

Quantification of calcium phosphates in sediment

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Quantification of octacalcium phosphate, authigenic apatite and detrital apatite in coastal sediments using differential dissolution and standard addition

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Knowledge of calcium phosphate (Ca-P) solubility is crucial for understanding temporal and spatial variations of phosphorus (P) concentrations in water bodies and sedimentary reservoirs. In-situ relationships between liquid and solid-phase levels cannot be fully explained by dissolved analytes alone and need to be verified by determination of particular sediment P species. Lack of quantification methods for these species limits the knowledge of the P cycle. To address this issue, we (i) optimized a specifically developed conversion-extraction (CONVEX) method for P species quantification using standard additions; and (ii) simultaneously determined solubilities of Ca-P standards by measuring their pH-dependent contents in the sediment matrix. Ca-P minerals including various carbonate fluorapatite (CFAP) specimens from different localities, fluorapatite (FAP), fish bone apatite, synthetic hydroxylapatite (HAP) and octacalcium phosphate (OCP) were characterized by XRD, Raman, FTIR and elemental analysis. Sediment samples were incubated with and without these reference minerals and then sequentially extracted to quantify Ca-P species by their differential dissolution at pH values between 3 and 8. The quantification of solid-phase phosphates at varying pH revealed solubilities in the following order: OCP > HAP > CFAP (4.5 % CO₃) > CFAP (3.4 % CO₃) > CFAP (2.2 % CO₃) > FAP. Thus, CFAP was less soluble in sediment than HAP, and CFAP solubility increased with carbonate content. Unspiked sediment analyses together with standard addition analyses indicated consistent differential dissolution of natural sediment species vs. added reference species and therefore verified the applicability of the CONVEX method in separately determining the most prevalent Ca-P minerals. We found surprisingly high OCP contents in the analyzed coastal sediments which supports the hypothesis of apatite formation by an OCP precursor.

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1 Introduction

Phosphorus cycles are influenced by P concentration and composition in both the liquid and solid phase. Although non-equilibrium behaviour is typical for natural systems (Carman and Rahm, 1997), precipitation and dissolution mechanisms may occasionally attain equilibrium between certain P minerals and dissolved inorganic P (DIP). Provided DIP levels are controlled by these mechanisms, P minerals may be qualitatively determined in natural (Avnimelech, 1983; Bray et al., 1973; Emerson and Widmer, 1978; Gaudette and Lyons, 1980) or experimental (Murrmann and Peech, 1969; Oxmann et al., 2008) environments using appropriate dissociation constants and apparent solubility products allowing for ionic strength effects and ion-pairing. However, detections of particular mineral species using solubility criteria are rarely reported for natural systems mainly due to kinetic control of mineral reactions and/or adsorption-controlled DIP levels. For instance, P may be sorbed to Fe, Al and Mn oxides, iron oxyhydroxides, kaolinite and montmorillonite (Bar-Yosef et al., 1988; Chen et al., 1973; Hutchison and Hesterberg, 2004; Parfitt et al., 1975).

Whether adsorption effects or mineral formation and dissolution influence soluble P concentrations depends largely on sediment types and physicochemical conditions. For example, the redox state affects DIP by promoting sorption to a FeOOH-phosphate complex in oxic sediments and release from this complex under reducing conditions (Einsele, 1936; Mortimer, 1941). A decrease of the redox potential (Eh) may therefore release P (Mortimer, 1971) and could cause a shift from sorption to precipitation (e.g. Ruttenberg and Berner, 1993). However, soil incubation experiments by Murrmann and Peech (1969) indicated that pH also significantly affects adsorption-desorption and precipitation-dissolution mechanisms: whereas DIP was controlled by sorption below pH values of 8, DIP was in equilibrium with octacalcium phosphate (OCP) above pH 8. A similar pH-mediated change from adsorption to precipitation was confirmed for coastal sediments (Oxmann et al., 2008) and agrees with a reduced sorption capacity by (hydr)oxide minerals with increasing pH (Oh et al., 1999; Spiteri et al., 2008; Barrow

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et al., 1980; Naidu et al., 1990) and lower OCP solubility under progressively increasing alkaline conditions (Fig. 1; Lindsay et al., 1989). Hence, pH- and Eh-mediated effects are difficult to separate. Phosphorus release during reduction could also be caused by apatite solubilization (Golterman, 2001) because a change to anoxic conditions is often accompanied by pore water acidification (Jourabchi et al., 2005).

Due to decreasing apatite solubility with decreasing acidity (Fig. 1), calcium phosphate (Ca-P) contents of soils and sediments generally increase with pH (Lindsay et al., 1989; Oxmann et al., 2010; Tyler, 2002). Accordingly apatite frequently constitutes the main P fraction of marine sediments (e.g. Daesslé et al., 2004; Morse and Cook, 1978). Apatite is classified into biogenic and inorganic apatite. The formation of inorganic apatite by chemical precipitation and recrystallization processes has been subject of extensive research during past decades (e.g. Bendor, 1980; Sheldon, 1981; Tribble et al., 1995). Earlier research focused on the formation of carbonate fluorapatite (CFAP) bearing phosphorite deposits, which are referred to as such if they contain more than 5 wt% P₂O₅ (Ruttenberg and Berner, 1993). Phosphorites form mainly at continental shelves and have been therefore closely linked to upwelling phenomena (e.g. Kazakov, 1937; Parrish et al., 1986). Locally occurring saturation states that support a sustained CFAP precipitation were either explained by chemical mechanisms, such as steady P supply by nutrient-rich deep ocean waters as well as temperature increase, pressure decrease, pCO₂ decrease and pH increase of ascending waters (Atlas, 1975) or by biochemical processes including the high productivity at upwelling zones and related P enrichment of interstitial waters. Concentrations of DIP may further increase due to additional diagenetic transformations such as P fixation in oxic and subsequent release in anoxic sediment (Ruttenberg and Berner, 1993). Even more factors, partly independent of upwelling processes, have been proposed for the concentration of pore water phosphate, such as river sources (e.g. Pevear, 1966) and P release by sulfur bacteria (Crosby and Bailey, 2012; Schulz and Schulz, 2005). Hence, different P enrichment mechanisms may increase the saturation state enough to enable spontaneous Ca-P precipitation for subsequent CFAP formation. However, the P precipitation

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in itself is chemically driven by the saturation state and kinetic factors without direct cellular control (e.g. Krajewski et al., 1994).

Solubility of CFAP is affected by the extent of carbonate inclusion within the crystal lattice and increases with carbonate content (Chen et al., 1973; Jahnke, 1984; LeGeros and LeGeros, 1984). Due to its extensive distribution within common sediments or deposits, CFAP has been suggested to be an important oceanic sink of reactive P (Ruttenberg, 1992; Ruttenberg and Berner, 1993). While numerous studies suggested that seawater is largely CFAP-undersaturated despite the extensive CFAP distribution (Baturin, 1981; Bendor, 1980, and references therein), Atlas and Pytkowicz (1977) and Sheldon (1981) argued that seawater is close to CFAP saturation but noted the extremely slow kinetics of homo- or heteronucleation observed by incubation experiments. According to these kinetic considerations, Ca-P precipitation is apparently prevented in the water column (Atlas, 1975; Atlas and Pytkowicz, 1977). Moreover, direct nucleation of apatite was found to be inhibited by magnesium and, thus, spontaneous inorganic precipitation suggested to be kinetically impossible in marine environments (Golubev et al., 1999; Gunnars et al., 2004; Martens and Harriss, 1970). In contrast, more soluble potential precursor phases are likely less inhibited by magnesium (Gunnars et al., 2004).

