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Alkalinity sources in the Dutch Wadden Sea

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Abstract. Total alkalinity (TA) is an important chemical property that plays a decisive role in the oceanic buffering capacity with respect to CO₂. TA is mainly generated by weathering on land as well as by various anaerobic metabolic processes in the water and sediments. The Wadden Sea, located in the southern North Sea, is hypothesized to be a source of TA for the North Sea, but quantifications are scarce. This study shows observations of TA, dissolved inorganic carbon (DIC), and nutrients in the Dutch Wadden Sea in May 2019. Surface samples were taken along several transects in order to investigate spatial distribution patterns and compare them with data from the late 1980s. A tidal cycle was sampled to further shed light on TA generation and potential TA sources. We identified the Dutch Wadden Sea as a source of TA and estimated an export of 6.6 Mmol TA per tide to the North Sea. TA was generated in the sediments, with deep pore water flow during low tide enriching the surface water. A combination of anaerobic processes and CaCO₃ dissolution were potential TA sources in the sediments. We deduce that seasonality and the associated nitrate availability specifically influence TA generation by denitrification, which is low in spring and summer.

1 Introduction

As the regulator of the ocean carbon dioxide (CO₂) sink, total alkalinity (TA) is of increasing scientific interest and has been investigated worldwide in the so-called "Anthropocene" (Abril and Frankignoulle, 2001; Bozec et al., 2005;

Chen and Wang, 1999; Dickson, 1981; Middelburg et al., 2020; Norbisrath et al., 2022; Renforth and Henderson, 2017; Thomas et al., 2004, 2009; Sabine et al., 2004). The Anthropocene describes the current era of our planet, during which environmental changes, driven by humans, have become identifiable in geological records (Zalasiewicz et al., 2010; Crutzen, 2002). One of the most threatening changes with respect to our climate is the anthropogenic increase in atmospheric greenhouse gases (GHGs), such as CO₂. To counteract the increasing atmospheric CO₂ concentrations and ongoing climate warming, a combination of several pathways is needed. In addition to a strict reduction in CO₂ emissions, net-negative emissions are also required; to achieve the latter, atmospheric CO_2 must be captured and stored, either on land or in the ocean (e.g., Keith et al., 2006; Matthews and Caldeira, 2008; Zhang et al., 2022). The climate and the increasing atmospheric CO₂ content is also naturally regulated by the open ocean, and around a quarter of the global anthropogenic CO₂ emissions are already removed by it (Friedlingstein et al., 2022). The carbon storage capacity of the North Sea is an important atmospheric CO_2 sink, as it exports the absorbed CO₂ to the deep layers of the Atlantic Ocean where it is then stored on longer timescales (Borges et al., 2005; Bozec et al., 2005; Burt et al., 2016; Brenner et al., 2016; Hu and Cai, 2011; Schwichtenberg et al., 2020; Thomas et al., 2004, 2009). Two important aspects of oceanic climate regulation are the oceanic circulation and TA. TA, primarily consisting of bicarbonate and carbonate, is generated by chemical rock weathering (Suchet and Probst, 1993; Meybeck, 1987; Berner et al., 1983) as well as in various stoichiometries by calcium carbonate (CaCO₃) dissolution and anaerobic metabolic processes, such as denitrification, which is the reduction process of nitrate to dinitrogen gas in the nitrogen cycle (Hu and Cai, 2011; Wolf-Gladrow et al., 2007; Chen and Wang, 1999; Brewer and Goldman, 1976). As TA, CO_2 uptake, and CO_2 export to the deep ocean are mainly disentangled in the open ocean, TA and the oceanic circulation interact closely in highly active and shallow-ocean areas, such coastal zones and continental and marginal shelves.

In these shallow areas, TA is susceptible to changes due to various metabolic processes and the influence of adjacent zones like rivers, estuaries, marshes, and tidal flats (e.g., Norbisrath et al., 2022, 2023; Wang et al., 2016; Voynova et al., 2019). A previous study by Norbisrath et al. (2022) showed that an enhanced riverine metabolic alkalinity would lead to increasing CO₂ absorption in the coastal zones of the North Sea, highlighting the need to further investigate TA regulation in adjacent zones of coastal oceans.

Coastal zones, which are the direct interface between most (if not all) compartments of the Earth system (i.e., atmospheric, terrestrial, aquatic, and oceanic) and human societies, appear particularly vulnerable to environmental and climate change (Glavovic et al., 2015). This holds true for the Wadden Sea, the shallow coastal sea along an approximately 500 km coastline of the Netherlands, Germany, and Denmark, in the southern North Sea, which has been declared a UNESCO world natural heritage site since 2009. Most of the Wadden Sea is located between protective barrier islands and the mainland, making it the world's largest uninterrupted stretch of tidal flats with multiple tidal inlets (Fig. 1). Due to the region's topography, the Wadden Sea is a highly dynamic ecosystem with influences from the mainland and the North Sea (Hoppema, 1993; Postma, 1954; van Raaphorst and van der Veer, 1990). Driving forces of the biogeochemical dynamics in the Wadden Sea are nutrient imports by rivers and high suspended particulate matter (SPM) and organic matter (OM) imports from the North Sea (van Beusekom et al., 2019, 2012; Postma, 1954). Physical sources of variability in the Wadden Sea are ocean-driven wind, waves, and tidal currents as well as the counterclockwise circulation of the North Sea (Elias et al., 2012). A large tidal amplitude and currents in conjunction with shallow water depths allow for vertical water column mixing and an exchange between the pelagic and benthic realms, including deep pore water exchange (Røy et al., 2008). The strong tidal currents also impact the biogeochemistry of the North Sea (Postma, 1954), as they cause an exchange of water between the North Sea and the Wadden Sea and play an important role in the import of particulate matter from the North Sea (Burchard et al., 2008).

