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# Spring-time zooplankton size structure over the continental shelf of the Bay of Biscay

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## Abstract

Linking lower to higher trophic levels requires a special focus on the pivotal role played by mid-trophic levels, i.e. the zooplankton. One of the most relevant information on zooplankton in term of fluxes of matter lies in its size structure. We present here an extensive dataset of size measurements covering part of the western European shelf and slope, from the Galician coast to the Ushant front, during springs from 2005 to 2012. Zooplankton size spectra were estimated using both measurements carried out in situ by the Laser-Optical Plankton Counter (LOPC, 816 records) and WP2 net (200 µm mesh size) samples scanned following the ZooScan methodology and image analysis (a total of 89 samples were analyzed). The LOPC counts and sizes all particles in the range 100 to 2000 µm of spherical equivalent diameter (ESD), whereas the WP2/ZooScan allows the counting, sizing and identification of zooplankton from ~ 400 µm ESD. The difference between the LOPC (all particles) and the WP2/ZooScan (zooplankton only) is assumed to provide the size distribution of non-living particles whose descriptors are further related to a set of explanatory variables (including physical, biological and geographic descriptors). A statistical correction based on these explanatory variables is then applied to LOPC measurements to removed the part due to non-living particles and estimate zooplankton size spectra. This extensive data set provides a new look at regional and inter annual variability of the pelagic ecosystem of the Bay of Biscay.

## 1 Introduction

Size matters and pelagics ecosystems are not an exception (Stemmann and Boss, 2012). Physiological rates as well as predator-prey interactions of zooplankton are thought to be size-dependent (Peters and Wassenberg, 1983; Glazier, 2005; Barnes et al., 2011). Therefore, the zooplankton size structure was used as a powerful ecological indicator of the dynamics of zooplankton communities (Sheldon et al., 1972;

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Krupica et al., 2012) and is nowadays considered and resolved in an increasing number of ecosystem models (e.g., Baird and Suthers, 2007; Ward et al., 2012). In particular, size-based approach reveals appropriate in trophodynamics observation or modeling studies linking lower and upper trophic level ~~models~~ (Daewel et al., 2013), i.e. looking at potential top-down control on zooplankton, or estimating food availability to fish. For the latter case indeed, size of prey field is key, especially for fish larvae (e.g., anchovy and sardine, Poulet et al., 1996; Morote et al., 2010). To improve model realism, Daewel et al. (2008) first proposed a simple approach by sorting modeled zooplankton biomass in size classes according to the average slope of ~~observed in-situ~~ zooplankton size-spectra. Size structured observations of the zooplankton are thus needed to provide prey field to fish models, investigate possible simplification through, e.g., biomass ~~slopes~~ relationships and to calibrate size structured zooplankton models.

However, measuring the size distribution of the zooplankton is not straightforward; various instruments exist, each of them having pros and cons. The size structure of the zooplankton could be measured by ~~in-situ~~ particles counters such as the (Laser) Optical Plankton Counter (OPC and LOPC Herman et al., 2004) that measure the size of all particles crossing a light or laser field. ~~The LOPC being able to count particles~~ from a diameter of 100  $\mu\text{m}$  to few millimeters. The discrimination between living and non-living particles with these instruments is ~~however~~ not possible although using derived parameters, such as transparency, allows some discrimination **for larger particles** (> 2 mm for the LOPC) in particular cases (Checkley Jr et al., 2008). Recently, Petrik et al. (2013) proposed a way of discriminating small (< 2 mm) living and non-living particles counted by the LOPC by fitting a log-normal curve to the volume spectrum; residuals are considered to be living particles. Another way of measuring the zooplankton size structure is through traditional net samples followed by manual or automated counting and sizing (Vandromme et al., 2012). Even though this procedure may be the most accurate, the large amount of time required generally prevent its widespread use. The last way of measuring zooplankton size-structure is through in situ imaging systems (e.g., Picheral et al., 2010). **Yet**, these instruments are still recent and not yet widely used to observe

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the zooplankton. In the present work, a combination of the LOPC (Herman et al., 2004) and net sampling followed by in lab scanning of the samples (Vandromme et al., 2012) is used.

Few studies provide zooplankton biomass or size structure at the scale of the whole Bay of Biscay. Information is available either dispersed in time and space as reviewed by Poulet et al. (1996), as long time-series from transects but locally for the north of Spain (Valdés et al., 2007) or in front of the Gironde estuary (Albaina and Irigoien, 2004), from a single season and one (Nogueira et al., 2004) or two years (Sourisseau and Carlotti, 2006) as biomass and size structure of the mesozooplankton from an Optical Plankton Counter, or as a time-series of biomass and size-spectrum maps over the southern french shelf of the Bay over a decade in spring (Irigoien et al., 2009).

The objective of the present work is to propose an homogeneous estimation of the size structure of the zooplankton over the whole shelf of the Bay of Biscay, using an extensive dataset covering the period 2005–2012 in spring and a region running from the north of Portugal to Brittany in France. This by combining the use of an in situ Laser Optical Plankton Counter (LOPC, Herman et al., 2004) and from net samples (WP2 net) followed by an image analysis procedure (Zooscan (WP2/Zs) Gorsky et al., 2010; Vandromme et al., 2012). These two instruments allows the quantitative measurement of particles between 100 and  $\sim 2000 \mu\text{m}$  ESD (Equivalent Spherical Diameter) for the LOPC and between 400 and  $\sim 2500 \mu\text{m}$  ESD for the WP2 net (Nichols and Thompson, 1991; Vandromme et al., 2012) adapted to the sampling of the meso-zooplankton. Comparison between size structure observed by in situ particles counters and net catches generally show lower or equal estimates of abundance in each size by net catches (González-Quirós and Checkley, 2006; Schultes and Lopes, 2009; Gaardsted et al., 2010). This was attributed mainly to aggregates and detritus that are often not analyzed in net catches but also disaggregated by the net passage and thus not correctly sampled. E.g., González-Quirós and Checkley (2006) found that the abundance of particles estimated with the OPC in the size range of 1.26 to 6.35 mm ESD was 4 times higher than the abundance of zooplankton estimated by concomitant bongo net

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in the California current ecosystem. This is confirmed by in situ data from imaging systems were zooplankton accounts for a small portion of total particles (often less than 5 %, Stemann and Boss, 2012). In the Bay of Biscay, Poulet et al. (1996) estimated the mesozooplankton biomass to be in the range of 4 to 20 % of the total Particulate Organic Carbon from a literature review. Fragile gelatinous zooplankton could also be destroyed by the net passage and therefore may account for part of the difference between in situ particles counters and net catches.

By investigating differences between size-spectra measured by the LOPC (zooplankton and non-living particles) and by the WP2/Zs (zooplankton only) we propose a statistical correction based on correlations between the differences observed and environmental data to estimate the zooplankton size-spectra where WP2/Zs data are missing. This allows the large available dataset from surveys made with the LOPC to be used for the characterization of the size structure patterns of the zooplankton in the Bay of Biscay. Estimated zooplankton size spectra are further investigated to depict correlations between total zooplankton biomass and the shape of the size distribution. We then discuss the results in terms of spatial and interannual variability of the habitats as defined by the size spectra distribution, as well as in terms of trophic transfer to upper trophic levels, in both an ecology and for a modelling perspectives.

## 2 Materials and methods

### 2.1 Study area and sampling

The Bay of Biscay is a large gulf of the Atlantic Ocean located off the western coast of France and the northern coast of Spain, between 43.5 and 48.5° N and 3 and 8° W. The principal rivers in decreasing order of drainage area are: the Loire, Garonne–Dordogne (Gironde complex), Adour, Vilaine and Charente rivers (Fig. 1). The continental shelf reaches widths of about 140 km off the coast of Brittany but narrows to less than 15 km off the Spanish shore. The physical and hydrological features of the Bay of Biscay

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are of great complexity, e.g. coastal upwelling, coastal run off and river plumes, seasonal currents, eddies, internal waves and tidal fronts (Lavin et al., 2004). These abiotic processes greatly influence the phytoplankton dynamics and as a consequence, the whole food web composition, structure and functioning (Varela, 1996). The samples come from the PELGAS and PELACUS cruises, French and Spanish small pelagic surveys held in spring in the Bay of Biscay on board *NO Thalassa*. At each station and at night a structure containing a CTD probe (seabird SBE19) and several sensors including an LOPC is dropped from the surface to the bottom depth (max 600 m) at  $1 \text{ ms}^{-1}$ . For some stations, a WP2 net is dropped to 100 m depth (or to the bottom for coastal stations) and towed vertically to the surface. The total numbers of stations with LOPC and/or WP2/Zs data are presented on Table 1. The dataset comprises sampling from PELGAS cruises in 2005, 2007, 2009, 2010, 2011 and 2012, and from PELACUS cruises in 2005, 2006, 2007 and 2009. The LOPC was not operated in missing years.

## 2.2 Measurement of the size distribution

### 2.2.1 Laser-Optical Plankton Counter (LOPC)

The Laser Optical Plankton Counter (Herman et al., 2004), is an optical instrument capable of measuring in situ the size of particles crossing its field by means of recording laser intensity attenuation. Smallest recorded objects are of  $\sim 100 \mu\text{m}$  ESD and largest are of few millimeters to centimeters. In the present work we limited the analysis to sizes below 1.9 mm ESD that corresponds to the maximum size of SEPs (MEPs lower than 1.9 mm ESD were added to the size distribution). SEPs are Single Element Particles and MEPs are Multiple Element Particles, which means that MEPs activate more than one diode when crossing the laser field of the LOPC. The LOPC takes two records per seconds and has a sampling window of  $49 \text{ cm}^2$ . Problems of coincidence in case of high particles concentration that arise often with the OPC (Woodd-Walker et al., 2000) are largely decreased with the LOPC (Herman et al., 2004). The LOPC is mounted on the CTD-Rosette and samples the water column at a rate of 2 Hz giving, at a maximal

lowering speed of  $1 \text{ ms}^{-1}$ , a minimal vertical resolution of 0.5 m. A total of 816 LOPC casts are used in the present work (Table 1).

