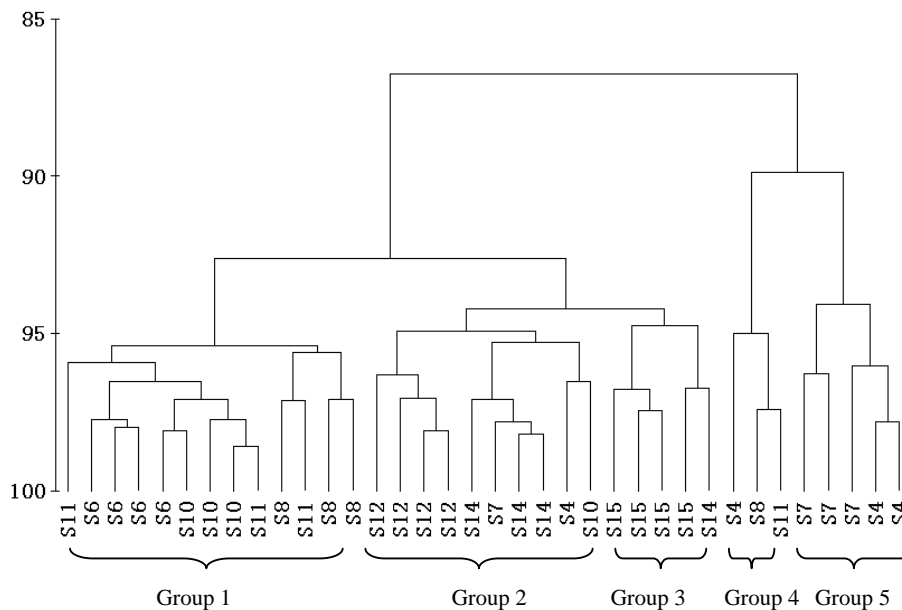
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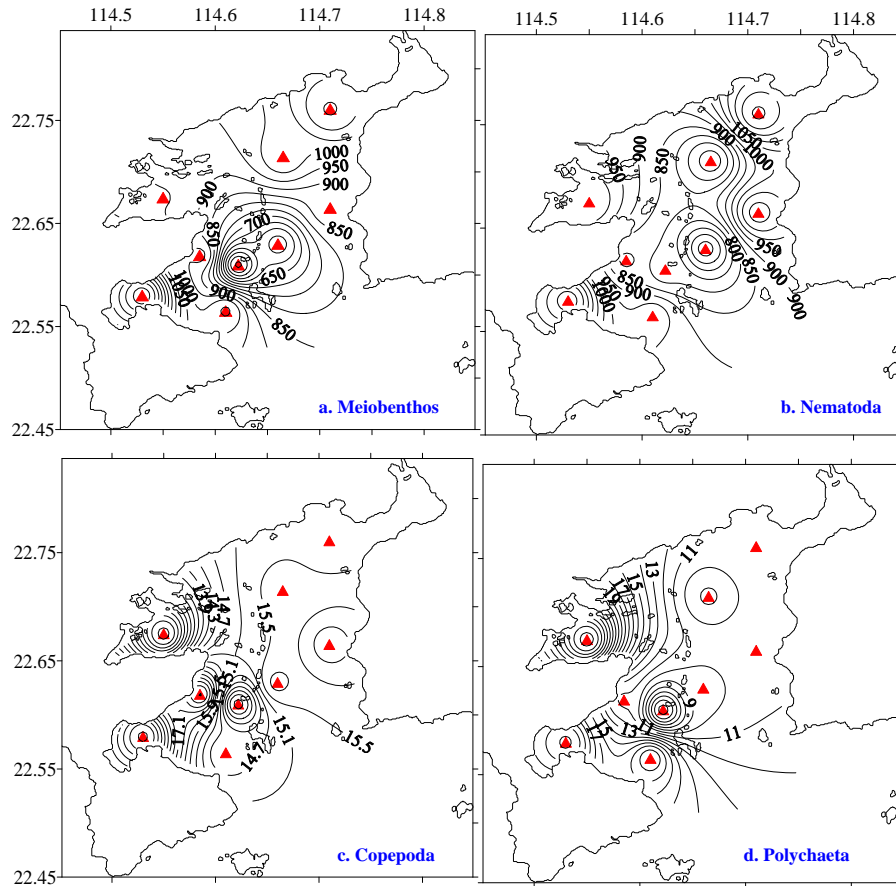
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**Fig. 6.** CLUSTER analysis of stations and seasons based on environmental parameters.