Previous studies have identified the Wadden Sea as a TA source for the North Sea, with a loading of between 39 Gmol yr⁻¹ (Schwichtenberg et al., 2020) and 73 Gmol yr⁻¹ (Thomas et al., 2009). Both of these prior publications suggested that the entire Wadden Sea is one of the most important TA sources impacting the carbon storage capacity of the North Sea. Burt et al. (2016) highlighted the importance of coastal TA production with respect to regulating the buffer system in the North Sea and suggested denitrification as the major TA source. Due to the strong connection between the North Sea and the Wadden Sea, a better understanding of TA generation in the latter is required. Here, we focus on the Dutch Wadden Sea, which has been well-studied during the past decades (Hoppema, 1990, 1991, 1993; De Jonge et al., 1993; Elias et al., 2012; Ridderinkhof et al., 1990; Postma, 1954; van Beusekom et al., 2019; Schwichtenberg et al., 2017). In particular, in the late 1980s, Hoppema (1990, 1993) observed the spatiotemporal variability in TA in the month of May; we compare those findings with our observed transect data to detect potential differences over the last 30 years. In addition, we further discussed potential TA sources in the Dutch Wadden Sea.

2 Methods

2.1 Study site and sampling

This study is based on samples collected on a research cruise (no. LP20190515) on RV Ludwig Prandtl in the Dutch Wadden Sea (Frisian Islands) in May 2019 (Fig. 1). We collected water samples in the Wadden Sea starting at Harlingen and traveling along the following path: through the Vlie Inlet along the islands of Vlieland and Terschelling, through the Ameland Inlet to Ameland Island, via the Frisian Inlet to Lauwersoog, and around Schiermonnikoog Island via the Ems-Dollard Inlet to Emden. In addition, we sampled half a tidal cycle during ebb tide (from high tide to low tide) on 21 May 2019 (Table B2). To set the range of ebb tide data, we also sampled half a tidal cycle during flood tide (from low tide to high tide) on 23 May 2019 for comparison. Both of the half tidal cycles were sampled each day using an anchor station in the waterway on the western side of Ameland in the Ameland Inlet.

Nearly every 30 min, we collected discrete surface (1.2 m depth) water samples with a bypass from the onboard flowthrough FerryBox system, which also provided essential physical parameters, such as salinity, with an accuracy of 0.02 PSU (practical salinity units) and temperature with an accuracy of 0.1 °C (Petersen et al., 2011). The FerryBox was cleaned and checked prior to the cruise, and salinity was occasionally checked using discrete samples; this workflow is considered sufficient for gradients in near-shore investigations (Yoana Voynova, personal communication, 2024). We complemented our salinity and temperature data with data from three Rijkswaterstaat stations (Dantziggat, Terschelling 10, and Vliestroom; Table B3), which were close to our stations.

For TA and dissolved inorganic carbon (DIC) measurements, we sampled water with overflow into 300 mL BOD (biological oxygen demand) bottles and preserved it with $300 \,\mu$ L of saturated mercury chloride solution (HgCl₂) to



Figure 1. Sampling site in the Dutch Wadden Sea. The sampling stations around the Frisian Islands in May 2019 are visualized using white triangles. The yellow circle highlights the anchor stations used to sample the tidal cycle in the Ameland Inlet on 2 d. During the sampling day, from low tide to high tide, we had two samples that were taken slightly more to the west due to drift. The island and city names are shown in white, whereas the inlets are shown in yellow. The tidal flats and sedimentary structures are visible between the barrier islands and the mainland.

stop biological activity. Each BOD bottle was filled without air bubbles and closed using a ground-glass stopper coated in Apiezon[®] M grease and a plastic cap. The samples were stored in a cool, dark environment prior to analysis in the lab.

Water for nutrient samples was filtered through precombusted (4 h at 450 °C) GF/F filters, and the filtrate was stored frozen in three 15 mL Falcon[®] tubes for triplicate measurements in the lab.

To determine the total carbon (C), organic carbon (C_{org}) and nitrogen (N) concentrations in SPM, and associated C_{org} : N ratios, we used pre-combusted (4 h at 450 °C) GF/F filters; the filters were dried after sampling at 50 °C to remove all humidity and then stored frozen until measurement.

2.2 Carbon species analyses

The parallel analyses of TA and DIC were carried out in March 2020 using the MARIANDA (MARIne ANalytics and Data) VINDTA 3C (Versatile INstrument for the Determination of Total dissolved inorganic carbon and Alkalinity) instrument, which measures TA (via potentiometric titration) and DIC (via coulometric titration), both with a measurement precision of $< 2 \,\mu$ mol kg⁻¹ (Shadwick et al., 2011). Certified reference material (CRM batch no. 187), provided by Andrew G. Dickson (Scripps Institution of Oceanography), was measured before and after the samples and used to ensure a consistent calibration of both measurements.

The calcite and aragonite saturation states (Ω), the pH, and the seawater partial pressure of CO₂ (*p*CO₂) were computed

with the CO2SYS program (Lewis and Wallace, 1998), using the measured parameters TA and DIC, salinity, temperature, silicate, and phosphate as input variables as well as the dissociation constants from Mehrbach et al. (1973), as refitted by Dickson and Millero (1987). Reported calculation uncertainties are ± 0.0062 for pH (Millero et al., 1993), ± 4.9 % for the aragonite saturation state, and ± 3.5 % for pCO_2 (Orr et al., 2018).

2.3 Nutrient analyses

The nutrients were measured with a continuous-flow automated nutrient analyzer (AA3, SEAL Analytical) and a standard colorimetric technique (Hansen and Koroleff, 2007) for nitrate (NO₃⁻), nitrite (NO₂⁻), phosphate (PO₄³⁻), and silicate (Si), whereas a fluorometric method (Kérouel and Aminot, 1997) was employed for ammonium (NH₄⁺) (Grasshoff et al., 2009). The nutrient samples were measured against Eurofins Scientific reference materials VKI SW4.1B (for NO_x, NO₂, and NH₄) and VKI SW4.2B (for Si and PO₄) in July 2019. The maximum standard deviations were 0.322 µmol L⁻¹ for NO₃⁻, 0.014 µmol L⁻¹ for NO₂⁻, 0.081 µmol L⁻¹ for NH₄⁺, 0.014 µmol L⁻¹ for PO₄³⁻, and 0.165 µmol L⁻¹ for Si.

For C_{org} determination, filters were acidified with 1N HCl and dried overnight to remove all inorganic carbon content. Filters were measured with a CHN (carbon-hydrogen-nitrogen) elemental analyzer (Eurovector EA 3000, HEKAt-ech GmbH) at the Institute of Geology, University of Ham-

burg, and calibrated against a certified acetanilide standard (IVA Analysentechnik). The standard deviations were 0.05 % for carbon and 0.005 % for nitrogen.