## 2.2.2 WP2 net samples and in lab imaging analysis (WP2/Zs)

Zooplankton sampling was done at night by vertical tows (100 m depth or bottom depth to surface) of a WP2 net (mesh size of  $200 \mu\text{m}$ , mouth aperture of  $0.25 \text{ m}^2$ ). Since no flowmeters were mounted on nets, the sampled volume was simply calculated as the maximum depth times the mouth aperture of the net. According to Nichols and Thompson (1991) a net is quantitative for objects which have a width of a minimum of  $4/3$  the mesh of the net (here  $200 \mu\text{m}$ ). Considering a minor: major axes ratio (the minor axis is here considered as a proxy of the width) of 0.45 (Vandromme et al., 2012, average for copepods) it implies that the WP2 will quantitatively sampled objects from  $\sim 400 \mu\text{m}$  ESD, which corresponds to the mode of the size spectra found from WP2 samples (Vandromme et al., 2012). Through comparison with larger net, the upper size limit of quantitative sampling is  $\sim 2500 \mu\text{m}$  ESD (Vandromme et al., 2012). A selection of available WP2 samples representative of different locations of the area were processed with the ZooScan/ZooProcess system, a laboratory flatbed scanning system used for the digitization of fixed wet net samples developed at the Laboratory of Oceanography of Villefranche-sur-mer (Gorsky et al., 2010). Each sample was separated in two size fractions with a mesh of  $1000 \mu\text{m}$  to avoid underestimation of large objects. Then, each size-fraction subsample was fractionated separately with a Motoda box. Each subsample was scanned after a manual separation of objects on the scanning tray to reduce the occurrence of touching objects (Vandromme et al., 2012). The ZooScan used here had a scanning surface of  $15 \times 24 \text{ cm}$  and a pixel width of  $10.56 \mu\text{m}$  (scan at 2400 dpi;  $14\,200 \times 22\,700$  pixels). Images are analyzed by a dedicated imaging software, ZooProcess. This software, (i) scans a high quality raw image, linked with associated metadata, (ii) divides the raw image into separate valid targets, defined by the continuity of pixels with grey levels lower than a threshold of 243 (8 bits grayscale;

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black = 0 and white = 255), and (iii) measures 46 variables for object characterization, including grey levels, fractal dimension, shape and size (Gorsky et al., 2010). Using these variables, a supervised classification of objects into groups was performed consisting of a rough pre-sorting that was further visually corrected to ensure discriminating zooplankton from non-living particles. Zooplankton images from PELGAS cruises were then classified in 18 categories. A total of 89 WP2/Zs samples are used in the present work (Table 1).

### 2.2.3 Size spectra calculation

The size spectra are expressed as Normalized Biomass Size Spectra (NBSS) in the sense of Platt and Denman (1977) and Blanco et al. (1994), i.e.

$$\beta(w) = \frac{B(w)}{\Delta(w)} \quad (1)$$

where  $w$  is the carbon weight for a given size class,  $\beta(w)$  is the normalized form,  $B(w)$  is the unnormalized form and  $\Delta(w)$  is the width of each size class. The carbon weight is calculated from the ESD of each objects (both for the LOPC and the WP2/Zs) following conversions for general zooplankton of Lehet and Hernández-León (2009) that relate ESD to dry weight and according to a dry weight to carbon ratio of 0.447 (Mauchline, 1998):

$$ar = \pi (ESD/2)^2 \quad (2a)$$

$$dw = 43.38 ar^{1.54} \quad (2b)$$

$$w = 0.447 dw \quad (2c)$$

$ar$  is the projected 2-D area of the object (in  $mm^2$ ),  $dw$  is the dry weight (in  $\mu g$ ) and  $w$  is in  $\mu gC$ . Size spectra are displayed in logarithmic scales and a log-linear regression is performed to compute the slope ( $s$ ) and the intercept ( $a$ ) for each of them. The

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log-linear regression was made from the average mode of size spectra. While nonlinear functions usually produce a better fit to size distribution data (Gasol et al., 1991; Vidondo et al., 1997; Brucet et al., 2005), the ecological meaning of the model parameters is normally difficult to interpret (Quintana et al., 2008) and so, those nonlinear functions have not been applied in the present analysis.

## 2.3 Environmental dataset

For each station a set of hydrological parameters were extracted, using mainly measure of the CTD probe. For temperature, salinity and density we used the value at surface and bottom (surface  $\sim 4$  m depth, bottom corresponds to 100 m or less, depending on the bathymetry). We included also the total fluorescence integrated from surface to bottom, the depth of the maximal fluorescence and the value of fluorescence at its maximum. Fluorescence data were calibrated each year against in situ measurements of chlorophyll *a*. From density we computed the stratification (first differences) and used the mean value as well as the maximum one. We also included the mixed layer depth which is calculated by a finite difference criteria method ( $0.02 \text{ kg m}^{-2}$  more than the surface (4 m depth) value, Kara et al., 2000). In addition we included daily satellite data, chlorophyll *a* and inorganic Suspended Particulate Matter (SPM) processed with a specific algorithm for Case 2 coastal waters (Gohin et al., 2005); at the sampling date as well as 7 and 14 days before. Finally, bathymetry, distance to the coast, latitude and julian day were also included. This builds a total of 22 explanatory variables. Integrated and maximum fluorescence, satellite chlorophyll *a* and suspended matter as well as bathymetry were log transformed. At some stations, the surface chlorophyll *a* was measured for three size fraction, i.e. below  $3 \mu\text{m}$ , from  $3$  to  $20 \mu\text{m}$  and above  $20 \mu\text{m}$ .

## 2.4 Analysis

We are assuming that the WP2/Zs estimates the size distribution of the zooplankton only while the LOPC estimates the size distribution of both the zooplankton and

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non-living particles (organic and non-organic particulate matter) and thus gives always a larger estimate than the WP2/Zs. The LOPC size spectra were first averaged over the same depth intervals as the WP2/Zs (i.e., up to 100 m or bottom depth). According to the size specificities of each sampling device (LOPC and WP2/Zs, see Sect. 2.2), the window of comparable size ranges from  $\sim 400$  to  $2000 \mu\text{m}$  ESD. We then considered the size distribution of non-living particles as the subtraction of the WP2/Zs NBSS from the LOPC NBSS on this size range. On it we performed a log-linear regression and we will consider the slope ( $a_{\text{nl}}$ ) and the intercept ( $b_{\text{nl}}$ ) of this regression as the descriptors of the non-living (nl) particles size distribution. We can notice that even if  $a_{\text{nl}}$  and  $b_{\text{nl}}$  are generally independent, in the special case of the NBSS, where biomass values are constrained by the carrying capacity of the ecosystem, some dependency exists between these two descriptors. Statistical analyses performed hereafter however took this dependency into account.

In order to assess the main factors explaining the size distribution of non-living particles we performed a Stepwise Redundancy Analysis (Stepwise RDA Legendre and Legendre, 2012, Chap. 11.1). RDA is the direct extension of multiple regression to the modelling of multivariate response data, redundancy being synonymous with explained variance. A table of explanatory variables ( $\mathbf{Y}$ ) is used to explain a table of responses variables ( $\mathbf{X}$ ). The ordination of  $\mathbf{Y}$  is constrained in such a way that the resulting ordination vectors are linear combinations of the variables in  $\mathbf{X}$ . Here, the table  $\mathbf{X}$  corresponds to the two coefficients ( $a_{\text{nl}}$  and  $b_{\text{nl}}$ ) that will be applied on the LOPC NBSS to estimate the WP2/Zs NBSS. The table  $\mathbf{Y}$  is a set of explanatory variables described in Sect. 2.3 together with the NBSS LOPC derived biomass (in  $\log_{10}$ ) and slope. A selection of explanatory variables is performed via a Stepwise RDA. Variables are sequentially added by selecting the one that yields the largest variance (partial F-statistic) and by removing it for the next selection step. Variables are thus sorted from the one that explains the most variance to the lowest.

To calculate the coefficients of the regression that estimate  $a_{\text{nl}}$  and  $b_{\text{nl}}$  we used a Partial Least Square regression (PLS regression, de Jong, 1993; Rosipal et al., 2006). PLS

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regression is used to find fundamental relations between two matrices ( $\mathbf{X}$ , the table of responses variables and  $\mathbf{Y}$ , the table of explanatory variables), i.e. a PLS model will depict the multidimensional direction in the  $\mathbf{Y}$  space that explains the maximum multidimensional variance direction in the  $\mathbf{X}$  space through computing principal components of the matrix  $\mathbf{Y}$ . PLS regression are generally used when there is a large number of explanatory variables and a covariance among them. The robustness of the PLS regression was assessed by performing a total of 5000 permutations with a random removal of 4 stations for each permutation. Average PLS regression coefficients are then applied on the whole LOPC size spectra to estimate the zooplankton size distribution for each station of each year occurring in the dataset. These steps are shown on Fig. 2.

Further, the zooplankton estimated productivity is calculated by using the formulation of Zhou et al. (2010) based on the model of Huntley and Boyd (1984) and Hirst and Bunker (2003). The growth of zooplankton is a function of water temperature, chlorophyll  $a$  and weight of individuals (in mgC). The production is the growth rate multiply by the total biomass of zooplankton. The total biomass is calculated as the integral of an assumed linear NBSS:

$$B = \int_{x_{\min}}^{x_{\max}} ax^s dx \quad (3a)$$

$$B = \frac{ax_{\max}^{s+1}}{s+1} - \frac{ax_{\min}^{s+1}}{s+1} \quad (3b)$$

where  $B$  is the biomass between  $x_{\min}$  and  $x_{\max}$  and  $s$  is the slope and  $a$  the intercept of the NBSS. Here,  $x_{\min}$  corresponds to 200  $\mu\text{m}$  ESD and  $x_{\max}$  corresponds to 2000  $\mu\text{m}$  ESD, which cover the size range of the meso-zooplankton. the production is summed from the surface to a max of 100 m depth to have an estimated productivity per square meter.