Given the kinetic constraints of apatite nucleation under supersaturated conditions, more soluble Ca-P species may drive apatite formation according to the Ostwald step rule (Morse and Casey, 1988; Nancollas et al., 1989). Thus other Ca-P minerals, such as OCP and amorphous calcium phosphate (ACP), may represent significant sinks during early diagenesis and may influence soluble P levels. Incubation experiments with seawater indicated apatite formation by prior precipitation of metastable precursor phases, and a corresponding mechanism has been suggested for certain phosphorite deposits (Gunnars et al., 2004; Jahnke et al., 1983; Krajewski et al., 1994; Van Cappellen and Berner, 1988; Schenau et al., 2000). Apatite precursors are however difficult to identify, even in relatively simple mixtures without interfering sample

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matrices. Therefore, direct evidence for the sedimentary occurrence of OCP or ACP is scarce owing to methodological issues with respect to complex matrices.

Phosphorus analysis in sediment is generally performed by sequential extraction methods, which mainly separate the following P fractions: (i) exchangeable or loosely sorbed P; (ii) Al/Fe-bound P (Al/Fe-P); (iii) Ca-P; and (iv) organic P. Sequential extraction methods were developed for agricultural purposes (e.g. Hedley et al., 1982) and subsequently modified for sedimentary studies (e.g. SEDEX method, Ruttenberg, 1992). Phosphorus fractions were sub-divided in some schemes in order to yield additional pools, such as CFAP (Ruttenberg, 1992) and Al-P (Kuo, 1996). To support the interpretation of these widely used extraction methods a complementary speciation method was recently developed to quantify OCP and different apatites (CONVEX; Fig. 2; Oxmann et al., 2008). This method combines a conversion of P compounds by batch titration with subsequent sequential extraction and was proposed to be used together with conventional methods.

For validating the specificity of the CONVEX method, Oxmann et al. (2008) conducted standard addition experiments using OCP, hydroxylapatite (HAP), strengite and variscite. For Ca-P, addition experiments revealed that OCP can be well distinguished from apatites due to its significantly higher solubility (Oxmann et al., 2008). Using CONVEX, we found that OCP was the major Ca-P species in alkaline sediments (Oxmann et al., 2008). Added HAP dissolved at lower pH and a third distinguishable compound dissolved upon further acidification. The latter was not identified due to lack of available standards but suggested to be fluorapatite (FAP). Carbonate fluorapatite remained also unverified by addition experiments despite its relevance. Theoretically, CFAP could dissolve similar to OCP or HAP based on the apparent solubility product of Perrone et al. (2002; see Fig. 1). To demonstrate CONVEX method applicability, especially for purposes of marine Ca-P analyses, we provide supplementary standardization by addition experiments in this study and show appropriate selective dissolution.

Our specific objectives were (i) to validate the specificity of the CONVEX method for FAP and CFAP; and (ii) to verify consistent differential dissolution of natural

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of the sample was dried at 37 °C, ground with a planetary mill (< 300 μm mesh; PM₁₀₀; Retsch, Haan, Germany) and used for subsequent conversion-extraction analyses (see CONVEX method: Sect. 2.4). Basic P fractionation into Al/Fe-P and Ca-P was carried out according to Kurmies (1972) with minor modifications as described in Oxmann et al. (2008).

With an Eh of −98 mV and a pH of 7.467, the sediment was reduced and slightly alkaline and had clay, silt and sand fractions of 8, 34 and 58 %, respectively. Calcium phosphate bound P was the predominant fraction with a dry weight content of 5.51 μmolPg^{−1}, while the Al/Fe-P pool had an amount of 3.02 μmolPg^{−1}. Sediment moisture content was 58 % and salt concentration was 16.5 ‰. Results of CONVEX analysis of the sediment from the temperate tidal flat were compared with those of sediment from a tropical tidal flat (Oxmann et al., 2008) to test cross-regional transferability (see Sect. 3.1).

2.2 Preparation and identification of calcium phosphate reference minerals

In total, eleven Ca-P minerals were prepared for standard addition experiments. Their origin is given in Table 1. Specimens were acquired from mineral distributors, synthetic HAP was purchased from Sigma Aldrich (St. Louis, Missouri, USA) and OCP was synthesized as described in Christoffersen et al. (1990).

A large single FAP crystal was ground to powder and used without further preparation. All CFAP specimens with visually distinguishable mineral attachments had characteristics of either (i) numerous small needle-like or small hexagonal crystals attached to a matrix; or (ii) spheres of a botryoidal cluster embedded into a matrix (Table 1). To generate pure mineral samples, crystals were picked and spheres were isolated from the matrix under a binocular microscope. Received fragments were again inspected to avoid any significant contamination and subsequently ground. Cryptocrystalline collophane could not be purified and in this case the entire specimen was ground. Correspondingly, all bones of a fresh Australasian snapper (*Pagrus auratus*) were ground after they were cleaned from attached tissue with a knife and dried at 40 °C. Heating

at elevated temperatures, boiling in water or chemical clean-up was avoided to prevent changes of the crystalline structure or carbonate content. As colophane and bone specimens were not purified, their carbonate content was not determined. In contrast, care was taken during purification of other samples for precise quantification of their mineral carbonate content.

Mineral analyses were performed using x-ray diffraction (XRD; PW 1130 HV generator; automation-modified 1050/25 goniometer fitted with PW 1752 curved graphite crystal monochromator and PW 1965 proportional detector; Philips Analytical, Amsterdam, the Netherlands; Cu-tube, 40 kV, 20 mA), Raman spectroscopy (RM1000 confocal micro-Raman system; Renishaw, Gloucestershire, UK; laser excitation at 488 nm, 26 mW) and Fourier transform infrared spectroscopy (FTIR; Spectrum One FT-IR Spectrometer; Perkin Elmer, MA, USA; 4 cm^{-1} spectral resolution). Briefly, crystals and spheres were pure, highly crystalline and correctly identified. In contrast, snapper bones and colophane were found to be impure and less crystalline. Despite impurities, both samples had relatively high P contents. Snapper bone contained more than 50% of the P content of pure minerals and colophane approximately five-fold less than pure minerals as determined by photometric determination (see Sect. 2.4).

2.3 Carbonate content of Ca-P reference minerals

In order to determine mineral carbonate content, FTIR and elemental analysis were used. Calibration of FTIR by apatite samples with known carbonate contents is an accurate method for carbonate quantification, which uses the extinction ratio of the FTIR carbonate to phosphate band. For FTIR, 2 mg of sample was ground with 200 mg of potassium bromide and pressed in a mechanical press to form pellets for spectroscopy. Analysis was carried out as described by Featherstone et al. (1984) with two modifications: (i) carbonate contents of apatite reference samples were determined using an elemental analyzer (TruSpec CNS; LECO Corp., MI, USA) instead of preparing blended apatite standards with barium carbonate; and (ii) area ratios ($E_{1380-1440}/E_{550-590}$; Omnic Software Suite; Thermo Scientific, MA, USA) were used instead of extinction ratio at

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specific wave lengths. For extinction ratio calibration, data of five minerals with known carbonate content were used (cf. Table 1). Contents of other minerals listed in Table 1 were calculated using the correlation given in Fig. 2.

Results of elemental analysis correlated with those of FTIR at a significance level of 0.0001. Hexagonal crystals from the UK had lowest carbonate contents and spheres of botryoidal clusters from Germany and Australia had highest contents (Table 1). Carbonate contents of natural FAP, purchased HAP and synthesized OCP were comparable and much lower than those of natural CFAP (Table 1). To verify the effect of mineral carbonate content on sedimentary CFAP solubility by a lower number of addition experiments CFAP samples were partly pooled, excluding collophane. The upper six CFAP samples in Table 1 were pooled into three composite samples according to similar CO₃ inclusions of each pooled pair (4.4 and 4.7 %; 3.4 and 3.4 %; 2.1 and 2.3 %).