2.4 Data analyses

The data analyses were performed using RStudio (Version 1.3.1073, © 2009–2020 RStudio, Posit PBC; R Core Team, 2019). The linear regression Model II was performed using the "Imodel2" R package (Legendre, 2018), and the plots were created with the "ggplot2" R package (Wickham, 2016).

3 Results

3.1 Spatial parameter distribution

To investigate the spatial distribution of TA in the Dutch Wadden Sea and compare its general status with earlier studies (in particular Hoppema, 1990), we observed TA and related parameters in surface water along a transect from the coastal mainland towards the North Sea.

The temperature varied between 12 and 16 °C, with higher temperatures towards the coastal mainland (Fig. 2a). We identified two main subregions based on the salinity values: (1) the Ems-Dollard Inlet, which showed salinity values lower than 28 PSU and had a minimum value of 20.25 PSU at the most upstream station, and (2) the area around Ameland Island and the remainder of our investigated region in the Dutch Wadden Sea, which showed salinity values from 28 to 33 PSU with little variation (Fig. 2b).

The spatial transect of TA concentrations ranged from 2332 to 2517 μ molTA kg⁻¹. We observed lower concentrations on the North Sea side of the Frisian Islands, with somewhat higher concentrations around Ameland (Fig. 2c). In contrast to the North Sea side, the values were higher (> 2380 μ molTA kg⁻¹) in the Wadden Sea. In the Ems-Dollard Inlet, the concentrations were even higher, with values of up to 2517 μ molTA kg⁻¹ at the most upstream station.

Silicate (Si) showed higher concentrations in the Wadden Sea and lower values towards the North Sea (Fig. A1a). The highest concentrations were observed at the coastal mainland and in the Ems-Dollard Inlet. Silicate concentrations ranged between 0.3 and 56.3 μ mol Si L⁻¹. Both the calcite and aragonite saturation states (Ω) were supersaturated throughout the study region. The Ω values ranged from 2.3 to 4.6 for calcite (Fig. A1b) and from 1.4 to 2.8 for aragonite (Table B3). The highest values were observed on the North Sea side of the barrier islands, whereas the lowest values were found near Harlingen and in the Ems-Dollard Inlet. Like the calcite and aragonite saturation states, the pH values were higher in the North Sea, whereas they were lower in the Wadden Sea and near the coastal mainland (Fig. A1c). The pH values ranged from 7.86 to 8.19, with the lowest readings observed near Harlingen and in the Ems-Dollard Inlet. The nitrate (NO₃⁻) concentrations fell within in a low range (< $3 \mu mol NO_3^- L^{-1}$) throughout the study region. Higher concentrations (< $6 \mu mol NO_3^- L^{-1}$) were only observed at a few stations close to land, and the maximum concentrations (< $38 \mu mol NO_3^- L^{-1}$) were observed in the Ems-Dollard Inlet (Fig. A1d). DIC concentrations ranged from 2097 to 2430 µmol DIC kg⁻¹ (Fig. A1e). DIC values showed a similar pattern to TA values: higher concentrations near the coastal mainland and in the Ems-Dollard Inlet and decreasing concentrations toward the North Sea, where DIC reached minimum values.

Compared with the other transects of this study region, the strong influence of the inner Ems Estuary is visible at the most upstream station in the Ems-Dollard Inlet, showing the lowest salinity values; the lowest pH and calcite saturation state values; and the highest TA, DIC, nitrate, silicate, and phosphate values. The outer side of the Vlie Inlet reflects the North Sea conditions: lower temperatures and higher salinities. The North Sea impact is also visible in the mixing plot between TA and salinity (Fig. 3). Statistically significant linear mixing behavior was observed in the transect through the Ems-Dollard Inlet ($R^2 = 0.81$) and through the Vlie Inlet $(R^2 = 0.77)$, where TA concentrations decreased with increasing salinities from the mainland towards the North Sea (Fig. 3). Whereas in the Ems-Dollard Inlet mixing is dominated by riverine water with high TA concentrations, the mixing in the Vlie Inlet showed a more prominent mixing of Wadden Sea and North Sea water. The TA concentrations in the Vlie Inlet and around Ameland, both on the North Sea side (Ameland NS) and the Wadden Sea side (Ameland WS), were higher than the TA concentration computed for the salinity end-member in the Ems-Dollard Inlet, suggesting the Dutch Wadden Sea as a source of TA (Fig. 3). Both Ameland NS and Ameland WS data clearly indicated nonconservative behavior with a range of TA concentrations at near-constant salinities.

3.2 Tidal cycle

In order to (1) identify potential TA sources and (2) to quantify potential TA export to the North Sea, we observed half a tidal cycle at an anchor station in the Ameland Inlet during ebb tide. We identified patterns in several biogeochemical parameters in water leaving the tidal flats (Fig. 4, Table B1). Temperature increased from 13.25 to 14.7 °C (Fig. 4a). Salinity was constant at around 32.5 PSU (Fig. 4b), which is within the range of coastal southern North Sea water excluding admixture of local freshwater sources.

During ebb tide, TA ranged from $2387 \,\mu\text{mol}\,\text{TA}\,\text{kg}^{-1}$ during high tide to $2438 \,\mu\text{mol}\,\text{TA}\,\text{kg}^{-1}$ during low tide (Fig. 4c). We observed an increase of $51.6 \,\mu\text{mol}\,\text{TA}\,\text{kg}^{-1}$ (Δ TA) during ebb tide (6.8 h), resulting in a TA increase of 7.6 $\mu\text{mol}\,\text{TA}\,\text{kg}^{-1}\,\text{h}^{-1}$ at the sampling location.

DIC concentrations behaved similarly to TA, with minimum values at high tide $(2172 \,\mu mol \, DIC \, kg^{-1})$ and max-



Figure 2. Spatial distribution of (a) temperature (°C), (b) salinity (PSU), and (c) total alkalinity (TA; μ mol kg⁻¹) from surface water samples in May 2019.

imum values $(2273 \,\mu\text{mol}\,\text{DIC}\,\text{kg}^{-1})$ at low tide, resulting in an increase of $101.3 \,\mu\text{mol}\,\text{DIC}\,\text{kg}^{-1}$ (Δ DIC), or $14.9 \,\mu\text{mol}\,\text{DIC}\,\text{kg}^{-1}\,\text{h}^{-1}$ (Fig. 4d). DIC increased almost twice as much as TA.