Maps of zooplankton estimated biomass, slopes and productivity are computed for each year on a 50  $\times$  50 grid ( $\sim 20 \times 20$  km cells) covering the whole Bay of Biscay. Val-



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ues at nodes are computed as the average of values in a radius of 30 km weighted by the inverse of the squared distance. Climatological maps is then calculated by taking the mean of each annual map of zooplankton biomass, slopes and productivity. The biomass of zooplankton is presented both as the average concentration, in  $\text{mgCm}^{-3}$ , from the surface to the bottom or to 100 m depth and as the integrated value, in  $\text{mgCm}^{-2}$ , from the surface to the bottom or to 100 m depth. Maps of standard deviation and number of years with value at each node are also presented.

In the last part, 5 groups are isolated by means of  $K$  means clustering computed on normed values of slopes and log-transformed biomass. The number of groups is arbitrary and was chosen as the best trade-off between the level of details needed to detect patterns and lisibility. The inter-annual and spatial variability of these groups is further investigated.

All analysis were performed with Matlab R2012a (The Mathworks) with the use of the statistics toolbox (The Mathworks) and the Fathom toolbox for the Stepwise RDA (maintained by David L. Jones, [www.marine.usf.edu/user/djones](http://www.marine.usf.edu/user/djones)).

## 3 Results

### 3.1 Size distribution of non-living particles

The LOPC size spectra averaged from surface to 100 m depth (or bottom depth) show slopes ranging from  $-1.73$  to  $0.40$  among all 816 stations, with a mean of  $-0.97 \pm 0.24$  (of standard deviation). Intercepts range from  $-1.88$  to  $5.15$  with a mean of  $1.76 \pm 1.39$ . The coefficients of determination against the linear regression shows a mean of  $0.95 \pm 0.03$  highlighting the high linearity of the size distribution. WP2/Zs size spectra show slopes ranging from  $-2.08$  to  $0.10$  with a mean of  $-0.86 \pm 0.40$ , and intercepts ranging from  $-2.36$  to  $2.54$  with a mean of  $0.15 \pm 1.20$ . The coefficients of determination of WP2/Zs size spectra against the linear regression were  $0.92 \pm 0.06$ , slightly less than LOPC size spectra. LOPC and WP2/Zs slopes have a Pearson correlation coefficient of

0.57 and their intercepts of 0.62. On Fig. 3 various examples of LOPC and WP2/Zs size spectra measured at same stations are presented. Examples include an overestimation by the LOPC for each size (Fig. 3a), an overestimation for lower size only (Fig. 3b) and no differences between both spectra (Fig. 3c). The first case corresponds to a larger

LOPC intercept but no difference in the slope estimate, in the second case the LOPC intercept is larger and its slope is steeper than of the WP2/Zs, then in the third case no difference exist in the intercept and slope of both. Differences in slope and intercepts for all the 89 stations are shown on Fig. 4. It corresponds to intercepts and slopes of the size distribution of non-living particles within this size range. For some cases, the slope of the WP2/Zs was steeper than that of the LOPC (red colored), however, the WP2/Zs never overestimated the LOPC in the present dataset. These cases appear in PELGAS 2009 and 2010 and are mainly located on very coastal stations. Stations where both devices estimate are comparable (small white circles) are generally located off-shore in the middle of the continental shelf or over the slope of the French slope. For other cases, no geographical pattern clearly emerged. During PELGAS 2011 (Fig. 4e) non-living particles seem mainly distributed in the southern part of the French continental shelf, yet, this was not the case for other years.

To depict the main variables ( $Y$  matrix) affecting the size structure of non-living particles ( $X$  matrix) we performed a stepwise Redundancy Analysis (RDA, see Sect. 2.4), results are shown in Table 2. The cumulative explained variance (Cum. Exp.) for all variables included reaches 73 % of the total variance of non-living particles slopes and intercepts. The variable that explains the most variance is the total biomass of particles in the water column estimated from LOPC (40 % of variance explained), followed by the chlorophyll  $a$  measured by satellite at the sampling time (52 % Cum. Exp.), the bathymetry (56 % Cum. Exp.) and the slopes of the size distribution of all particles measured with the LOPC (59 % Cum. Exp.). Since the variance explained by a variable is removed prior to the selection of the next variable, a variable strongly correlated with a previous extracted one is not likely to add a lot of cumulated explained variance. The variance explained by each variable on the initial  $X$  matrix is shown in the column Tot.

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Expl. of the Table 2. For example, the julian day explains 12 % of the initial **X** matrix but appears on the 9th position in the stepwise RDA and add only 1 % to the cumulative explained variance.

To further depicts variables explaining the variance of the non-living particles size distribution we added supplementary variables that were available only for some stations, thus we worked on a reduce dataset. The Table 3 shows the effect of including fractionated surface chlorophyll *a* to the **Y** matrix. The chlorophyll *a* was measured for the fraction below 3 µm (chlorophyll *a* 1), then from 3 to 20 µm (chlorophyll *a* 2), and above 20 µm (chlorophyll *a* 3). A total of 68 stations (among the 89) were available and belong to PELGAS 2009, 2010 and 2011. On these 68 stations the total cumulated explained variance without fractionated chlorophyll *a* is 74 % and reaches 80 % with it. The first variable to be extracted is, as above, the LOPC derived biomass (41 % Cum. Exp.) followed by the chlorophyll *a* in the larger size class (51 % Cum. Exp.), the bathymetry (56 % Cum. Exp.) and the chlorophyll *a* in the second size class (59 % Cum. Exp.). This first four variables accounts for the same explained variance as the first four above (Table 2). A last stepwise RDA was done, using taxonomic data (total of 18 zooplankton categories) from the WP2/Zs (75 stations belonging to PELGAS 2009, 2010 and 2011). Results are shown on Table 4. The stepwise RDA with same explanatory variables as above (Table 2) shows a total cumulated explained variance of 71 %, whereas all the 18 zooplankton categories explain a total of 92 % of the variance. The first four zooplankton categories explains 82 % of the variance (*Acartia* sp., cladocerans, *Calanus* sp. and other calanoids smaller than 1 mm length, see Table 4).

### 3.2 Accuracy of estimated zooplankton size-spectra

Using explanatory variables we estimated the size distribution of the zooplankton from size spectra measured with the LOPC, i.e., we removed the non-living part from LOPC size-spectra to approach size-spectra measured with the WP2/Zs that are assumed to represent the best estimate of the zooplankton size distribution. For that purpose we performed a Partial Least Square (PLS) regression (see Sect. 2.4) of the explanatory

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variables (matrix **Y**) on the response matrix (**X**). For the 5000 permutations performed the average explained variance was of  $74.7 \pm 1.9\%$ , similar to the stepwise RDA. Estimated parameters of the non-living particles size spectra were then subtracted to the initial LOPC size spectra (hereafter, LOPC) to obtain the estimation of the zooplankton size spectra (hereafter, PLS reg-). Correlations between WP2/Zs slopes and intercepts and LOPC ones before and after the subtraction of non-living particles are shown on Fig. 5 and on Taylor diagrams (Fig. 6, Taylor, 2001). The Pearson correlation between WP2/Zs slopes and intercepts with the ones of the LOPC are respectively of 0.57 ( $p < 0.001$ ) and 0.62 ( $p < 0.001$ ). In the case of the intercepts (Fig. 5b) a strong overestimation by the LOPC is also observed. After the subtraction, Pearson correlations reach 0.78 and 0.85 (both  $p < 0.001$ ) for slopes and intercepts respectively. The overestimation of the intercept by the LOPC is reduced with the PLS reg-. This is clearly seen on the Taylor diagram (Fig. 6b).

WP2/Zs slopes ( $s$ ) and intercepts ( $a$ ) are significantly correlated (Pearson  $r = -0.86$ ,  $p < 0.001$ ) and related by a linear fit on the form  $a = p_1 s + p_2$ , where  $p_1 = -2.568(-2.895, -2.241)$  and  $p_2 = -2.062(-2.372, -1.752)$ , numbers in parenthesis indicate the 95 % confidence interval. For the LOPC, the Pearson correlation is of  $-0.70$  ( $p < 0.001$ ) and the linear fit has coefficients  $p_1 = -4.144(-4.445, -3.842)$  and  $p_2 = -2.251(-2.551, -1.951)$ . Slopes and intercepts estimated by the PLS regression are also significantly correlated (Pearson  $r = -0.84$ ,  $p < 0.001$ ) and have coefficients  $p_1 = -3.181(-3.331, -3.031)$  and  $p_2 = -2.606(-2.727, -2.486)$ . These correlations are shown on Fig. 7. The subtraction of non-living particles tends to reproduce the initial correlation that exists between slopes and intercepts of the zooplankton size spectra.

### 3.3 Spatial and inter-annual distribution of zooplankton

On Fig. 7 the estimated biomass computed as the integral of a linear NBSS defined by its slope and intercept is displayed as a color scale in  $\log_{10}$ . The strength of the relation between biomass and size spectra parameters is affected by the position of the

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subset slopes vs. intercepts. For the values of slopes and intercepts observed here we have a Pearson correlation of biomass and slopes of  $-0.51$  ( $p < 0.001$ ), the relation is displayed on Fig. 11:  $\log_{10}(B) = -0.571s + 0.544$ . The estimated zooplankton biomass concentrations range from 2.9 to  $63.7 \text{ mg C m}^{-3}$  with a mean of  $11.4 \pm 8.2 \text{ mg C m}^{-3}$  (median of  $8.9 \text{ mg C m}^{-3}$ ). The depth integrated zooplankton biomass ranges from 37.7 to  $4614.5 \text{ mg C m}^{-2}$  with a mean of  $804.2 \pm 421.0 \text{ mg C m}^{-2}$  (median of  $753.6 \text{ mg C m}^{-2}$ ). A general tendency of large biomass associated with steeper size spectra is observed. This tendency also emerged from climatological maps (Figs. 8, 9 and 10) where maximum values of biomass are observed near the coast of the northern part of the Bay of Biscay (especially off the Vilaine estuary) and on the coast of Galicia which roughly corresponds to the steepest slopes observed. Lowest biomass values are observed on average in the southern part of the Bay, in the Cap Breton area and off Portugal. Low values occur also throughout the French continental shelf. In term of biomass, one of the most variable area observed is the Gironde plume (Fig. 8c).