2.4 CONVEX method

The CONVEX method is shown in Fig. 3. For the conversion procedure (1A/1B), sediment subsamples of 1 g were incubated for seven days in 10 mL pH-adjusted 0.01 M CaCl₂ using two wash steps with 0.01 M CaCl₂ after two days. Subsequently, subsamples were centrifuged and pH was measured in the clear supernatant solution with the instrumental set-up described above but using an InLab Micro electrode (Mettler Toledo GmbH, Greifensee, Switzerland). Soluble P analyses of supernatant solutions were not performed because corresponding DIP concentrations were already shown (i) to alter with pH in a typical manner for different sediment and soil types; (ii) to vary inversely with total inorganic P; and (iii) to be insignificant with respect to solid-phase levels within a certain pH range despite large P transfers between Al/Fe-P and Ca-P (see Murrmann and Peech, 1969; Oxmann et al., 2008; and Sect. 3.5). For sequential extraction, dried sediment pellets were extracted by a series of steps (2A–3C) that enable an accurate separation of inorganic P into Al/Fe-P (mineral and adsorbed P) and Ca-P (mineral and residual P). Total inorganic P was calculated by summing Al/Fe-P and Ca-P. Sediment subsamples without prior incubation served as controls while

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differences between spiked and unspiked incubations yielded pH-dependent losses for each added standard mineral.

For addition experiments, minerals were weighed into small polyethylene caps using an ultra-micro balance (XP6U; Mettler Toledo GmbH, Greifensee, Switzerland) and then added to the sediment subsamples before incubation. The separate addition ensured homogeneity of added P among subsamples as opposed to a possible mineral addition to a larger sediment amount before subsample weighing. Care was taken to maintain experimental conditions and to minimize sample loss during centrifugation and decanting steps. For example, tubes were briefly centrifuged (3 min, 3000g) and shaken once before each centrifugation step (15 min, 3000g) in order to remove remaining particles from tube walls with clean supernatant solution. This procedure ensured minimal loss by subsequent decanting. Phosphate concentrations in solutions obtained by sequential extraction were determined according to Murphy and Riley (1962) using a UV-Vis spectrophotometer (Cintra 2020; GBC Scientific Equipment, Dandenong, Australia). A detailed description of the CONVEX method is given in Oxmann et al. (2008).

2.5 Data analysis

To quantify mineral loss for all standard addition experiments at equal pH values, data sets for Ca-P vs. pH were interpolated (0.001 pH intervals) using piece-wise cubic hermite interpolating polynomial (Octave, version 3.2.4; Eaton et al., 2008). Based on maximum changes in curve slopes ($\Delta c/\Delta \text{pH}$) for unspiked sediment (Sect. 3.2), differential dissolution of sediment FAP, CFAP and OCP occurred at pH ranges of ca. 3.0–3.5, 3.5–7.0 and 7.0–8.1, respectively. Selective losses of sediment species (sediment FAP, CFAP, OCP) and total species (added species plus corresponding sediment species) at different pH values were calculated according to:

$$\Delta c_{\text{loss}} = \Delta c_{\text{initial}} - \Delta c_{\text{actual}}, \quad (1)$$

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where $\Delta c_{\text{initial}}$ (initial concentration of each species) equals the difference between Ca-P concentrations at the beginning (upper cutoff pH values of 3.5, 7.0 and 8.1 for FAP, CFAP and OCP, respectively) and end (lower cutoff pH values of 3.0, 3.5 and 7.0 for FAP, CFAP and OCP, respectively) of each selective loss, and Δc_{actual} (actual concentration of each species after acidification-induced losses; actual pH < upper cutoff pH) equals the difference between Ca-P concentrations at actual and lower cutoff pH values (see example of calculation for OCP loss at pH 7.5 in Fig. 5a). For calculation of selective losses of added reference species (FAP, CFAP, OCP), $\Delta c_{\text{initial}}$ and Δc_{actual} were defined by the difference between Ca-P concentrations of spiked and unspiked incubations at upper cutoff pH values and actual pH values. The difference in Ca-P content ($\Delta c_{\text{initial}}$) between spiked and unspiked incubations at upper cutoff pH values of 3.5, 7.0 and 8.1 for FAP, CFAP and OCP (average of $2.04 \mu\text{mol P g}^{-1}$, relative standard deviation of 3.1%) closely matched the quantity of spiked P ($2 \mu\text{mol P g}^{-1}$) for each addition experiment. Hence, added FAP, CFAP and OCP were not significantly dissolved during incubations at pH values of 3.5, 7.0 and 8.1, respectively, and were quantitatively recovered by the Ca-P fraction at these pH values. Results from Eq. (1) were subsequently used to calculate loss percentages of sediment species, of total species and of added reference species for different pH values (Table 2). Based on results of standard addition experiments a slightly lower final pH (6.9) was preferable to calculate OCP contents (see Sect. 3.3).

3 Results and discussion

3.1 Sedimentary Ca-P speciation

For both unspiked sediments curve slopes changed at pH values of approximately 7, 3.5 and 3 (Fig. 4a and b) due to differential dissolution of Ca-P species, as proven by standard addition experiments (see Sect. 3.2). Octacalcium phosphate dissolved within a narrow pH range above pH 7, CFAP within a large pH interval of ca. 3.5–7

and FAP within a small pH range below pH 3.5. Due to these dissolution ranges and given species concentrations, incremental acidification-induced losses of OCP, CFAP and FAP per pH ($\Delta\text{OCP}, \text{CFAP}, \text{FAP} / \Delta\text{pH}$) resulted in a steep, a less steep and a steep curve, respectively (cf. unspiked sediment in Fig. 4a and b).

Species contents of unspiked sediments are indicated by columns at the right of Fig. 4a and b. Carbonate fluorapatite in the sample from New Zealand was elevated (43%) based on a lower amount of FAP (8%) and residual P (12%) if compared to the sample from Vietnam (CFAP: 25%; FAP: 18%; residual P: 20%). Thus, species analysis showed different P compositions despite similar sediment properties (reduced and slightly alkaline; similar Ca-P content) of the two contrasting sites. However, OCP contents of both sediments were equal and with a value of 37% surprisingly high. These findings generally agree with a kinetic control of species compositions because OCP precipitates in natural systems more rapidly (days to weeks; Bell and Black, 1970) than apatite (ten to some thousand years; Schenau et al., 2000; Jahnke et al., 1983; Gulbrandsen et al., 1984). Hence, more soluble species might better mirror ongoing processes whereas sparingly soluble species obviously reflect varying conditions over time due to slow precipitation. In accordance with such a kinetic control, OCP has been consequently proposed as a precursor for apatite formation (Gunnars et al., 2004; Krajewski et al., 1994).

The occurrence of OCP supports suggestions that apatite forms via an OCP precursor. The precursor hypothesis agrees with earlier reported kinetic barriers but tends to contrast with the often suggested strong undersaturation of seawater with respect to CFAP. If direct nucleation of apatite is kinetically inhibited in marine environments by magnesium (Golubev et al., 1999; Gunnars et al., 2004; Martens and Harriss, 1970), the ambient water must be supersaturated with respect to a precursor to facilitate apatite formation by a less magnesium-affected pathway. For example, in long-term incubation experiments apatite precipitated from natural brackish seawater only in case of supersaturation with respect to OCP (Gunnars et al., 2004). The liquid phase must be several times supersaturated with respect to apatite, though, in order to attain

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supersaturation with respect to OCP or even ACP. Conversely, it is difficult to explain the occurrence of a highly soluble Ca-P mineral with rapid dissolution kinetics, such as OCP, when ambient water is undersaturated even with respect to a much more insoluble Ca-P species like CFAP. However, an undersaturation of bottom water with respect to marine apatite is difficult to reconcile even with the occurrence of comparatively stable ancient CFAP because the latter did not dissolve despite exposure to undersaturated conditions for millions of years (Sheldon, 1981). Accordingly, Atlas (1975) inferred that prolonged apatite persistence requires at least saturation of sediment with respect to apatite. It is even more remarkable that recent studies suggest CFAP formation in sinking particulate matter throughout the water column (Faul et al., 2005; Lyons et al., 2011) using extractions of Ruttenberg's SEDEX method (1992). Although protective coatings and microenvironments may be considered for the prevented dissolution of ancient CFAP and potentially occurring precipitation in the water column, apparent undersaturation is frequently not confirmed by field observations (Sheldon, 1981).