Nitrate increased by $0.92 \,\mu mol NO_3^- L^{-1} (\Delta NO_3^-)$ during ebb tide, from a minimum of $1.26 \,\mu mol NO_3^- L^{-1}$ to a maximum of $2.17 \,\mu mol NO_3^- L^{-1}$ (Fig. 4e), resulting in a nitrate increase of $0.13 \,\mu mol NO_3^- L^{-1} h^{-1}$.

Silicate showed a similar pattern, with low values $(1.8 \,\mu\text{mol}\,\text{Si}\,\text{L}^{-1})$ at high tide that increased during ebb tide to a maximum of $11.2 \,\mu\text{mol}\,\text{Si}\,\text{L}^{-1}$, resulting in a silicate increase (Δ Si) of $9.4 \,\mu\text{mol}\,\text{Si}\,\text{L}^{-1}$, or $1.4 \,\mu\text{mol}\,\text{Si}\,\text{L}^{-1}\,\text{h}^{-1}$, during ebb tide (Fig. 4f).

Ammonium increased from 3.47 to $6.22 \,\mu mol \, NH_4^+ L^{-1}$ during ebb tide (Fig. 4g), resulting in an ammonium increase (ΔNH_4^+) of 2.74 $\mu mol \, NH_4^+ L^{-1}$, or 0.4 $\mu mol \, NH_4^+ L^{-1} h^{-1}$. The calcite and aragonite saturation states had maximum values ($\Omega_{Ca} = 3.8$, $\Omega_{Ar} = 2.4$) at high tide and decreased to their minimum ($\Omega_{Ca} = 3.1$, $\Omega_{Ar} = 2.0$) during ebb tide (Fig. 4h). The influence of the North Sea is indicated by the observed maximum at high tide, which decreased during the ebb.

The seawater pCO_2 had minimum values at high tide (385.1 µatm) and increased up to 576.6 µatm during low tide (Fig. 4i).

Like Ω , the maximum pH was 8.07 at high tide and decreased to a minimum (7.93) during ebb tide (Fig. 4j).

The C_{org} : N ratios of SPM increased during ebb tide (Fig. 4k). A minimum C_{org} : N ratio of 5.6 was observed around high tide and increased to a maximum of 13.0 during ebb tide. Simultaneously, the SPM concentration increased during ebb tide, from 12.8 mg SPML⁻¹ to a maximum of 82.4 mg SPML⁻¹ at the second-last station (Table B1).



Figure 3. Mixing plot of total alkalinity (TA) and salinity (PSU) on the North Sea side of Ameland and the Frisian Inlet (Ameland NS), on the Wadden Sea side of Ameland (Ameland WS), around Schiermonnikoog and in the Ems-Dollard Inlet (Ems-Dollard), and in the Vlie Inlet (Vlie).

3.3 TA generation

Tidal forcing leads to an exchange between Wadden Sea and North Sea water almost twice a day. The tidal forcing also induces a strong benthic–pelagic coupling (Huettel et al., 2003; Røy et al., 2008). Many studies support the fact that outflowing water exports material from the sediment, including remineralization products from organic matter (e.g., Billerbeck et al., 2006; Røy et al., 2008). Here, we focus on the hypothesis that the sediments are a significant source of TA.

For an initial rough estimate of maximum TA export during ebb tide, we used the mean observed TA increase $(\Delta TA/2)$ of 25.8 µmol TA kg⁻¹ during ebb tide (in the Ameland Inlet, part of the Borndiep tidal basin), a tidal prism of 478×10^6 m³ of the Borndiep tidal basin, and a share of intertidal flats of 53 % (Louters and Gerritsen, 1994). Assuming that only the intertidal sediments exchange TA, we estimated a TA export of 6.6 Mmol TA per tide to the North Sea. Assuming two ebb tides and a lunar cycle of 24.8 h, this would result in a daily export of 12.7 Mmol TA.

The significant correlation of TA and silicate ($R^2 = 0.93$) as well as the insignificant relation between TA and salinity ($R^2 = 0.32$) and between silicate and salinity ($R^2 = 0.21$) suggest that TA originates from the tidal flats in this part of the Dutch Wadden Sea, not from admixture carried by river runoff. The significant correlation between TA and silicate during ebb tide point to the same source (Fig. 5b).

To further elucidate potential TA sources in the Dutch Wadden Sea, we correlated TA with DIC, silicate, nitrate, and ammonium, respectively, in half a tidal cycle, from high tide to low tide (Fig. 5).

The correlation between TA and DIC is a measure between anaerobic and aerobic processes. Our data show a strong positive correlation between DIC and TA ($R^2 = 0.93$), with TA concentrations being higher than DIC concentrations (Fig. 5a). We observed a release of excess DIC compared with TA, as indicated by the slope of 1.89 and by an increase in DIC (Δ DIC = 101.3 µmol kg⁻¹) that is almost twice as high as TA (Δ TA = 51.6 µmol kg⁻¹) (Fig. 5a). This excess DIC may be caused by strong CO₂ production due to high aerobic OM degradation, which can be supported by the seawater pCO_2 being supersaturated with respect to the atmosphere (Fig. 4i). The TA increase can be fueled by various processes, which we will discuss below. We detected a positive linear correlation of increasing TA and silicate $(R^2 = 0.93)$ during ebb tide, supporting pore water outflow (Fig. 5b), as pore water is the major Si source during summer (van Bennekom et al., 1974). A stronger influence of pore water with the ongoing ebb tide is indicated by increasing values. The positive correlation between nitrate and TA $(R^2 = 0.67)$ (Fig. 5c) was less strong than the correlations between TA and DIC and between TA and Si, which could be traced back to an effect of the first four sampling points that were probably at the tipping point from high tide to low tide. In the remaining samples, the increasing nitrate and TA concentrations suggest stronger TA generation than nitrate production, balancing TA that may be consumed by nitrification (i.e., nitrate production).