The inter-annual and spatial patterns were investigated through the 5 groups identified by the  $K$  means clustering based on estimated slopes and biomass of the zooplankton (Fig. 11). Annual and spatial distribution of these groups is shown in Fig. 12. The first group (blue circles) corresponds to largest biomasses concentration (median of  $32 \pm 11.8 \text{ mg C m}^{-3}$ ) and steepest size spectra (median of  $-1.19 \pm 0.14$ ). This group is almost exclusively found in the most coastal areas, notably in the main estuaries of the french coast (Gironde, Charente, Loire, Vilaine) but also in the coastal area south of the Gironde estuary (the Landes), in the coast of western Brittany and along the Galician coast. An important inter-annual variability is observed with the quasi absence of this group during the spring of 2011 compared to springs of 2009 and 2010, the two years with the most occurrence of this group. The second group (green squares) shows the second largest biomass concentration (median of  $15.9 \pm 5.2 \text{ mg C m}^{-3}$ ) and second steepest size spectra (median of  $-0.94 \pm 0.13$ ). This group occurs, as group 1, mainly in coastal areas, although it extends more over the plateau. This group shows less inter-annual variability than the first one with, yet, a higher occurrence during spring 2011,

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notably in estuarine areas. The third group (red losanges) has a median biomass concentration of  $7.9 \pm 1.5 \text{ mg C m}^{-3}$  and slopes of  $-0.83 \pm 0.10$ . In 2005 and 2007 this group is observed mainly in the spanish and portugese coasts, while, from 2009, many occurrence of this group were observed further north, specifically in the Cap-Breton area (south-east of the bay), mid-plateau and continental slope. The forth group (black triangles) has the lowest biomass concentration (median of  $4.9 \pm 1.0 \text{ mg C m}^{-3}$ ) and second flatest size spectra (median of  $-0.66 \pm 0.10$ ). As the latter group, the forth group is mainly observed at southern location (Cantabrian sea and portugese slope) in 2005, 2006 and 2007. In 2005, this group was also observed in the Cap-Breton area. From 2009, an extension northward is observed with many occurrence in the french slope and off the Cap-Breton. In 2012, this group is also observed mid-plateau up to Brittany. The last group (pink stars) has biomass concentrations of  $9.5 \pm 2.5 \text{ mg C m}^{-3}$  and the flatest size spectra (median of  $-0.55 \pm 0.10$ ). This group is the most off-shore, with most occurrence mid-plateau and at the slope. This group was also observed, mainly in 2007, in the spanish coast. In 2009 and 2010, stations close to the Gironde estuary belonging to this group were recorded. In 2012 this group was also observed at some locations along the Brittany coast.

### 3.4 Zooplankton productivity

The productivity of zooplankton, in term of  $\text{mg C m}^{-2} \text{ d}^{-1}$ , was computed using a model based on the size of the zooplankton, water temperature and chlorophyll *a* (see Sect. 2.4). Estimates ranged from 0.0011 to  $0.58 \text{ mg C m}^{-2} \text{ d}^{-1}$ , with a mean of  $0.036 \pm 0.050$  and a median of  $0.021 \text{ mg C m}^{-2} \text{ d}^{-1}$ . The climatological map of zooplankton productivity, calculated as for Figs. 8 and 10, is presented on Fig. 13. On average, the less productive area, but also the most variable, is the area off Cap Breton and the eastern coast of the Cantabrian Sea. Most productive area are the river plume of Gironde and Loire/Vilaine, but also the French continental slope followed by the Galician area.

## 4 Discussion

### 4.1 A large scale homogeneous sampling

Several studies aim to compare efficiency of sampling methodologies in providing size structure of mesozooplankton, among them LOPC and net tow combined with identification or image analysis (Nogueira et al., 2004; Schultes and Lopes, 2009). Here we took advantage of both methodologies, taxa resolution of the zooscan with convenience of the LOPC to provide homogeneous size structure information at the scale of the Bay of Biscay shelf. French Pelgas survey directly follows spanish Pelacus survey aboard the same vessel, and both operate similar LOPC and WP2 net tows. This limits sources of errors from sampling differences when further comparing spatial variability over this large area. However this also means a large difference between starting date of first survey in the south and end of second one in the north (~ 2 months). This potentially bias the interpretation we make on spatial variability, which encompass seasonal evolution, especially during the spring season when rapid changes in the environment occurs (Koutsikopoulos et al., 1996). This effect is partly mediated since the productive season progresses from south to north following temperature and stratification increases (Koutsikopoulos et al., 1996), which has the effect of attenuating differences arising from seasonality. In addition, the repeatability across years of spatial patterns when looking at NBSS slope with low standard deviation, or habitat classification from clustering, especially over the French shelf for which more years are available, provide arguments for considering our climatologies as robust in representing the spring situation.

The sampling strategy of the survey prevents any assessment of observed variability at short temporal (Sourisseau and Carlotti, 2006) and spatial scale, while interpretation may be affected by population dynamic patterns or patchiness. However, as noticed by Sourisseau and Carlotti (2006), the spatial variability at our sampling scale is higher than the temporal variability, giving confidence to our results. Distance between stations are approximately 24 nm (~ 44 km) in the alongshore direction, and 10 to 25 nm (~ 19

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to 46 km) in the cross-shelf direction which shows strongest gradients, with refinement for the latter in the known frontal areas and over the Iberian shelf. Based on analysis of Albaina and Irigoien (2004), a minimum resolution of 12 nm ( $\sim 22$  km) is necessary in the cross-shelf direction to obtain realistic patterns of mesozooplankton distribution, thus our sampling should allow identification of major spatial patterns of size distribution and habitat at mesoscale.

Non-linearity of the size spectrum is a common observed feature (Nogueira et al., 2004; Sourisseau and Carlotti, 2006) and reflects a non equilibrium state as compared to theoretical models (Kerr and Dickie, 2001). However, two arguments were in favour of using a linear regression model to calculate the NBSS. First we found high coefficients of determination for the adjusted models at each station and for both instruments. Second, in our study and with our low temporal resolution data we are more interested here at looking at principal scalings of the ecosystem structure (zooplankton size spectrum and its spatial variability, Kerr and Dickie, 2001), that is physiological (primary scaling) and ecological (secondary scaling), rather than short term population dynamics, one of the factor leading to these non-linearities. The linearisation of the measured spectrum in our analysis, as well as presentation of results through climatologies tends to filter out the short spatio-temporal variability.

### 4.2 Correction of the size spectra

To separate living and non-living particles counted by the LOPC we use a direct comparison with net catches performed at the same position and at the same time. The statistical correction was made only on the common valid part of the size spectrum for each instrument ( $\sim 400$  to  $2000 \mu\text{m}$  ESD), to reduce uncertainties arising from undersampling at lower and upper limits of the mesozooplankton size range. Differences between both spectra were assumed to originate from non-living particulate matter. LOPC counts were, within the present dataset, always higher or equal to the WP2/Zs counts giving weight to the assumption that the LOPC accurately counts both living and non-living particles. Properties of this difference (slope and intercept of the log-linear



regression) were related to the ones of the LOPC size spectra and to the environmental variables. The resulting multiple regression was further used to estimate the size distribution of non-living particles at location where only LOPC data are available.

~~In the present work,~~ the correction significantly improved the reliability of the LOPC-derived NBSS, both in terms of correlation with the WP2/Zs-derived NBSS, and of properties of the NBSS, i.e. correlation between slope and intercept. Remaining unexplained variability may have several of the following origins difficult to further assess from our study: (i) the lacking information included in the explanatory environmental variables, as well as the associated errors associated to their measurement, (ii) errors from various origin in the estimation of volume sampled by the net (Nogueira et al., 2004; González-Quirós and Checkley, 2006; Schultes and Lopes, 2009) as well as further fractioning before Zooscan processing, and (iii) various efficiency between instruments especially at the tails of the size range even if caution was taken in its selection, with potential avoidance of larger individuals.

#### 4.3 Interpretation of the correction

Our correction methodology from the LOPC NBSS towards the WP2/Zs NBSS makes best use of available environmental information, both in situ and satellite. One could think of direct statistical models to estimate the correct NBSS properties and propose some extrapolation for periods without sampling. But the occurrence of LOPC-derived biomass, and to a lesser extent of the LOPC slope, in first positions of the explanatory variables (Tables 2 and 3) for the differences between spectrum properties give strong interest of using the LOPC despite its contamination by non-living particles.

In addition to the uncorrected NBSS properties, chlorophyll (either from satellite, Table 2, or as fractionated measurements, Table 3) appears as the second explanatory variable. This confirms that the majority of the difference between both instruments is due to particulate organic matter, being highest in areas with detrital products of large phytoplankton blooms (fractions higher than 30  $\mu\text{m}$ ) or associated zooplankton blooms. Looking only at satellite-derived chlorophyll, highest variability is explained by one week

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past situation (4.47 %), again highlighting the potential role of past bloom in detritus concentration, even if in the stepwise RDA current chlorophyll situation takes priority. In opposition, inorganic SPM from satellite does not explain much variability, their small size being unavailable to both type of measurements. Julian day, taken separately, explains more than 10 % of the variability. This may be explained also by the timing of sampling with respect to bloom phenology, with highest detritus concentrations in post-bloom conditions later in spring. Non-living particulate material may also has terrestrial or resuspension origin, which is reflected in the salinity or distance to coast covariables, but here again redundancy with other variables (likely with bathymetry but also coast-to-offshore gradient of the size spectra) prevent them to be at the top of the list of the cumulated explained variance.