However, a suggested precursor pathway obviously relies more on continuously high pH values and elevated DIP and calcium concentrations compared to direct CFAP nucleation. For example, Ca-P formation via OCP precipitation was detected after approximately two years of incubation in calcite-seeded brackish seawater using a DIP concentration of 500 μM , which is slightly higher than the range reported for interstitial water of Baltic Sea sediments (Gunnars et al., 2004). In addition, relatively low dissolved magnesium levels are likely necessary, because OCP growth seems to be less but not completely uninfluenced by magnesium (e.g. Bell and Black, 1970). Yet, equilibrium calculations by means of pore water data imply that most interstitial waters of the Baltic Proper are supersaturated for long periods, at least with respect to apatite (Carman and Rahm, 1997), and thus agree with observed P transformations into apatite (van Beusekom and De Jonge, 1997).

In view of the proposed but not unambiguous OCP precursor pathway, Gunnars et al. (2004) highlighted the difficult in situ identification of authigenic apatite. However, OCP detection may pose an even larger problem because OCP is not quantifiable in

sediment using any conventional method. Furthermore, OCP is not included in operationally defined fractions of common sequential extraction methods. Calcium phosphate fractions of these methods are consequently incompletely defined and those definitions may result in overestimates of other fractions and the exclusion of OCP. Possible misinterpretations might be not insubstantial in view of OCP contents detected in the present study. Overall, the proposed method offers a good opportunity to further evaluate the issues mentioned above by field or experimental studies. Initial results support suggestions of an OCP precursor during apatite formation for the sediments investigated. However, further analyses of samples from other regions are required to determine spatial OCP distributions; to clarify whether successive or direct nucleation is favoured in environmental systems; and to identify factors that may facilitate or inhibit one or the other pathway at different locations.

3.2 Solubility of calcium phosphates in sediment: verification of differential dissolution

Results of Ca-P addition experiments (Figs. 4–6) show mineral species successively dissolved by acidification according to their theoretical solubility, agree with thermodynamic considerations and verify the applicability of the CONVEX method. Moreover, solubility of added CFAP increased with its carbonate content, which is in line with previous studies (Chien and Black, 1976; Jahnke, 1984; LeGeros and LeGeros, 1984).

The OCP and HAP addition experiments showed that the CONVEX method can distinguish OCP from HAP in sediment. Almost complete dissolution of sedimentary and added OCP at pH 7 (cf. ST-OCP in Fig. 4a and b) verified OCP quantification by the Ca-P loss at this pH because added HAP was not significantly dissolved at or above pH 7 (cf. ST-HAP in Fig. 4a and b). Addition experiments with HAP showed Ca-P loss above pH 7 but this loss was caused by the sedimentary OCP content. The detected OCP presence only at pH values above ca. 7 is in line with soil column incubations by Bell and Black (1970), which confirmed OCP formation only in the alkaline range. Precipitation of OCP in the alkaline range also agrees with DIP concentrations in supernatant

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solutions of pH batch experiments of soils and sediments because soluble P followed the OCP isotherm (Murrmann and Peech, 1969; Oxmann et al., 2008).

The CFAP addition experiments verified that the CONVEX method can also distinguish CFAP from OCP (differential dissolution of CFAP and OCP in Fig. 4a) despite the apparently higher CFAP solubility in comparison to HAP (cf. Figure 1; apparent solubility product of CFAP). Further, all incubations with various CFAP minerals revealed a reduced loss compared to HAP spiked incubations within the pH range 3.5–7. Therefore, natural CFAP minerals with differing carbonate content were even more stable in the sedimentary environment than synthetic HAP and consequently did not bias determined OCP contents. Lower solubility of CFAP in comparison to HAP is in agreement with reported HAP conversion into more stable CFAP or increased HAP dissolution in the water column and near-surface sediments (e.g. Burdige, 2006; Schenau and De Lange, 2000). In contrast to the separation of CFAP or HAP from OCP, quantitative discrimination between HAP and CFAP is not possible without further methodological optimization because both apatites dissolved within the same pH range. Yet, the presence of significant quantities of HAP can be verified because approximately half of added HAP dissolved in the pH range 6.3–7 (cf. ST-HAP in Fig. 4a and b) while CFAP did not. Thus, incubations with HAP additions show a steeper curve within that interval compared to unspiked sediment incubations (illustrated by curved and straight arrows in Fig. 4b, respectively) and all FAP and CFAP additions shown in Fig. 4a (cf. DU-FAP, CW-CFAP, KC-CFAP, MS-CFAP). Loss of sediment Ca-P within the range of ca. 3.5–7 was previously ascribed to HAP based on concurrent HAP dissolution during corresponding addition experiments (Oxmann et al., 2008). However, as unspiked incubations and CFAP additions did not show an increased curve slope in the pH range 6.3–7, contrary to HAP additions, sedimentary Ca-P loss in the pH range 3.5–7 was likely caused by a predominance of sediment CFAP. Yet presence of some HAP in the unspiked sediments could not be ruled out.

Standard addition with igneous FAP from Mexico (cf. DU-FAP in Fig. 4a) demonstrated that Ca-P dissolution below pH 3.5 is caused by FAP and therefore showed that

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the CONVEX method is able to distinguish FAP from other Ca-P phases. All other apatites almost completely dissolved at ca. pH 3.5, whereas added FAP dissolved below this value. The observed significantly lower FAP solubility is consistent with the pronounced differences between solubilities of laboratory synthesized FAP and CFAP detected by Jahnke (1984). His and other investigations (Chien and Black, 1976; LeGeros and LeGeros, 1984) did not only point to solubility differences between pure FAP and CFAP but also showed that increasing carbonate content strongly increases the solubility of CFAP minerals with different degrees of carbonate substitutions. Regarding the effect of crystal carbonate content on CFAP solubility, increased carbonate substitution also caused a greater extent of CFAP dissolution in the present study (cf. CW-CFAP, KC-CFAP, MS-CFAP in Fig. 4a; Table 2).

Due to this dependency on carbonate substitutions and naturally occurring carbonate traces in FAP (Table 1), separation of sedimentary FAP from CFAP could have been insufficient. Presence of both, FAP with low carbonate content and CFAP with varying extent of carbonate substitution, could have resulted in a significant concurrent dissolution. However, there is a pronounced slope change for unspiked sediment at ca. pH 3.5, and the slope height in the pH range 3–3.5 obviously discriminates a certain amount of less soluble apatites from more soluble apatites. The ratio of slope heights is not constant for both sediments indicating that this is not an acidification-caused constant feature similarly related to all sedimentary apatites. Sediment from Vietnam contained less CFAP but more FAP than sediment from New Zealand. Finally, the slope change for unspiked sediment agrees with FAP and CFAP addition experiments. In conclusion, there must be certain mineral features that permit discrimination of these sedimentary apatites in the sedimentary environment in addition to the clear separation of added reference species.

However, the different mineral features that permit discrimination of FAP and CFAP are attributable to the different origins of sedimentary apatites. In contrast to authigenic CFAP, FAP is in general derived from detrital sources and delivered to marine environments by continental weathering. The pH at which differential dissolution of FAP and

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CFAP occurred during our addition experiments agrees approximately with a pH of 4, at which best separation of FAP and CFAP is attained in acetate buffer solutions or dilute mineral acids (Ruttenberg, 1992). Therefore, the CONVEX method distinguishes detrital from authigenic apatite within the sediment matrix. Whether this discrimination and hence the large solubility differences between sediment FAP and CFAP were primarily attributable to different carbonate contents remained unclear. Addition experiments showed that a rather small variation in carbonate content between DU-FAP and CW-CFAP (1.0 and 2.2 %, respectively) coincided with large solubility differences, whereas a larger variation between CW-CFAP and MS-CFAP (2.2 and 4.5 %, respectively) corresponded to less pronounced solubility differences (cf. Table 2). As detrital FAP and authigenic CFAP can contain various impurities, which are dependent on depositional and postdepositional conditions (Guidry and MacKenzie, 2003), other factors may also contribute to their strongly differing dissolution characteristics.