4 Discussion

4.1 TA spatial variability

Hoppema (1990) reported TA distributions in the westernmost part of the Dutch Wadden Sea, around the barrier islands of Texel, Vlieland, and Terschelling. The aforementioned work focused on the tidal basins drained by the tidal inlets of Marsdiep and Vlie, located more to the west than our sampling stations (not visible on the map). Hoppema (1990) did not observe a continuous increase in salinity in the Wadden Sea from the freshwater source towards the North Sea and associated this with the influence of tidal differences and an arbitrary sampling scheme. The presence (dominance) of North Sea water in the Dutch Wadden Sea and on the tidal flats is supported by our transect data, which show relatively high salinities at the coastal North Sea level. Brackish salinity values were only detected in the Ems-Dollard Inlet, which receives freshwater from the river Ems, and close to Harlingen and Lauwersoog, which have direct freshwater inflows from smaller rivers and streams. The absence of clear salinity gradients in the more eastern part of the Dutch Wadden Sea investigated in our study suggests that most of the IJsselmeer discharge was exchanged with the North Sea through the Marsdiep Inlet (e.g., Duran-Matute et al., 2014).

The spatial TA data from Hoppema (1990) show lower TA concentrations at stations with more freshwater influence and higher TA concentrations in the tidal inlets. The data from this study also show high TA concentrations in the tidal inlets, suggesting TA generation in sediments, which



Figure 4. Half a tidal cycle (from high tide to low tide) showing the temporal distribution of (**a**) temperature, (**b**) salinity, (**c**) total alkalinity (TA), (**d**) dissolved inorganic carbon (DIC), (**e**) nitrate, (**f**) silicate, (**g**) ammonium, (**h**) calcite (upper points) and aragonite (lower points) saturation states, (**i**) pCO_2 , (**j**) pH, (**k**) the C_{org} : N ratio of SPM, and (**l**) the DIC : TA ratio. Note the different *y* axes and the +2 h time difference between the local summer time and UTC time.

is possibly fueled by high nutrient and OM imports (van Beusekom and De Jonge, 2002). The even higher TA concentrations at stations with lower salinities close to the mainland observed in this study also show the influence of the catchment area on the coast and, possibly, TA generation in the shallow sediments near the coast. In May 1986, Hoppema (1990) found TA concentrations ranging between 2319 and 2444 μ mol TA kg⁻¹ at salinities between 18.62 and 29.17 PSU. Our lowest observed TA concentration was 2332 μ mol TA kg⁻¹ at a salinity of 32.14 PSU, whereas our highest TA concentration was 2517 μ mol TA kg⁻¹ at a salinity of 20.25 PSU close to the coastal mainland. A comparison of both studies shows that the general TA levels are in a similar range but that the spatial gradients are opposite.



Figure 5. Correlations of TA with (a) dissolved inorganic carbon (DIC), (b) silicate, (c) nitrate, and (d) ammonium during ebb tide in the Ameland Inlet.

Conservative mixing between TA and salinity is only visible in the Ems-Dollard Inlet and the Vlie Inlet (Fig. 3). While conservative mixing in the Ems-Dollard Inlet is more dominated by the freshwater discharge from the river Ems, the conservative mixing in the Vlie Inlet is more dominated by North Sea water passing through this deep inlet and allowing more North Sea water to be transported towards the coast. After the Marsdiep Inlet, the Vlie Inlet has the highest average tidal prism and is the second-largest inlet in the Dutch Wadden Sea (Elias et al., 2012). Similar to our findings, Hoppema (1990) noted linear mixing of TA and salinity in the Vlie Inlet and also suspected a lower freshwater contribution in the inlet, which is in accordance with model data (Duran-Matute et al., 2014).

In the Ems-Dollard Inlet, conservative mixing was observed, indicating minor contributions from other sources. In a previous study, Norbisrath et al. (2023) observed very high TA concentrations and TA generation in the upper tidal river of the highly turbid Ems Estuary, which may explain the high levels of TA in the Ems-Dollard Inlet (at low salinities) observed in this study. Hoppema (1990) also observed a range of TA concentrations in the Dutch Wadden Sea and related these to different sinks and sources. TA sinks can be calcium carbonate (CaCO₃) precipitation or the extraction of seawater carbonate by mollusks (e.g., Chen and Wang, 1999; Hoppema, 1990). Variable freshwater inflows can either serve as a sink or a source (e.g., Chen and Wang, 1999; Hoppema, 1990). Other TA sources can be CaCO₃ dissolution, anaerobic metabolic processes in the sediment, or the erosion of sediments that enhance TA (e.g., Hoppema, 1990; Chen and Wang, 1999).

Except for the Ems-Dollard Inlet and close to Harlingen, we mainly observed marine salinities > 30 PSU but found higher TA values in the Dutch Wadden Sea than in the North Sea. Therefore, we exclude possible TA sinks and focus only on TA sources. According to Hoppema (1990), the main causes of TA variation in the Dutch Wadden Sea were freshwater inflows and sources in the sediment. In our study, freshwater inflows with high TA concentrations were only observed in the Ems-Dollard Inlet (but not around the islands and the tidal flats). For a further TA source identification in the Dutch Wadden Sea, we investigated the TA variability during ebb tide in a tidal channel close to Ameland.

4.2 Determination of TA generation

Burt et al. (2016) and Schwichtenberg et al. (2020) have indicated that TA generation in the Wadden Sea is an important source with respect to the North Sea's carbon storage capacity. Here, we want to further identify TA generation and potential TA sources.

In a study from the late 1980s, Hoppema (1993) observed a tidal cycle in the Marsdiep Inlet in May and September. Focusing on TA, DIC, and oxygen, this aforementioned work also observed increasing TA values during ebb tide and assumed that the tidal flats and discharging rivers and canals were TA sources. Comparing our present TA data to the historical TA data, there is not a large difference in the range of values observed during a tidal cycle. However, further indepth interpretation and comparison of both TA data sets is limited by the low number of data, leading us to focus on TA generation during our cruise.