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**Fig. 7.** The horizontal distribution of meiofauna and the main groups in Daya Bay.

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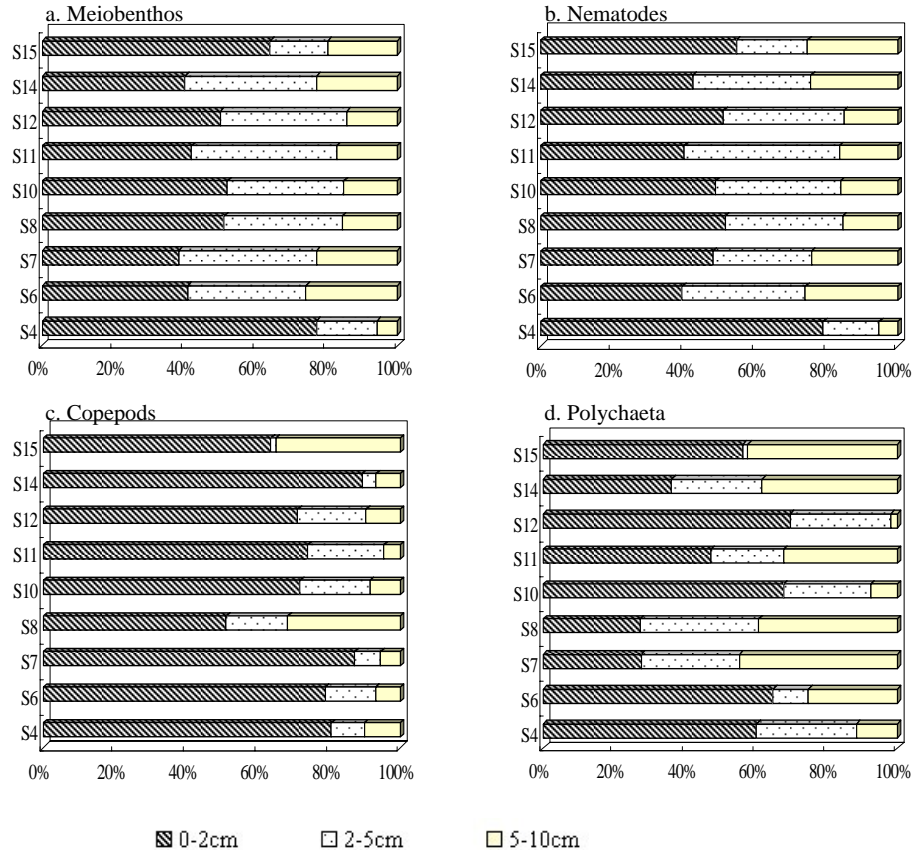
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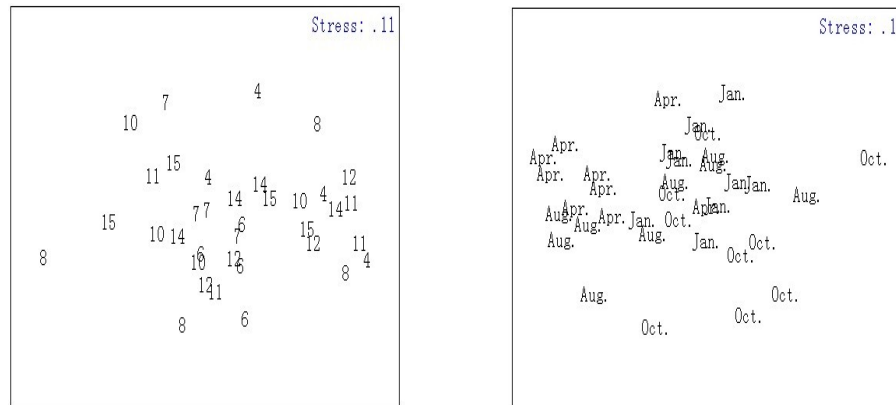
**Fig. 8.** The vertical distribution of **(a)** total meiofauna and its major taxonomical groups: **(b)** nematodes, **(c)** copepods, and **(d)** polychaetes.

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**Fig. 9.** Multidimensional scaling plots of meiobenthos assemblages based on fourth-root transformed abundance data from nine stations and four seasons. The plots are coded by stations and by seasons: Oct = autumn, Jan = winter, Apr = spring, and Aug = summer.

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