3.3 Congruency of species dissolution and possible sources of error

Based on maximum changes in curve slopes ($\Delta c/\Delta pH$) for unspiked sediment, complete dissolution of sediment OCP, CFAP and FAP was assumed to take place at pH values of 7.0, 3.5 and 3.0, respectively (Table 2). More than 80 % of added OCP dissolved at pH 7 (value in bold in Table 2; Fig. 5a for an example of calculation). However, CFAP only sparingly dissolved at slightly lower pH values (see loss percentages of CFAP at pH > 6.5) thereby minimizing a concurrent dissolution of CFAP and OCP. Thus, a slightly lower pH of 6.9 was preferable to calculate OCP contents (Fig. 5a). At this pH, the total OCP loss was approximately 100 %, loss of added OCP was 90 %, CFAP loss was lower than 5 % and sediment OCP might be overestimated by 5 % (Table 2).

In general more than 90 % of added CFAP and FAP dissolved at pH 3.5 and 3.0, respectively. Largest differences in CFAP dissolution due to varying carbonate inclusion were observed in the pH range 4 to 4.5 (Table 2, numbers in italic). Further, the dissolution of the CFAP sample with the highest degree of carbonate substitution (MS-CFAP:

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4.6%) was best correlated with the dissolution of sediment CFAP ($R^2 = 0.92$ for the values shown in Table 2). This correlation was progressively less pronounced with decreasing carbonate substitution of the other CFAP samples (KC-CFAP: 3.4%, $R^2 = 0.85$; CW-CFAP: 2.2%, $R^2 = 0.73$) and may therefore indicate a relatively high degree of carbonate substitution of authigenic apatite of the unspiked sediment sample. However, despite the effect of carbonate substitution on CFAP dissolution, different degrees of carbonate substitution did not significantly change the dissolution range of CFAP (Fig. 4a). All in all, dissolution ranges of added FAP, CFAP and OCP were almost congruent with the three dissolution ranges of sediment Ca-P (Table 2) verifying that standards were well-suited to explain these dissolution ranges.

Regarding potential transformations, incubations at in situ pH did not significantly change P compositions. For instance, OCP was not generated by desorption of adsorbed P because Ca-P and Al/Fe-P contents of controls without incubation (cf. Sect. 2.4.; Fig. 3) closely matched those of sediment incubated at in situ pH (Figs. 4 and 7). Hence, OCP did not form by P desorption (or any other non Ca-P source) because total Ca-P did not increase by incubation. Furthermore, one Ca-P species did not transform to another. If anything, one would expect extremely slow HAP formation by OCP hydrolysis (e.g. Gunnars et al., 2004) and certainly not OCP formation by apatite dissolution. None of this took place as shown by standard additions. Finally, controls without incubation were not biased by OCP formation during sequential extraction because the employed method was developed and validated to eliminate OCP precipitation (Kurmies, 1972; removal of Ca using three wash steps with KCl/EtOH prior to the alkaline extraction; Fig. 3).

However, control values fit even better to Ca-P concentrations of incubations at higher pH (see Ca-P contents closer to pH 8 in Fig. 4a and b) and were used for calculation of the most soluble species (OCP). Incubations with a final pH similar to field pH showed slight Ca-P dissolution because the adjusted pH was initially somewhat lower than the final pH and drifted subsequently upwards. Therefore, the final pH did not precisely reflect the pH of observed loss. However, this shift is not significant

because added Ca-P minerals (i) were subjected to the same conditions as sediment Ca-P; (ii) explained characteristic Ca-P dissolution ranges of unspiked incubations; and (iii) were measurable in the sediment matrix by differential dissolution.

3.4 Solubility of collophane and fish bone apatite in sediment

Collophane and fish bone specimens were also tested by standard addition experiments because various substitutions and crystal imperfections may result in increased solubility of collophane and bone apatite. Although biogenic apatite is likely an insignificant P sink on a global scale, it can constitute a considerable P fraction in some sediments. If so, biogenic apatite might bias the quantification of precursor phases due to its relatively high solubility compared to other apatites (see Schenau et al., 2000; Schenau and De Lange, 2000).

Dissolution of nanocrystalline snapper bone apatite (SB-AP) was comparable to that of crystalline CFAP with lowest carbonate inclusion (CW-CFAP) at higher pH but matched synthetic HAP (ST-HAP) dissolution at ca. pH 4 (Fig. 6). Bone apatite is structurally similar and frequently termed as HAP but also shows considerable chemical and structural differences (Wopenka and Pasteris, 2005). This may explain why the dissolution behaviour of the bone apatite was comparable to both, crystalline CFAP and synthetic HAP. Bone apatite contains typically ca. 7% carbonate, little or a complete lack of hydroxyl substitutions and other ions may be incorporated to adjust for required properties. For example, incorporation of fluorine into apatite reduces its solubility in organisms. However, bone apatite did neither bias the determination of OCP nor the detection of detrital apatite due to absent additional Ca-P loss above pH 6.9 (loss only caused by sediment OCP) and pronounced Ca-P loss at ca. pH 4. Likewise, OCP presence in the bone sample can be ruled out by the missing additional loss in the alkaline pH range. An OCP precursor during bone apatite formation has been proposed in the past but a recent publication concluded that OCP has not been detected hitherto in bone (Rey et al., 2009).

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Cryptocrystalline collophane (BL-CFAP) seemed to include an apatite mixture, as it dissolved in agreement with HAP, CFAP and FAP at high, medium and low pH, respectively. Thus, incubations with collophane only showed increased dissolution comparable to HAP (increased dissolution marked by curved arrow in Fig. 6) within the pH range 6.3–7 among all incubation experiments. This is in agreement with expected varying collophane compositions including presence of HAP. Thus, collophane incubations indicate that significant contents of HAP are likely detectable. This conclusion additionally suggests minor contents of HAP within the unspiked sediments because they did not show increased dissolution within the pH range 6.3–7 (straight arrow for unspiked sediment in Fig. 6).

3.5 Increase of Al/Fe-P due to calcium phosphate dissolution

The pH-dependent Ca-P variations were generally reflected by Al/Fe-P contents as P liberated by Ca-P dissolution was subsequently adsorbed and determined by the Al/Fe-P fraction (Fig. 7). Thus, Al/Fe-P mirrored Ca-P in the pH range 4–8 for each individual addition experiment with an accompanied sequence inversion among all addition experiments (cf. Figs. 7 and 4a). In consequence of the almost quantitative adsorption ($\Delta\text{Al/Fe-P} \approx \Delta\text{Ca-P}$), total inorganic P contents remained largely unchanged in the pH range 4–8 as shown in Fig. 5b for unspiked sediment and sediment with FAP, CFAP and OCP additions. These findings are in agreement with previous observations (Oxmann et al., 2008) and demonstrate that most pH-induced sedimentary P dissolution processes cannot be observed by total inorganic P variations or reverse soluble P levels because of almost quantitative solid-phase P transfers. Data of Al/Fe-P and Ca-P for unspiked sediment were also mirrored in the pH range 4–8. As they always involved lowest concentrations relative to other incubations (see Fig. 5b) they naturally did not follow sequence inversion.