We made a very rough first estimate of the daily TA export. Using a 3D ecosystem model, Schwichtenberg et al. (2020) estimated an annual export of 10-14 GmolTA yr⁻¹ for the entire Dutch Wadden Sea. Given that the Borndiep tidal basin covers about 14% of the Dutch Wadden Sea and assuming no seasonal dynamics, our estimate of 12.7 Mmol TA d^{-1} compares well with the annual average model estimate of 4.6 Mmol TA d^{-1} , but the overestimation suggests that seasonal dynamics may be involved. As our TA export was based on only a half-tidal-cycle observation, the inclusion of it into the model used by Schwichtenberg et al. (2020) would be unreliable (Johannes Pätsch, personal communication, 2022). To test whether the observed TA generation matches their suggested TA export, observational data of at least each season are required to run the model and gain a representative result (Johannes Pätsch, personal communication, 2022).

4.3 TA source attribution

4.3.1 Local sediment outwash

In order to gain further insight into potential sources of TA, we compared our TA and nutrient data. The main focus was on dissolved silicate (Si), as van Bennekom et al. (1974) showed that this nutrient is depleted in the Wadden Sea during the spring diatom blooms and further reported that pore water is the main source of dissolved silicate during summer. It is important to note that winter concentrations in the Rhine (the main contributor to the IJsselmeer) have not changed much since the 1970s, showing maximum concentrations of about $125 \,\mu\text{mol Si L}^{-1}$ in winter and clear seasonal dynamics due to uptake by diatoms (unpublished results based on data provided by Pätsch, 2024: available from https://wiki.

cen.uni-hamburg.de/ifm/ECOHAM/DATA_RIVER, last access: 30 July 2024). We identified a silicate increase of $1.4 \,\mu$ mol Si L⁻¹ h⁻¹ during ebb tide. Due to the absence of large estuaries nearby and salinity consistently being above 32 PSU at our tidal sampling station near the island of Ameland, we exclude freshwater runoff as a major silicate source and indicate TA sources within the Wadden Sea.

Submarine groundwater discharge (SGD) has been identified as a source for nutrient fluxes in tidal-flat ecosystems in previous studies (e.g., Billerbeck et al., 2006; Røy et al., 2008; Santos et al., 2021; Waska and Kim, 2011; Wu et al., 2013). As we observed relatively constant marine salinities, we suspect that deep pore water flow (e.g., Røy et al., 2008) which is enriched with respect to nutrients acts as a source for our observed increasing TA and nutrient parameters. TA generation in tidal flats was also observed by Faber et al. (2014), who focused on a large macro-tidal embayment in southern Australia. They also found increasing TA values during ebb tide, associated the TA increase with a higher fraction of pore water, and determined that the tidal cycle was the controlling force with respect to pore water exchange. Their findings and the observed silicate outwash support our assumption that TA is generated in the sediments of the tidal flats and is washed out during ebb tide. In addition, we exclude laterally advected signals from regions west of the Vlie Inlet, as the TA concentrations in the surface transect samples in the Vlie Inlet (except for the two samples close to the coastal mainland near Harlingen) were in the same range as the other observed TA concentrations and were lower than the increasing TA concentrations during ebb tide. Both increases in TA and silicate are tidal signals, and we identify TA generation in the sediments of the tidal flats here as the major local TA source.

4.3.2 TA generation processes

The observed TA increase of 7.6 μ mol TA kg⁻¹ h⁻¹ and the silicate increase of 1.4 μ mol Si L⁻¹ h⁻¹ indicated an excess of TA compared with silicate (also Fig. 5b). Considering a supposed TA : Si ratio of 2 : 1 (Marx et al., 2017), the observed 1.4 μ mol Si L⁻¹ h⁻¹ would then account for a TA generation of 2.8 μ mol TA kg⁻¹ h⁻¹. High silicate concentrations in tidal-flat pore water (Rutgers van der Loeff, 1974) and in situ production of silicate from dissolving diatom frustules are the most probable sources of the silicate (e.g., van Bennekom et al., 1974). As we observed more TA generated than silicate being washed out, other biogeochemical processes must be responsible for the TA generation in the sediments of the tidal flats in the Dutch Wadden Sea.

We exclude CaCO₃ dissolution as a TA source in the water column, as the Ω values were supersaturated with $\Omega > 1$ (Fig. 4h, Table B1). The continuous calcite supersaturation nicely indicated the inflow and dominance of North Sea water during the flood, with Ω values similar to previously observed North Sea values ($\Omega \sim 3.5$ –4) (Charalampopoulou et al., 2011; Carter et al., 2014). In pore water, carbonate undersaturation and associated $CaCO_3$ dissolution can only be driven metabolically, due to CO_2 production by OM remineralization or due to the reoxidation of compounds reduced previously by anaerobic processes (Brenner et al., 2016; Jahnke et al., 1994).

Other potential sources of TA generation in the sediments can be further narrowed down by a more detailed interpretation of changes in DIC (Δ DIC) and TA (Δ TA) during ebb tide as well as their combination with various nutrient ratios. The correlation of DIC and TA reveals an increase in DIC (ΔDIC) that is almost twice as high as that in TA (ΔTA) (Fig. 5a), as indicated by the slope of 1.89. The high Δ DIC points to high aerobic OM degradation and remineralization, resulting in high CO₂ production, which is also indicated by seawater being supersaturated with respect to pCO_2 . High aerobic OM degradation has also previously been observed in the heterotrophic Wadden Sea (e.g., De Beer et al., 2005; van Beusekom et al., 1999), assuming OM degradation and remineralization occurring in the water and sediment in about equal parts (van Beusekom et al., 1999). High OM degradation is indicated by the increasing Corg : N ratios of SPM during ebb tide (Fig. 4k, Table B1). Because we observed constant coastal North Sea salinities, we rule out freshwater runoff and terrestrial signals as a source of the increasing C_{org}: N ratios of SPM. We assume that fresh OM is rapidly degraded in the water column and that the older OM settles on and in the sediment, where the degradation continues and where it is resuspended at low prevailing water levels during ebb. Therefore, we assume that the increase in the SPM concentrations and their Corg : N ratios is an indicator of older and more refractory OM. The increase in TA concentrations points to anaerobic processes, CaCO₃ dissolution, or a combination thereof as TA sources occurring in the sediments.