Interestingly, using taxonomy information, we successfully explained a larger part of the correction variability. Size structure is implicitly considered by the abundance of each taxa, but in addition this reveals that taxonomy also implicitly introduce spatial information such as distance from coast with community structuration along a cross-shore gradient (Albaina and Irigoien, 2004), likely some information on phenology and succession of plankton species, and finally some information on quantity and quality of associated detrital matter. In case that living fragile particles such as gelatinous zooplankton are also part of the difference between both instrument measurements, then taxonomy information may also bring some information on the community assemblages not explained by other covariables. This latter analysis with taxonomy was mainly exploratory since no simultaneous information is available for systematic correction, but this motivates the effort to be set on further development of in situ imaging system.

### 4.4 Size spectrum patterns

The estimated mesozooplankton biomass ( $2.9$  to  $63.7 \text{ mg C m}^{-3}$ ) is in agreement with values reviewed by Poulet et al. (1996) in the area, or with high resolution sampling in spring over the southern shelf of the Bay by Irigoien et al. (2009). The slopes of the NBSS range from  $-1.4$  to  $-0.2$  with a mean of  $-0.8$  slightly flatter that the commonly

accepted slope of  $-1$ , typical of steady-state large ecosystems (Platt and Denman, 1977; Kerr and Dickie, 2001) indicating a small : large ratio in favor of larger individuals, most likely explained by the post-bloom conditions. ~~Spatial distribution of size structure as~~ climatology confirms the remarkable positive coast to off-shore gradient of the NBSS slope, with a slight decrease when reaching the shelf break, especially over the french shelf in coherence with observations by Sourisseau and Carlotti (2006) and Irigoien et al. (2009). We do not observe an opposite gradient over the North Iberian shelf, as clearly appeared from observations by Nogueira et al. (2004) during winter to spring transition in 2002, but we observe their west to east trend of steeper to flatter slopes. Fewer years are available to built a robust climatology in that area.

From 5 yr and high resolution sampling over a transect offshore the Gironde estuary, Albaina and Irigoien (2004) described the following structuration for the mesozooplankton community: (i) a river plume area with high abundance of small individuals, (ii) a shelf break frontal zone with relatively high abundance, (iii) the shelf zone with much lower abundance but large species, and (iv) an oceanic zone. Apart from the latter that is not well sampled in our surveys except for 2011 and 2012 in the south (Fig. 12), our results show a similar cross-shelf structure over the whole area north of the Gironde. Indeed, the coastal zone shows steeper slopes from a large proportion of small individuals, mid-shelf zone has flattest slopes with highest relative proportion of large individuals, and the slope then gets steeper over the shelf break.

Despite this size spectrum gradient, the biomass is relatively homogeneous as also observed by Albaina and Irigoien (2004), with only a slight decrease in the mid-shelf area and a really coastal strip of higher biomass north of the Gironde until Brittany. The bigger size of individuals in the mid-shelf area tend to compensate the fewer abundance. The absence of cross-shelf structuration in the south of the bay and over the Iberian shelf is certainly due to the absence of the mid-shelf habitat (low occurrence of group 5, see Fig. 12) with no transition between coastal and shelf break habitats. Large continental inputs along the coast and internal waves breakdown (Pingree et al., 1981) over the continental slope are strong drivers of the ecosystem dynamics over the

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French northern shelf. Those processes are lacking or really limited over the Iberian shelf which consequently shows higher spatio-temporal variability across years.

#### 4.5 Productivity and trophic control

The integrated mesozooplankton productivity gives a significantly different picture than the biomass. The high biomass areas such as along the French and Galician coasts still appear as highly productive, but in addition the shelf slope is also quite productive when compared to the mid-shelf, which is more conspicuous again over the french shelf. This is partly due to bathymetry which tend to lower the values in coastal areas after vertical integration as compared to offshore deeper waters, but this is also explained by differences in size structure of the zooplankton and potentially primary production. The mid-shelf, with flattest slopes of the bay and large zooplankton, is estimated as less productive even if the biomass seems close to the one over the slope. This region is often rapidly and strongly stratified in spring (Koutsikopoulos et al., 1996) with low primary production limited to the thermocline after the spring bloom, and has water masses with long residence time under low residual circulation (Charria et al., 2013). In opposition, coastal areas are continuously under the influence of rich river inputs, and the slope frontal structure regularly receives nutrient inputs from breakdown of internal waves.

Variation of the slope of the size spectrum around the theoretical value of  $-1$  typical of steady-state large ecosystems (Kerr and Dickie, 2001) has ecological significance.

At the scale of our defined habitats, the slope is the result of local structuration of the community, informing on efficiency of the transfer of matter across trophic levels or type of trophic control (Suthers et al., 2006). From a literature review, Daewel et al. (2013) could not find evidence of top-down control on zooplankton at the scale of the Bay of Biscay, even if at local scale and for particular season this can not be excluded. Steeper slopes are generally related to lower efficiency of the matter flux, potentially with top-down control, while flatter slopes areas represent efficient transfer under bottom-up control. So even if more productive, coastal areas (especially plumes) and to a lesser

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extent shelf break habitats, may have low transfer efficiency, while mid-shelf area may have high transfer efficiency through the zooplankton size range. The fact that similar ranges of biomass occur in the mid-shelf and over the slope despite higher productivity over the slope is a sign of higher efficiency for the mid-shelf habitat. There are usually low fish occurrence in that habitat in spring during the small pelagic survey, which has the effect of releasing predation pressure on large zooplankton that would have their biomass rapidly declining otherwise. In opposition, the southeastern bay have high fish occurrence, which could explain the lower observed biomass than in the north under top-down control.

## 5 Conclusions

The LOPC has been operated in the Bay of Biscay since 2005 during Spanish and French small pelagic surveys. Considering that fewer stations are sampled with net due to time constraint, and the considerable amount of lab work for taxon identification or/and size measurements (more than 1 month for scanning and classification from our 76 analysed samples from Pelgas) which often prevents complete analysis of the full set of sampled stations (here 874), the LOPC combined with our methodology provides a robust and rapid access (potentially at the end of a survey) to key information on the size structure of the mesozooplankton. Further investigations of selected samples may then be further analysed with binocular or Zooscan. Alternatively, in situ imaging instruments such as the UVP (Stemmann et al., 2008) can provide both size structure and taxonomy rapidly during a survey. The UVP was tested in the Bay of Biscay but the instrument would need further development for its full efficiency in loaded coastal areas.

Combining biomass from biogeochemical model results and size spectrum observation is an investigated approach to estimate available fraction of the total biomass to upper trophic levels (Daewel et al., 2013). The significant but relatively weak correlation between slope and biomass (Fig. 7) obtained over the whole domain does not permit

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a direct estimation of this fraction from only model biomass. Alternatively, combining model results and our NBSS slope climatology can provide a first estimation. Temporal stability of the maps of size structure presented in this study has to be verified before this approach is generalised for estimation over the whole year.

Even if able to provide abundance and biomass on lower size classes with more efficiency than the WP2 mounted with a mesh of 200  $\mu\text{m}$ , the LOPC is not efficient for the microplankton size classes, also key in the energy transfer to upper trophic level, in particular as food for fish larvae. Nogueira et al. (2004) showed some continuity between the size spectra obtained from a 20  $\mu\text{m}$  net and an in situ OPC. However this has to be verified over a larger set of stations before any extrapolation to be proposed from the LOPC-derived size spectra. No in situ instrument currently exist for this size range, however, the Flowcam (Fluid Imaging Technologies), operational on-board, may provide rapidly key additional information.

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**Table 1.** Number of stations with LOPC and/or WP2/Zs data per cruises. There is a total of 816 LOPC profiles and 89 WP2 net samples analyzed with the ZooScan. The table shows also the starting and ending dates of the cruises.

Cruise	First station	Last station	#LOPC	#WP2/Zs
Pelacus 2005	5 Apr	29 Apr	100	0
Pelgas 2005	5 May	30 May	52	0
Pelacus 2006	3 Apr	18 Apr	43	0
Pelacus 2007	11 Apr	21 Apr	55	14
Pelgas 2007	27 Apr	18 May	55	0
Pelacus 2009	2 Apr	20 Apr	77	0
Pelgas 2009	26 Apr	2 Jun	105	24
Pelgas 2010	26 Apr	2 Jun	117	23
Pelgas 2011	29 Apr	3 Jun	127	29
Pelgas 2012	27 Apr	24 May	85	0
$\Sigma =$			816	89

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**Table 2.** Results of the stepwise Redundancy Analysis (RDA) and simple RDA performed on residuals from NBSSs of [LOPC-WP2/Zs]. ~~See text for details on the procedure.~~ Par. F = partial F-statistic; Cum. Exp. = cumulative fraction of variance explained; Tot. Expl. = fraction of the total variance explained; Tot. F = total F-statistic;  $p$  =  $p$  values for significance of the F-statistic; Sat. = Satellite;  $T^\circ$  = temperature; surf. = surface; stratif. = stratification; Sal. = salinity; fluo = fluorescence; MLD = mixed layer depth; SPM = suspended particulate matter; Int. = Integrated (from surface to a max of 100 m depth); d-7/14 = Satellite data 7 and 14 days before the sampling date. ~~The last column represent the direction of the correlation with respectively the slope and the intercept of the residuals (i.e., the non-living matter).~~