Below ca. pH 4, less P was adsorbed despite large Ca-P losses (Fig. 7). The accompanied overall loss of sediment inorganic P below pH 4 (Fig. 5b) is in turn reflected by increased soluble P (see Oxmann et al., 2008). Very little free orthophosphate

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generated by Ca-P dissolution was adsorbed below pH 3 resulting in an almost complete liberation of added P from the sedimentary environment. Thus, even the relatively low original sedimentary Al/Fe-P content ($3.02 \mu\text{mol P g}^{-1}$) increased just marginally ($3.31 \mu\text{mol P g}^{-1}$) due to adsorption in the strongly acidic range (Fig. 7).

4 Conclusions

We found that OCP dissolved above ca. pH 7 and its quantification was not biased by FAP, CFAP, HAP or biogenic apatites. Thus, the CONVEX method is able to differentiate OCP from the apatite group. Carbonate fluorapatite dissolved together with HAP in the pH range 3.5–7. However, HAP dissolved to a greater extent at equal pH and was therefore more soluble than CFAP. Below pH 3.5, FAP began to dissolve and was therefore distinguishable from other apatites, verifying that the CONVEX method also distinguishes detrital from authigenic apatites. Solubility of CFAP increased with carbonate content, particularly in the pH range 4–4.5. Bone apatite and collophane dissolved in a similar manner to different identified reference apatites.

Overall, this study demonstrated that the CONVEX method can be used for experimental evaluation of P mineral solubilities in the sedimentary environment. Moreover, the method performs accurately for quantification of the most important sedimentary Ca-P species. Obtained results are not biased by common matrix interferences as the method of standard addition eliminates matrix effects.

The two coastal sediment samples differed in their FAP and CFAP content despite similar physicochemical properties. However, contribution of OCP to total Ca-P was equal for both sediments and, with a value of 37%, surprisingly high. The detected OCP occurrence supports suggestions of apatite formation by an OCP precursor thereby pointing to the importance of addressing corresponding compositions of species and their geographic distributions in future studies.

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Table 1. Basic properties, performed analyses and carbonate content (% by weight) of Ca-P reference minerals used for standard addition experiments.

ID-Mineral ^a	Origin	Type/Appearance	Characterization	Purity ^b	CO ₃
MS-CFAP	Moculta Phosphate Quarry, Angaston, Australia	Var. Staffelite, botryoidal, yellow-brown	XRD, FTIR, Raman, EA	PS	4.4
MS-CFAP	Staffel, Limburg, Germany	Var. Staffelite, botryoidal, brown	XRD, FTIR, Raman, EA	PS	4.7
KC-CFAP	Zheleznyi Mine, Kola Peninsula, Russia	Var. Staffelite, botryoidal, blue-green	XRD, FTIR, Raman, EA	PS	3.4
KC-CFAP	Holmbush Mine, Callington, Cornwall, UK	Var. Holmbushite, needle-like crystals	XRD, FTIR, Raman	PC	3.4
CW-CFAP	Geevor Mine, Pendeen, Cornwall, UK	Small hexagonal crystals, brown	XRD, FTIR, Raman	PC	2.3
CW-CFAP	Levant Mine, Pendeen, Cornwall, UK	Small hexagonal crystals, brown	XRD, FTIR, Raman	PC	2.1
BL-CFAP	Bonny Lake, Florida, USA (with HAP and FAP)	Var. Collophane, cryptocrystalline	XRD, FTIR, Raman	BS	ND
DU-FAP	Cerro de Mecado Mine, Durango, Mexico	Single large hexagonal crystal, green	XRD, FTIR, Raman, EA	PC	1.0
SB-AP	<i>Pagrus auratus</i> bones, New Zealand	Nanocrystalline apatite inclusion	XRD, FTIR, Raman	BS	ND
ST-HAP	Synthetic	Crystalline powder	XRD, FTIR, Raman, EA	S	0.9
ST-OCP	Synthetic	Crystalline powder	XRD, FTIR, Raman	S	0.8

^a The upper six CFAP samples were pooled into three composite samples according to their similar CO₃ content (see minerals with equal ID; MS-CFAP: 4.5%; KC-CFAP: 3.4%; CW-CFAP: 2.2% pooled CO₃ content).

^b Samples were prepared from pure spheres (PS), pure crystals (PC) or synthetic origin (S) and confirmed by XRD, FTIR and Raman or from bulk samples (BS) including other components.

Var.: Variety. EA: Elemental analysis. ND: Not determined.

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Table 2. Loss (%) of added reference species (FAP, CFAP, OCP), total species (added species plus corresponding sedimentary species) and sedimentary species (sediment FAP, CFAP, OCP) at varying pH as determined by the CONVEX method. See Table 1 for abbreviations of added standard minerals.

Final pH	DU-FAP	Total FAP	Sediment FAP	MS-CFAP	KC-CFAP	CW-CFAP	Total CFAP ^a	Sediment CFAP	ST-OCP	Total OCP	Sediment OCP
3.0	98	99	100								
3.2	40	48	76								
3.5	0	0	0	94	91	84	96	100			
4.0				79	55	40	70	82			
4.5				47	32	21	53	70			
5.0				31	23	18	43	58			
5.5				18	19	16	32	43			
6.0				8	10	15	20	27			
6.5				2	6	13	10	14			
6.8				1	4	8	6	7			
6.9				0	3	5	4	4	90	98	105 ^b
7.0				0	0	0	0	0	81	91	100
7.1									68	78	88
7.5									35	28	20
8.0									6	3	0
8.1									0	0	0

Values in bold indicate cutoff pH values for distinguishing Ca-P species (FAP: 3.0–3.5; CFAP: 3.5–7.0; OCP: 7.0–8.1). Italics: largest differences in CFAP dissolution due to different CO₃ substitution in the pH range 4–4.5 (MS-CFAP: 4.6%; KC-CFAP: 3.4%; CW-CFAP: 2.2% CO₃).

^a Among the different CFAP phases, KC-CFAP was used to calculate the total CFAP loss.

^b Indicates a possible overestimate if pH 6.9 is used for distinguishing between OCP and CFAP.

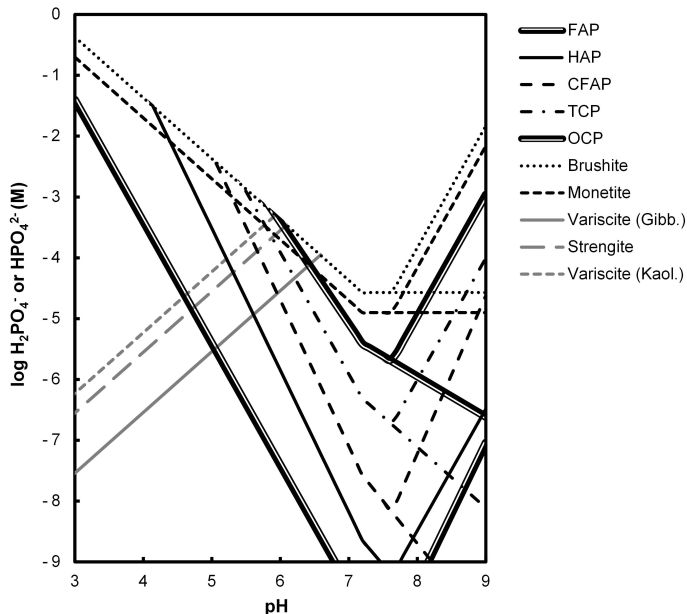


Fig. 1. Solubility of calcium phosphates, strengite and variscite (in equilibrium with gibbsite or kaolinite) when Ca^{2+} is 10^{-2} M or is fixed by calcite. $\text{CO}_2(\text{g})$: 0.0003 atm, F^- : 10^{-4} M (equilibrium level of CaF_2), CO_3^{2-} at the given $\text{CO}_2(\text{g})$. Strengite and variscite are more insoluble under acidic conditions while calcium phosphate solubility decreases with pH increase. At $\text{pH} > 7.88$ calcium phosphate solubility may increase due to Ca^{2+} depression caused by decreasing calcite solubility (see dividing curves at this pH for Ca^{2+} depression or maintained Ca^{2+} concentration). Adapted using calculation recommendations of Lindsay et al. (1989). Carbonate fluorapatite (CFAP) was introduced using the apparent $K_{\text{sp}} 10^{-103}$ given by Perrone et al. (2002). FAP: Fluorapatite; HAP: Hydroxylapatite; TCP: Tricalcium phosphate; OCP: Octacalcium phosphate.