For an upper-bound estimate of sedimentary CaCO₃ dissolution as source of TA, we considered a ΔDIC : ΔTA ratio of 1:2. Considering this ratio and the observed ΔTA of 51.6 µmol TA kg⁻¹, CaCO₃ dissolution would lead to a ΔDIC of 25.8 µmol DIC kg⁻¹. The remaining 75.5 μ mol DIC kg⁻¹ (101.3–25.8 μ mol DIC kg⁻¹) of the observed ΔDIC in this study could then be produced by OM degradation and remineralization, and it would, using the theoretical expected Redfield ratio of C: N (6.6) for fresh OM (Hickel, 1984), correspond to an estimated dissolved inorganic nitrogen (DIN) production of $11.4 \,\mu\text{mol}\,\text{DIN}\,\text{kg}^{-1}$. However, this estimated DIN production (11.4 µmol DIN kg⁻¹) of OM degradation and remineralization exceeds the observed increase in ΔDIN $(3.97 \,\mu\text{mol}\,\text{DIN}\,\text{L}^{-1}$, sum of NO₃⁻, NO₂⁻ and NH₄⁺; Table B1) during ebb tide. Based on this estimation and the assumption that all DIN produced is released and, thus, lost, TA is probably produced by CaCO₃ dissolution and anaerobic metabolic processes other than denitrification in the sediment. In addition to that, and with an N-focused perspective, the DIN loss also hints at the occurrence of other processes that consume nitrogen species but have no net effect on TA, such as anammox and coupled nitrification– denitrification (Hu and Cai, 2011; Middelburg et al., 2020). The suggested DIN loss can be supported by considering the marine DIN: Si ratio, which is supposed to be about 1:1 (Brzezinski, 1985). We observed DIN: Si ratios decreasing from 2.7 to 0.8 during ebb tide, showing that both parameter concentrations increased, whereby DIN concentrations increased less than silicate concentrations. The silicate excess with respect to DIN at the end of ebb tide supports the DIN loss.

Denitrification, the anaerobic irreversible reduction of NO_3^- to N_2 that generates 0.9 mol of TA using 1 mol of NO_3^- as an electron acceptor (Chen and Wang, 1999), is a net TA source. Denitrification depends on the supply of nitrate, which seasonally varies (van der Zee and Chou, 2005, and references therein). Generally, nitrate is depleted in summer due to high photosynthetic activity and occurs in higher concentrations in winter (Kieskamp et al., 1991; Jensen et al., 1996; van der Zee and Chou, 2005). This seasonality leads to denitrification rates also being lower in summer and higher in winter (Kieskamp et al., 1991; Jensen et al., 1996). In previous studies, Faber et al. (2014) identified denitrification as a minor source of TA due to low denitrification rates, and Kieskamp et al. (1991) also observed low denitrification rates in the Wadden Sea, with low nitrate concentrations (< 2.5 μ mol NO₃⁻ L⁻¹) in the water column. We observed nitrate concentrations ($< 2.17 \,\mu mol \, NO_3^- L^{-1}$) lower than the concentration sufficient for denitrification assumed by Kieskamp et al. (1991). Therefore, we do not exclude denitrification; rather, we suspect it as a minor source of TA in the Dutch Wadden Sea, at least in spring and summer, due to the seasonal lack of nitrate. Thomas et al. (2009) detected TA seasonality in the southern bight of the North Sea, which is also influenced by the TA generation in the Wadden Sea. In addition, the estimated DIN production compared to the observed DIN not only hints at other N-consuming processes that have no effect on TA but also suggests that allochthonous nitrate would be needed to fuel the TA increase by denitrification.

The simultaneous increase in ammonium and TA (Figs. 4c and d and 5d) is important to notice, because the occurrence of ammonium is coupled with nitrification, a process that consumes ammonium and also TA, under oxic conditions (Chen and Wang, 1999). However, under anoxic conditions, such as in deeper sediment layers, ammonium cannot be re-oxidized, accumulates, and is washed out during ebb tide. As we observed low nitrate concentrations and rule out terrestrial nitrate inputs here, the increase in ammonium and TA implies the occurrence of other anaerobic processes of the redox system, such as sulfate and iron reduction, to generate TA in the deeper, anoxic sediment layers in the Dutch Wadden Sea.

Sulfate reduction followed by iron reduction and the formation and burial of pyrite are net sources of TA, as TA consumption by reoxidation is excluded when buried in sediments (Berner et al., 1970; Faber et al., 2014). Whether and to what extent these processes contribute to TA generation in the deeper sediments of the Dutch Wadden Sea cannot be further identified without the necessary data. However, sulfate reduction was also mentioned as source of TA by Thomas et al. (2009), and the temporary slight appearance of a noticeable sulfuric odor could be another indirect indicator of the occurrence of sulfate reduction. In previous studies of tidal flats in the German Wadden Sea, Beck et al. (2008a, b) observed increasing TA concentrations with depth and identified sulfate reduction as the most important process for anaerobic OM remineralization in pore water cores.

A strict comparison of the more northern (north of the Elbe Estuary) and the more western (Texel – Elbe Estuary) parts of the Wadden Sea is difficult, as the areas vary in terms of OM import and eutrophication effects (van Beusekom et al., 2019), sediment composition, and the distance between the barrier islands and the mainland, all of which influence the occurrence and interaction of biogeochemical processes (Schwichtenberg et al., 2020). The area characteristics of the northern and western Wadden Sea specifically differ in terms of the OM turnover being lower in the more northern Wadden Sea. However, a previous study by Brasse et al. (1999) identified high TA and DIC concentrations in the sediment of the North Frisian Wadden Sea and identified CaCO₃ dissolution and sulfate reduction as major TA sources, which is consistent with our findings.

5 Conclusion

The Dutch Wadden Sea is a unique and highly dynamic ecosystem. We observed higher TA values in the Dutch Wadden Sea than in the North Sea and identified the Dutch Wadden Sea as a TA source for the North Sea's carbonate system. Compared to previous studies (Hoppema, 1990, 1993), the TA values that we observed were in a similar range, with high TA values in the tidal basins. In addition to the need for seasonal observations, future work should also focus on the regional and seasonal impacts of freshwater inflows of TA on the TA status in the Dutch Wadden Sea.

By observing salinity and using dissolved silicate as a tracer, we excluded freshwater and river runoff as significant TA sources on the tidal flats and, instead, deduced local outwash from the sediments as sources of TA. Considering various stoichiometries, we suggest that CaCO₃ dissolution generates TA in the more upper, oxic sediment layers and that anaerobic metabolic processes such as denitrification, sulfate, and iron reduction are potential TA sources in the deeper, anoxic sediment layers. However, in spring and early summer, denitrification seems to play a minor role in generating TA in the sediments of the Dutch Wadden Sea due to seasonality and associated limited nitrate availability.