Variables	Stepwise RDA			Simple RDA			
	Cum. Exp.	Par. F	$p$	Tot. expl.	Tot. F	$p$	
LOPC biomass	0.40	55.02	0.01	0.4	55.02	0.01	++
Sat. chl a	0.52	20.78	0.01	0.01	0.82	0.01	+-
Bathymetry	0.56	7.68	0.01	0.02	1.29	0.01	--
LOPC slope	0.59	6.28	0.01	0.25	28.41	0.01	+-
Density bottom	0.61	3.55	0.22	0.06	5.77	0.01	--
$T^\circ$ bottom	0.62	2.90	0.12	0.01	0.63	0.52	+-
Salinity bottom	0.66	7.89	0.01	0.03	2.85	0.01	--
Sat chl a d-7	0.68	5.61	0.01	0.05	4.47	0.15	++
Julian day	0.69	2.24	0.23	0.12	11.31	0.01	+-
Latitude	0.70	1.94	0.22	0.09	8.64	0.01	+-
Sat. chl a d-14	0.70	1.09	0.11	0.01	1.09	0.24	++
Sat. SPM d-7	0.70	0.84	0.47	0.01	0.43	0.48	++
Strat. mean	0.71	1.32	0.21	0.02	1.96	0.35	++
Sat. SPM d-14	0.71	0.60	0.30	0.00	0.33	0.64	++
Depth of fluo. max	0.71	0.30	0.76	0.01	1.11	0.29	--
MLD	0.71	0.15	0.6	0.03	2.61	0.01	--
Coast dist.	0.71	0.11	0.91	0.07	6.29	0.01	--
Fluo. max	0.71	0.07	0.81	0.00	0.32	0.75	++
Sat. SPM	0.72	0.08	0.91	0.02	1.42	0.34	++
$T^\circ$ surf.	0.72	0.04	1.00	0.02	2.04	0.30	--
Strat. max	0.72	0.05	0.91	0.07	5.88	0.01	++
Salinity surf.	0.72	0.04	0.91	0.06	5.67	0.01	--
Density surf.	0.73	3.33	0.10	0.04	3.26	0.14	+-
Fluo. int.	0.73	0.00	1.00	0.00	0.01	1.00	+-

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**Table 3.** Stepwise Redundancy Analysis (RDA) performed with environmental data (as in Table 2) and with fractionated chlorophyll *a* (three fractions) from Pelgas 2009, 2010 and 2011 (total of 68 stations located in the French Shelf of the Bay of Biscay). Only the cumulative explained variance is displayed (Cum. Exp.) for the 5 first explanatory variables, the first chlorophyll *a* fraction (plus the variable just before) and the last one. On the same 68 stations, environmental data only explained 0.74 of the total variance.

Variables	Cum. Exp.
LOPC biomass	0.41
chl 3	0.51
Bathymetry	0.56
chl 2	0.59
Fluo. int.	0.60
...	
LOPC slope	0.72
chl 1	0.73
...	
Depth of fluo. max	0.80

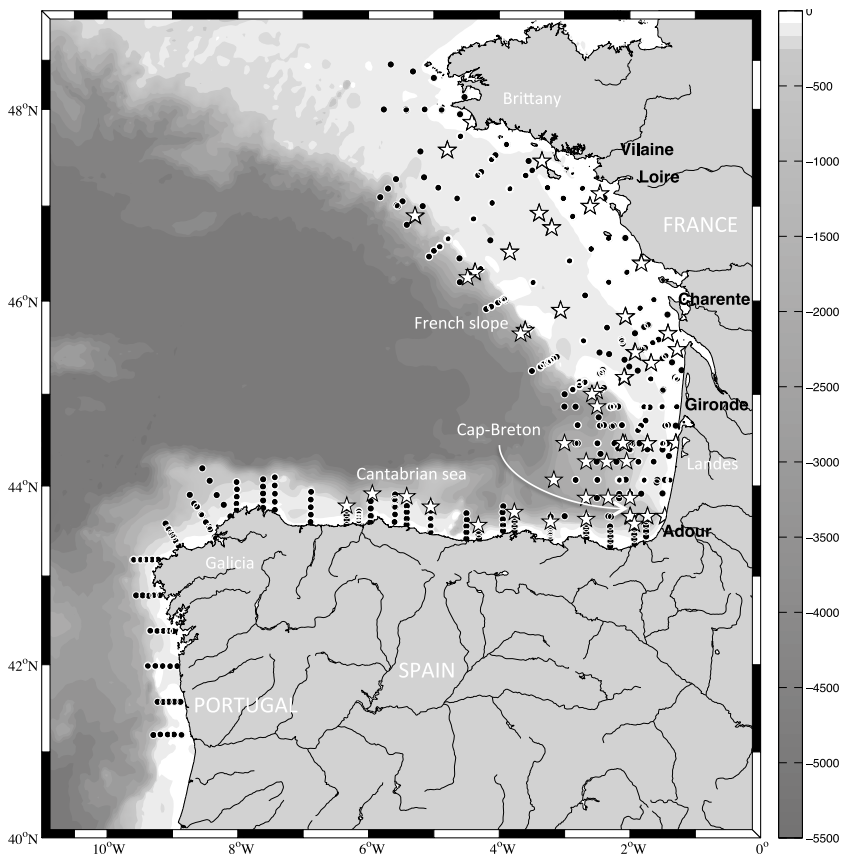
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**Table 4.** Stepwise Redundancy Analysis (RDA) performed with zooplankton group abundances from Pelgas 2009, 2010 and 2011 as explanatory variables (total of 75 stations located in the French Shelf of the Bay of Biscay). Here only the cumulative explained variance is displayed (Cum. Exp.) for the 5 first zooplankton groups and the last one. On the same 75 stations, environmental data only (as in Table 2) explained 0.71 of the total variance.

Variables	Cum. Exp.
<i>Acartia</i> sp.	0.49
cladocerans	0.66
<i>Calanus</i> sp.	0.76
small calanoids	0.82
larvaceans	0.87
...	
<i>Limacina</i> sp.	0.92

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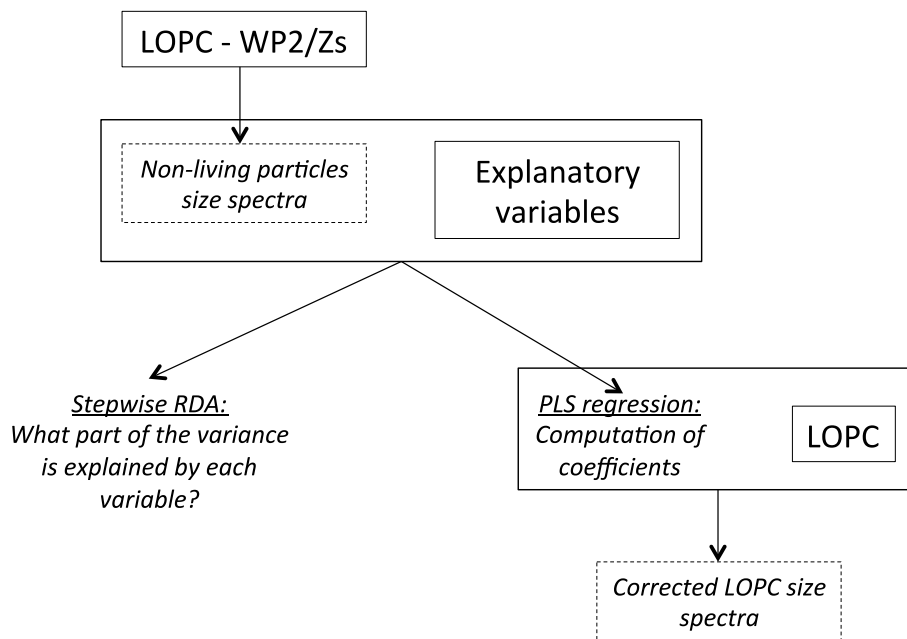



**Fig. 1.** Bathymetry of the Bay of Biscay and location of stations (black circle for LOPC only and white star for LOPC and WP2 net). Location and name of main rivers are indicated in black whereas geographical name are indicated in white.



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**Fig. 2.** Diagram showing the different analyses performed on the LOPC dataset.

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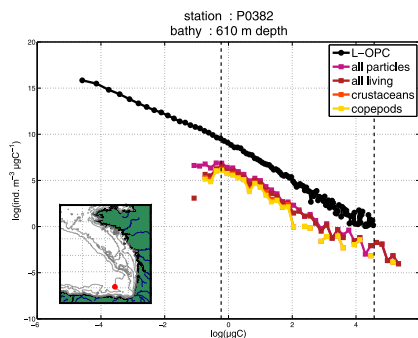
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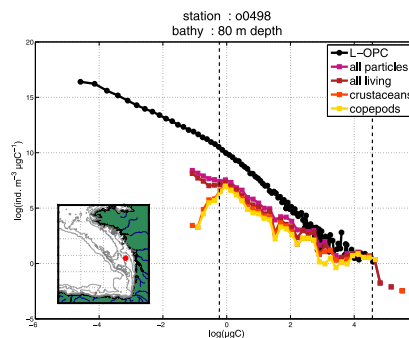
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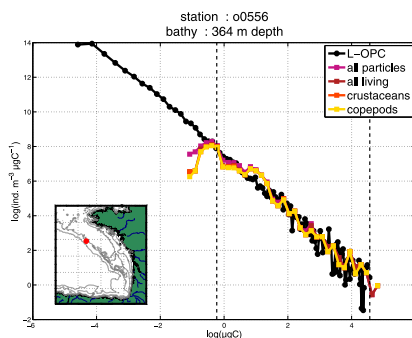
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(a)



(b)

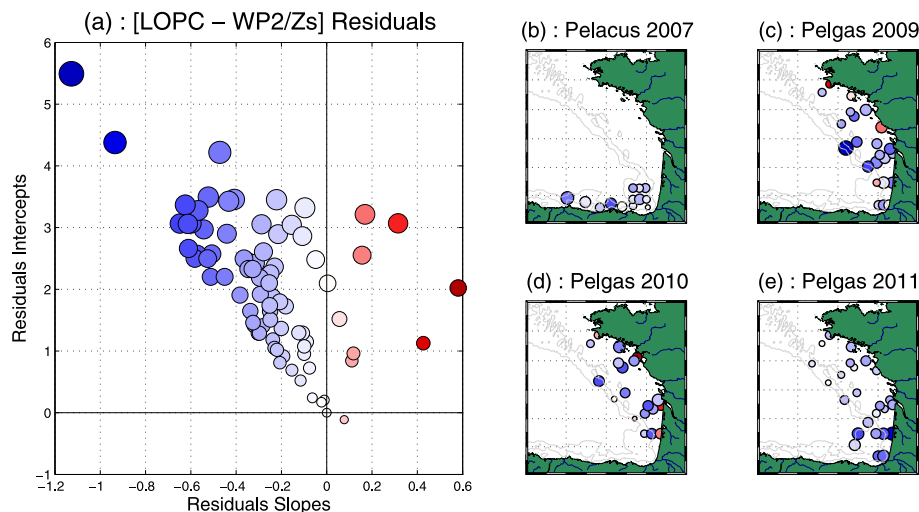


(c)

**Fig. 3.** Examples of simultaneous Normalized Biomass Size Spectra (NBSS) measured by the LOPC and by the WP2/Zs. For the WP2/Zs the NBSS of all objects, all zooplankton, all crustaceans and copepods only are shown. The two vertical dash lines indicate the limits of comparable size classes of the LOPC and the WP2/Zs. Location of station is shown as insets of each graph. (a) shows an example where the LOPC overestimates the WP2/Zs for all size classes, (b) where the overestimation is only for smaller size classes and (c) where there is no overestimation.