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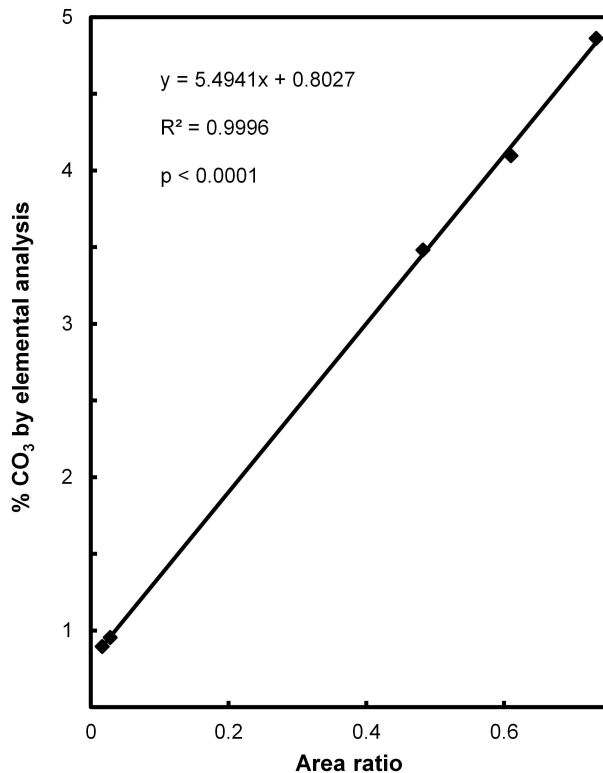


Fig. 2. FTIR calibration for carbonate content determination of calcium phosphate minerals. Ratio of the extinction of the FTIR carbonate band to phosphate band (areas, $E_{1380-1440}/E_{550-590}$) vs. carbonate content (% by weight) determined by elemental analysis. For calibration, data of five minerals with known CO₃ content were used (cf. Table 1, minerals with performed elemental analysis). Contents of other minerals listed in Table 1 were calculated using the indicated correlation (method after Featherstone et al., 1984 with modification).

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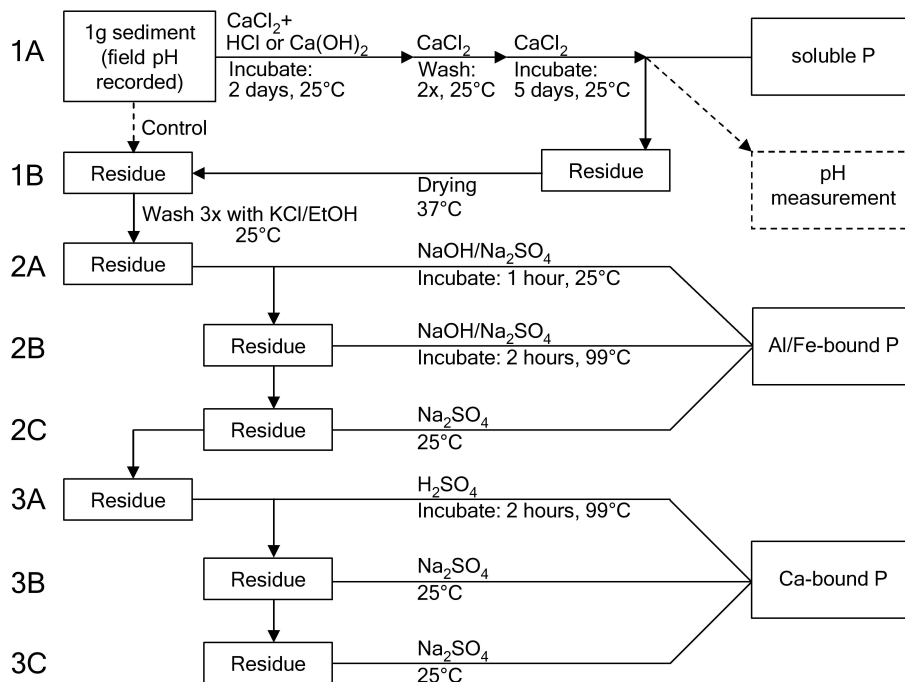


Fig. 3. CONVEX scheme for the quantification of phosphorus species in sediment. 1A/1B: Conversion procedure, 2A–3C: extraction procedure. For addition experiments, reference minerals were added to the sediment subsamples before incubation. Adapted from Oxmann et al. (2008).

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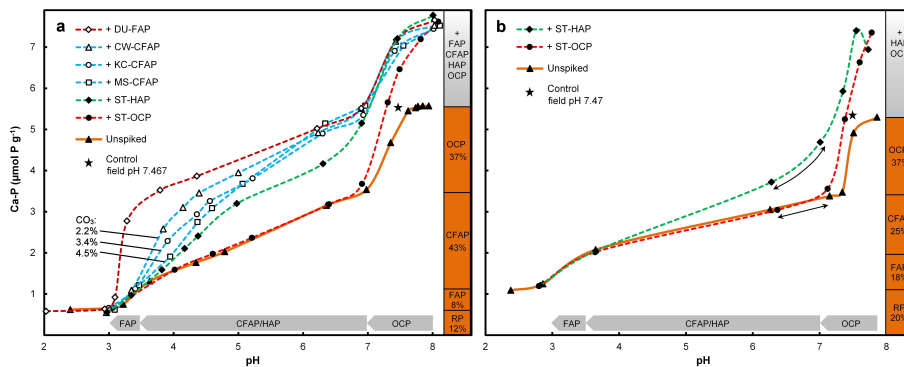


Fig. 4. Differential Ca-P dissolution in sediment at varying pH as determined by subsample incubation of sediment (unspiked and with reference mineral additions) and subsequent fractionation (CONVEX method). **(a)** Standard addition experiments with natural and synthetic Ca-P using sediment from the Firth of Thames, New Zealand. **(b)** Standard addition experiments with synthetic Ca-P using sediment from the Saigon River Delta, Vietnam (adapted from Oxmann et al., 2008). Columns indicate sedimentary Ca-P species composition in % (CFAP fraction may contain some HAP; see text) and standard additions. Note the greater extent of CFAP dissolution with increasing carbonate substitution. Note the increased slope of HAP addition for both sediments in the pH range 6.3–6.9, which was absent for unspiked sediment (arrows for visualization in **b**) and other additions (cf. Fig. 4a) except collophane (cf. Fig. 6). Control: fractionation without prior incubation. RP: Residual P. See Table 1 for abbreviations and origins of added standard materials.