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**Fig. 4.** Size structure characteristics of residuals between NBSS estimated by the LOPC and by the WP2/Zs procedure. **(a)** shows the relation between the slope and the intercept of residuals. the colormap indicates the value of the slope of the residuals (blue for negative slope, red for positive ones and white for null slopes). The size of points indicates the value of the intercept of residuals (the bigger the points, the larger the intercepts). **(b–e)** show the spatial distribution of residuals for the four cruises in which we have data for both LOPC and WP2/Zs (Pelagus 2007 **(b)**, Pelgas 2009 **(c)**, Pelgas 2010 **(d)** and **(e)** Pelgas 2011). The color are the same as in **(a)** but for clarity size of points have been divided by three.

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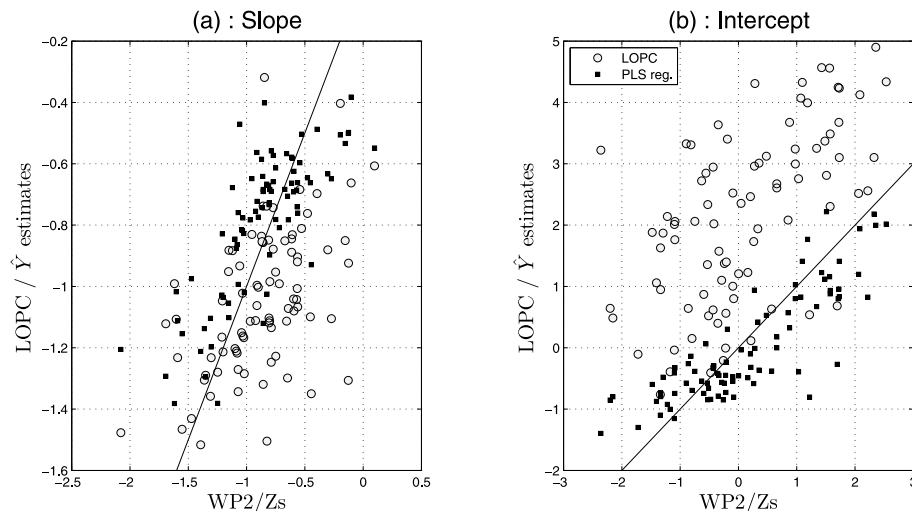
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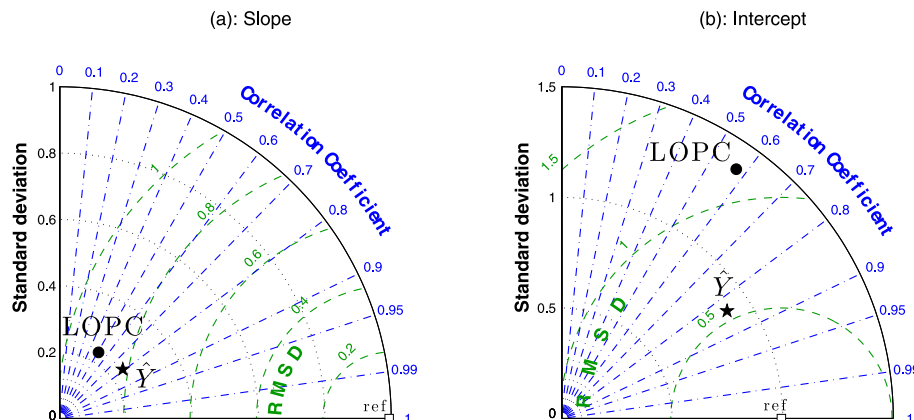


**Fig. 5.** Scatter graph of the relations of NBSS slopes **(a)** and intercepts **(b)** estimated by the WP2/Zs vs. the raw LOPC (grey points), and after the PLS (Partial Least Square) regression was performed on the LOPC ( $\hat{Y}$ , black plain square). The  $x = y$  line is added to each plot.

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**Fig. 6.** Taylor diagram (Taylor, 2001) of the correlation ( $R$ ), the centered root mean square difference (RMSD) and standard deviation (STD) between NBSS parameters (**a**: slope and **b**: intercept) estimated by the WP2/Zs and estimated by the LOPC (black plain circles) and after the PLS (Partial Least Square) regression performed on the LOPC ( $\hat{Y}$ , black plain stars). The white square indicates the maximum ( $R = 1$ , RMSD = 0 and STD = 1).

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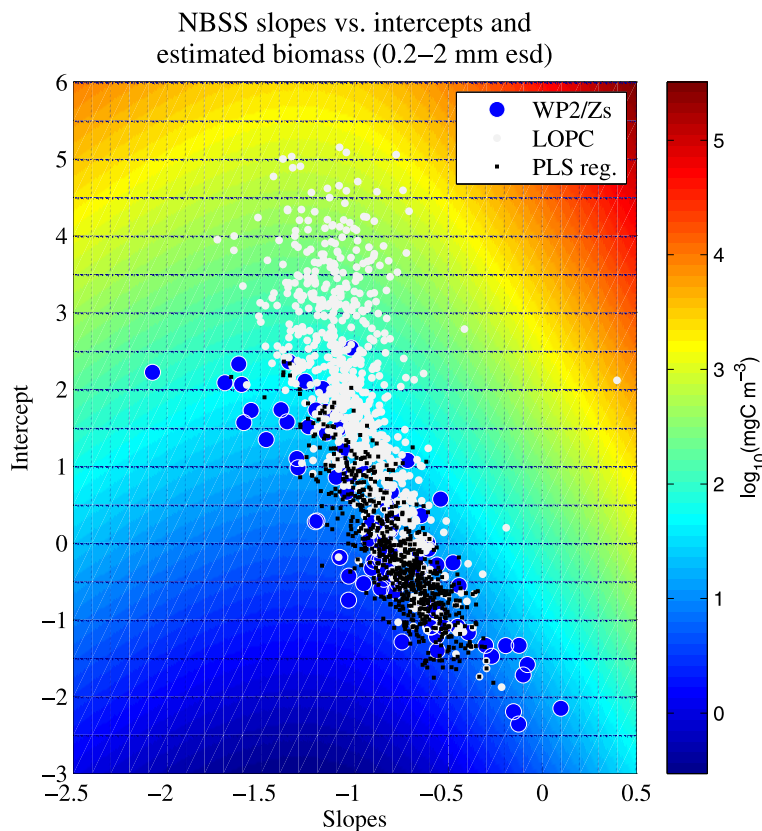
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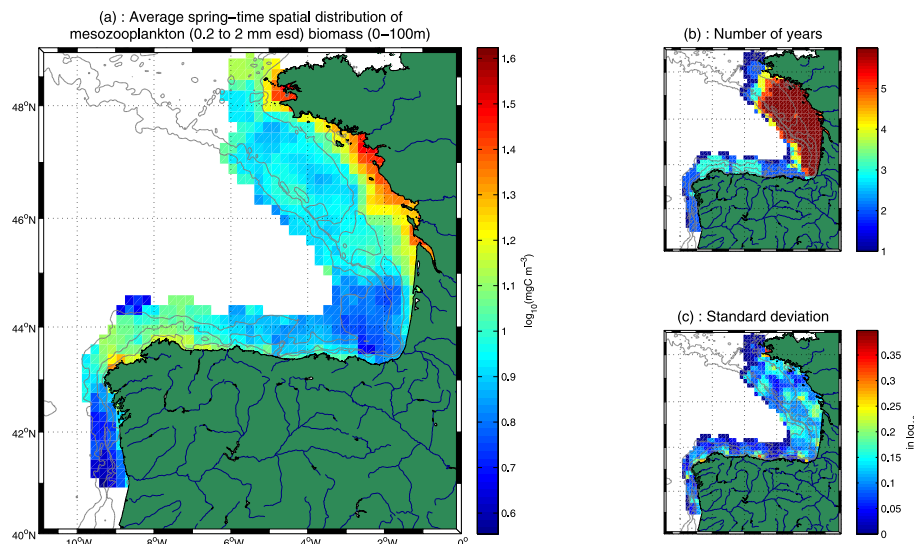




**Fig. 7.** Scatter graph of the relations between slopes and intercepts computed from NBSS estimated by the WP2/Zs (blue circles), the LOPC (grey circles) and by the PLS (Partial Least Square) regression (black square). The colormap corresponds to the biomass computed as the integration of the log-linear NBSS in the range 0.2–2 mm ESD.

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**Fig. 8.** Spatial distribution of mesozooplankton (from 0.2 to 2 mm ESD) biomass concentration (in  $\log_{10} \text{ mg C m}^{-3}$ ) from the surface to a maximum of 100 m depth and calculated as the integral of the NBSS estimated by the PLS (Partial Least Square) regression. A regular grid was generated each year and nodes were interpolated from available data within a radius of 30 km and weighted by the square of their distance. **(a)** shows the average distribution, **(b)** the total number of year used to calculate each points and **(c)** shows the standard deviation.

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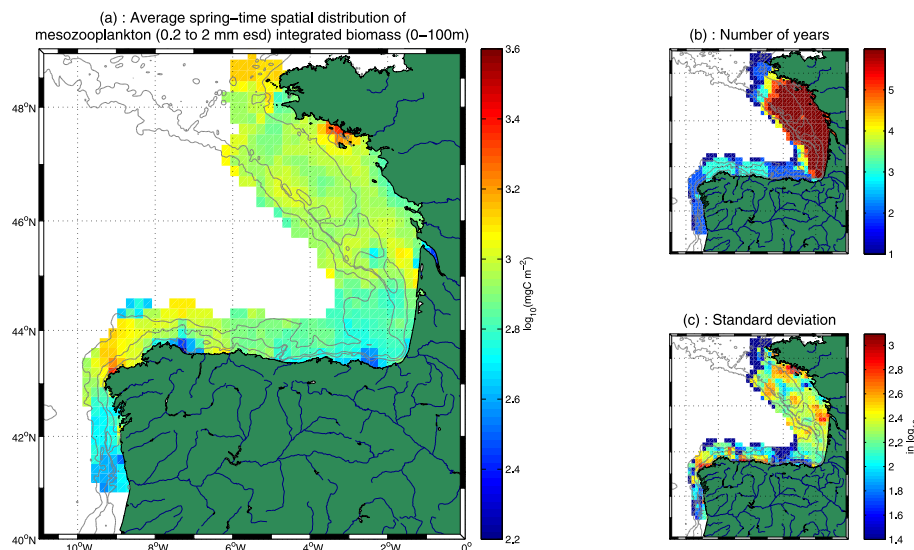
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**Fig. 9.** Spatial distribution of mesozooplankton (from 0.2 to 2 mm ESD) integrated biomass (in  $\log_{10} \text{mg C m}^{-2}$ ) from the surface to a maximum of 100 m depth and calculated as the integral of the NBSS estimated by the PLS (Partial Least Square) regression. A regular grid was generated each year and nodes were interpolated from available data within a radius of 30 km and weighted by the square of their distance. **(a)** shows the average distribution, **(b)** the total number of year used to calculate each points and **(c)** shows the standard deviation.

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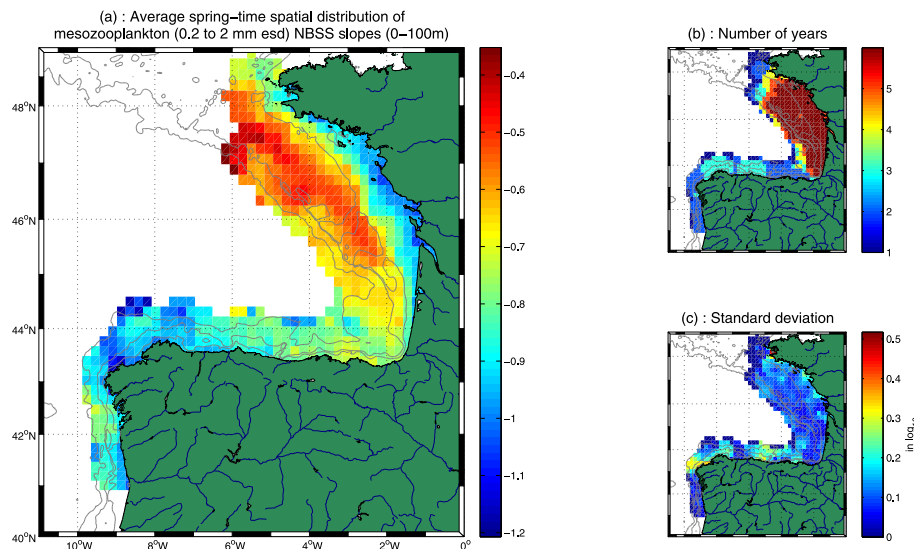
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**Fig. 10.** Spatial distribution of NBSS slopes estimated by the PLS (Partial Least Square) regression. A regular grid was generated each year and nodes were interpolated from available data within a radius of 30 km and ponderated by the square of their distance. (a) shows the average distribution, (b) the total number of year used to calculate each points and (c) shows the standard deviation.

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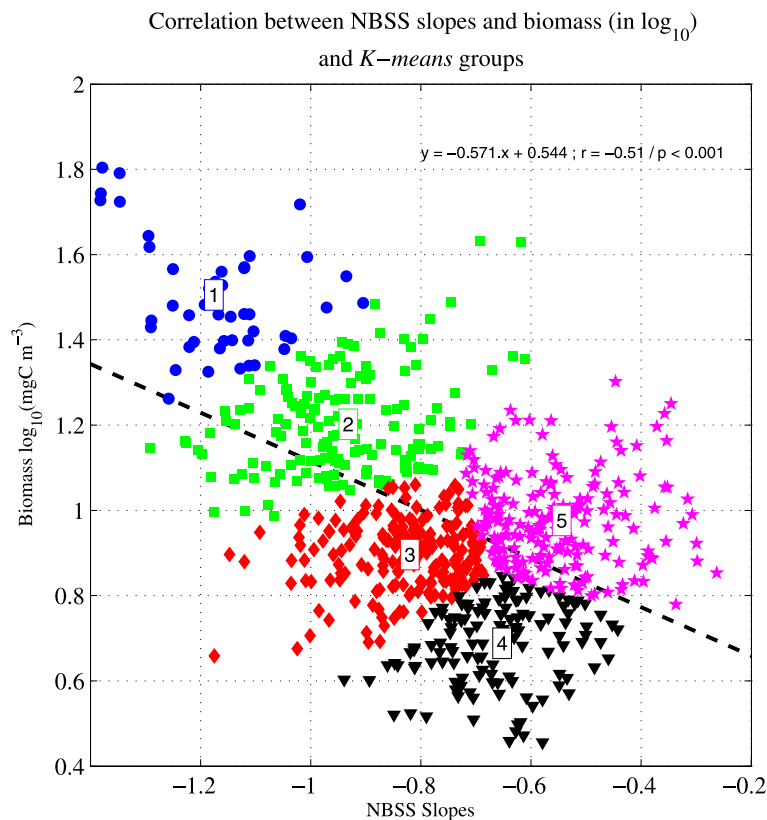
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**Fig. 11.** Correlation between slopes and the biomass computed as the integration of the log-linear NBSS in the range 0.2–2 mm ESD. The correlation (Pearson) is significant with a  $r = -0.51$ . Result of a 5 groups *K* means clustering based on estimated slope and biomass is shown with different colors and shapes. Group 1: blue circles, group 2: green squares, group 3: red losanges, group 4: black triangles and group 5: pink stars.

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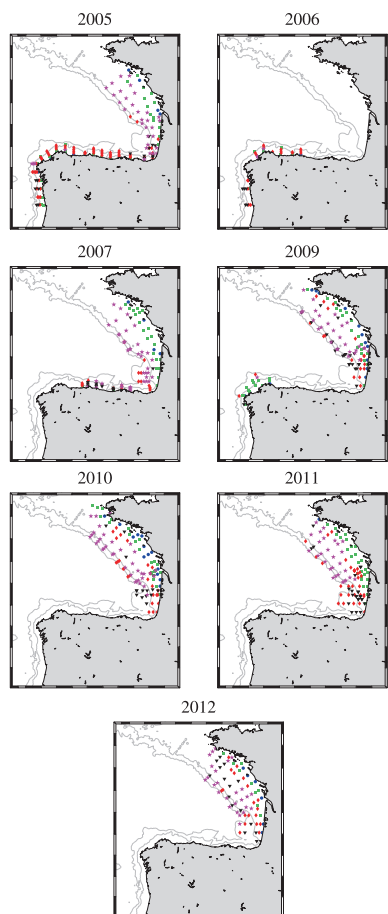
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**Fig. 12.** Spatial and interannual distribution of  $K$  means groups as established in Fig. 11 for the seven years. Groups are represented by the same colors and shapes as in Fig. 11.

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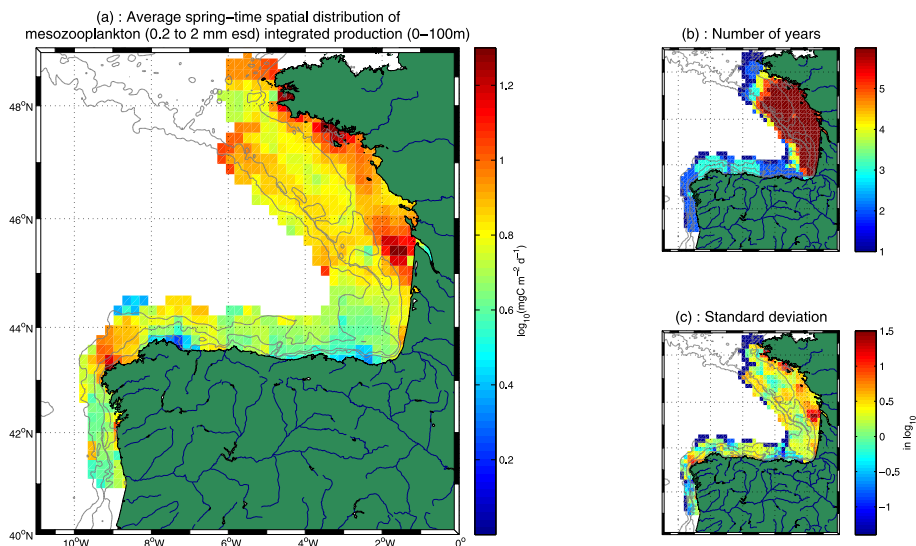
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**Fig. 13.** Spatial distribution of mesozooplankton (from 0.2 to 2 mm ESD) productivity (in  $\log_{10} \text{mg C m}^{-2} \text{ d}^{-1}$ ) calculated from the NBSS estimated by the PLS (Partial Least Square) regression (Sect. 2.4). A regular grid was generated each year and nodes were interpolated from available data within a radius of 30 km and ponderated by the square of their distance. **(a)** shows the average distribution, **(b)** the total number of year used to calculate each points and **(c)** shows the standard deviation.

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