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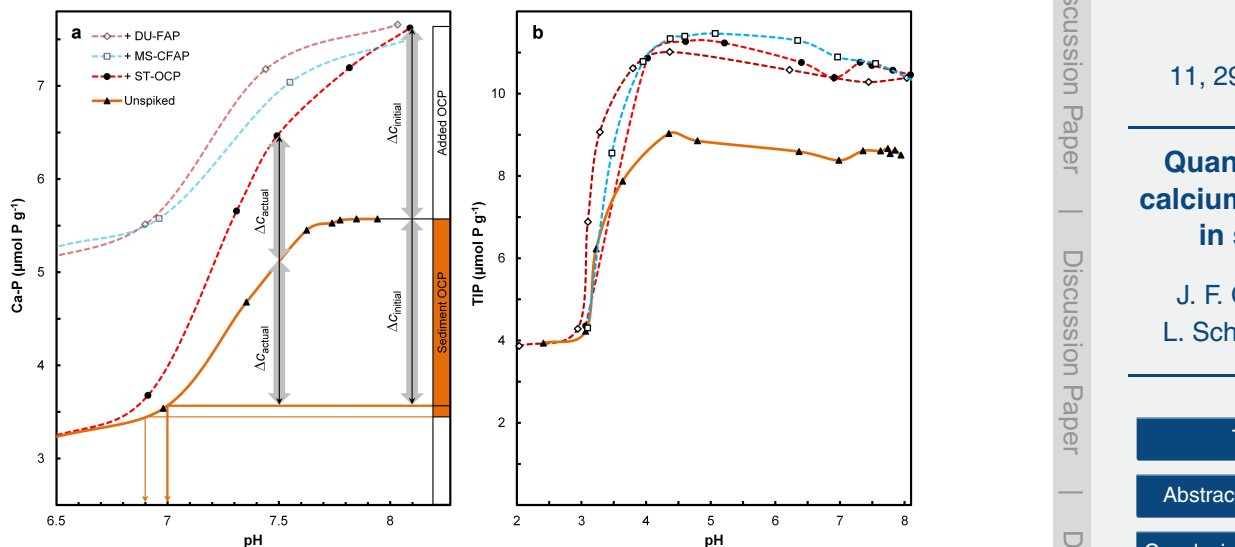


Fig. 5. (a) Calculation of Ca-P species dissolution by acidification-induced losses of the Ca-P fraction as indicated for OCP. Differential OCP dissolution occurred at ca. pH 7 (thick orange arrow) and a slightly lower pH (6.9; thin orange arrow) was preferable to calculate the OCP content (see text). Curves were fitted by piece-wise cubic hermite interpolating polynomial to calculate selective losses of species ($\Delta C_{\text{loss}} = \Delta C_{\text{initial}} - \Delta C_{\text{actual}}$) at different pH values. Grey arrows denote initial and actual concentrations (exemplified for pH 7.5) of added OCP (+ ST-OCP) and sediment OCP (unspiked). Superimposed black arrows denote corresponding concentrations of total OCP (added OCP plus sediment OCP). Calculated loss percentages for different P species at different pH values are given in Table 2. Standard addition of FAP and CFAP is shown for comparison (+DU-FAP; +MS-CFAP; insignificant dissolution in the range of OCP dissolution). **(b)** Acidification-induced changes of total inorganic P (TIP = Ca-P + Al/Fe-P) for unspiked sediment and standard additions of OCP, FAP and CFAP (same symbols as in Fig. 5a).

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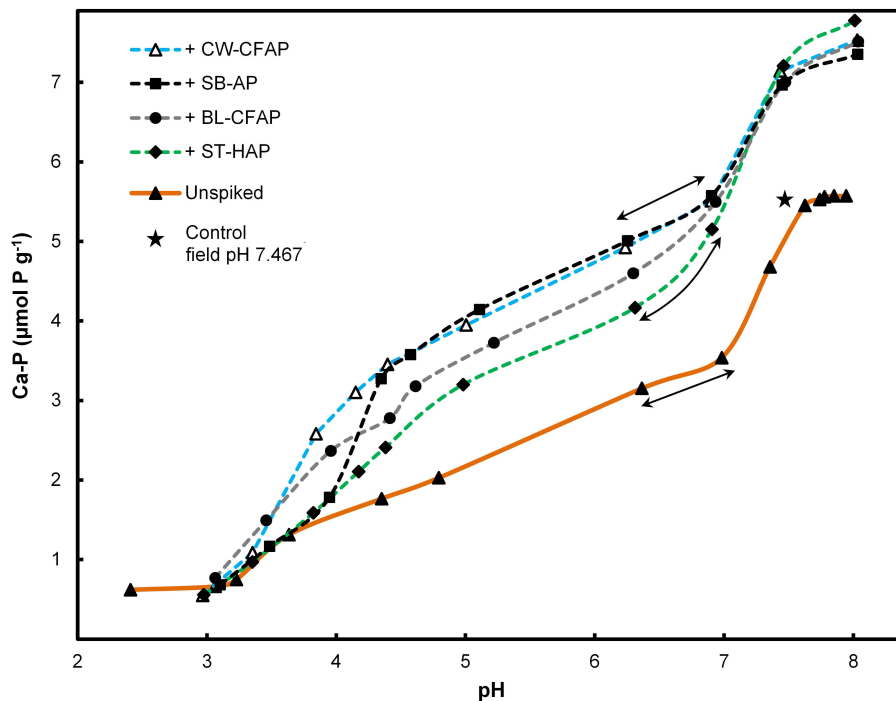


Fig. 6. Differential Ca-P dissolution in sediment at varying pH as determined by subsample incubation of sediment (unspiked and with reference mineral additions) and subsequent fractionation (CONVEX method). Cryptocrystalline colophane (BL-CFAP) dissolved similar to HAP (ST-HAP) in the range 6.3–6.9 (see increased slope, arrows). Analysis was performed using sediment from the Firth of Thames, New Zealand. Control: fractionation without prior incubation. See Table 1 for origins of added standard materials.

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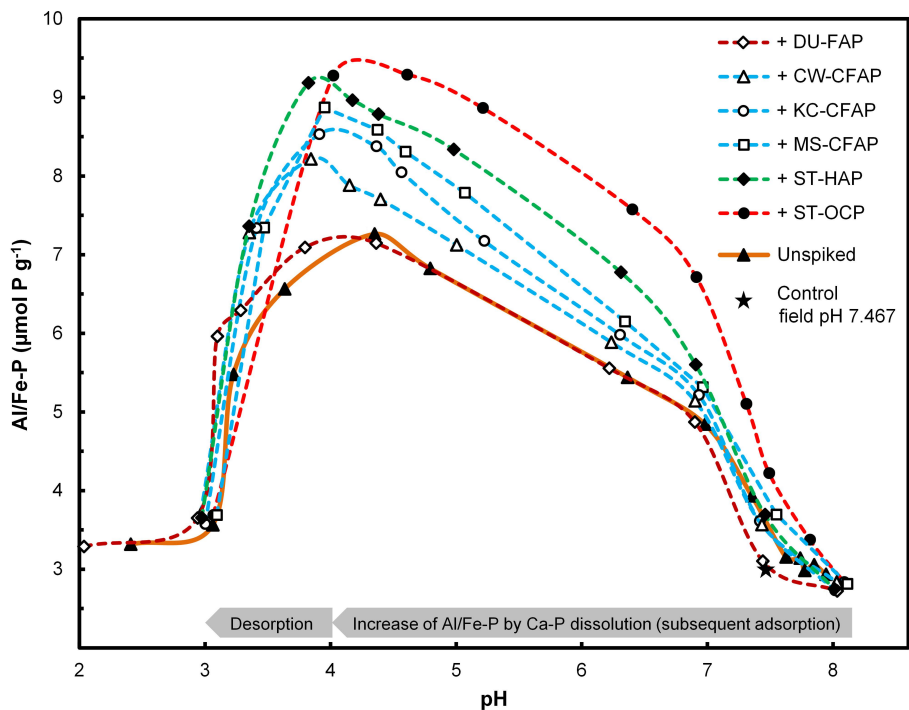


Fig. 7. Increase of Al/Fe-P due to calcium phosphate dissolution (subsequent adsorption; pH 4–8) and decrease of Al/Fe-P due to desorption (pH < 4) as determined by subsample incubation of sediment (unspiked and with reference mineral additions) and subsequent fractionation (CONVEX method). Note the mirror-like reflection in the pH range 4–8 when compared to Fig. 2a (except for unspiked sediment; see text). Analysis was performed using sediment from the Firth of Thames, New Zealand. Control: fractionation without prior incubation. See Table 1 for origins of added standard materials.

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