Appendix A



Figure A1. Spatial distribution of (a) silicate (Si; μ molL⁻¹), (b) calcite saturation state (Ω), (c) pH, (d) nitrate (NO₃⁻; μ molL⁻¹), and (e) dissolved inorganic carbon (DIC; μ molkg⁻¹) from surface water samples in May 2019.

Appendix B

Table B1. Half-tidal-cycle sample parameters during ebb tide. Sample no. 545 is the first sample at high tide and sample no. 557 is the last sample at low tide on 21 May 2019 (53.38° N and 5.62° E). Shown are values of temperature (Temp), salinity (Sal), total alkalinity (TA), dissolved inorganic carbon (DIC), silicate (Si), nitrate (NO₃⁻), nitrite (NO₂⁻), ammonium (NH₄⁺), phosphate (PO₄³⁻), dissolved inorganic nitrogen (DIN), the amount of carbon (C) and organic carbon (C_{org}) in SPM, the amount of nitrogen (N) in SPM, the calcite (Ca) and aragonite (Ar) saturation states, the pH, and the seawater partial pressure of CO₂ (*p*CO₂) per sample.

Sample no.	Time (UTC)	Temp (°C)	Sal (PSU)	TA/DIC (µmol kg ⁻¹)	Si (µmol L ⁻¹)	NO_3^- (µmol L ⁻¹)	NO_2^- (µmol L ⁻¹)	NH_4^+ (µmol L ⁻¹)	PO_4^{3-} (µmol L ⁻¹)
E 1 E	10.46	12.20	22.52	2207/2172	1.04	1.20	0.10	2.47	0.12
545 546	10:40	13.20	32.52	238772172	1.84	1.26	0.19	3.47	0.12
540	11:19	13.23	32.32	238372190	1.77	1.24	0.19	3.40	0.11
547	11:49	13.28	32.52	2389/2185	1.72	1.21	0.19	3.35	0.11
548	12:23	13.38	32.52	2391/2183	1.6	1.19	0.19	3.52	0.12
549	13:35	14.32	32.50	2400/2204	2.11	0.91	0.25	3.57	0.32
550	14:05	14.61	32.50	2400/2221	2.78	1.09	0.29	3.98	0.42
551	14:36	14.64	32.51	2405/2216	2.72	1.01	0.29	4.27	0.47
552	15:26	14.73	32.51	2411/2234	4.59	1.23	0.34	5.51	0.57
553	15:42	14.77	32.51	2402/2228	4.24	1.26	0.33	5.08	0.54
554	16:04	14.72	32.51	2419/2234	5.66	1.46	0.36	5.33	0.54
555	16:38	14.66	32.51	2428/2256	8.18	1.77	0.43	6.04	0.58
556	17:07	14.68	32.51	2433/2271	9.79	1.87	0.47	6.27	0.62
557	17:32	14.70	32.50	2438/2273	11.22	2.17	0.50	6.22	0.63
Sample	Time	DIN	C/Corg (SPM)	N (SPM)	Corg : N	SPM	Ca/Ar	pН	pCO ₂
Sample no.	Time (UTC)	$\begin{array}{c} DIN \\ (\mu mol L^{-1}) \end{array}$	$\begin{array}{c} C/C_{org} \ (SPM) \\ (\mu mol L^{-1}) \end{array}$	N (SPM) (µmol L ⁻¹)	C _{org} : N (SPM)	$\frac{\text{SPM}}{(\text{mg}\text{L}^{-1})}$	Ca/Ar (Ω)	pH	pCO ₂ (µatm)
Sample no. 545	Time (UTC) 10:46	DIN (µmolL ⁻¹) 4.93	C/C _{org} (SPM) (µmol L ⁻¹) 86.8/65.1	N (SPM) (μmol L ⁻¹) 8.8	C _{org} : N (SPM) 7.4	SPM (mg L ⁻¹) 12.8	Ca/Ar (Ω) 3.8/2.4	рН 8.07	<i>p</i> CO ₂ (µatm) 385.1
Sample no. 545 546	Time (UTC) 10:46 11:19	DIN (µmolL ⁻¹) 4.93 4.83	C/C _{org} (SPM) (μmol L ⁻¹) 86.8/65.1 72.7/42.4	N (SPM) (µmol L ⁻¹) 8.8 7.4	C _{org} : N (SPM) 7.4 5.8	SPM (mgL ⁻¹) 12.8 8.7	Ca/Ar (Ω) 3.8/2.4 3.5/2.3	pH 8.07 8.03	<i>p</i> CO ₂ (µatm) 385.1 430.2
Sample no. 545 546 547	Time (UTC) 10:46 11:19 11:49	DIN (µmolL ⁻¹) 4.93 4.83 4.76	C/C _{org} (SPM) (µmol L ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \\ \hline 8.8 \\ 7.4 \\ 9.6 \end{array}$	C _{org} : N (SPM) 7.4 5.8 9.7	SPM (mgL ⁻¹) 12.8 8.7 15.4	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3	pH 8.07 8.03 8.05	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4
Sample no. 545 546 547 548	Time (UTC) 10:46 11:19 11:49 12:23	DIN (µmolL ⁻¹) 4.93 4.83 4.76 4.91	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6	N (SPM) (µmolL ⁻¹) 8.8 7.4 9.6 9.9	C _{org} : N (SPM) 7.4 5.8 9.7 10.5	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4	pH 8.07 8.03 8.05 8.05	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1
Sample no. 545 546 547 548 548 549	Time (UTC) 10:46 11:19 11:49 12:23 13:35	DIN (µmolL ⁻¹) 4.93 4.83 4.76 4.91 4.73	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8	N (SPM) (µmolL ⁻¹) 8.8 7.4 9.6 9.9 8.8	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3	pH 8.07 8.03 8.05 8.05 8.05 8.01	<i>p</i> CO ₂ (µаtm) 385.1 430.2 411.4 404.1 452.3
Sample no. 545 546 547 548 549 550	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05	DIN (µmolL ⁻¹) 4.93 4.83 4.76 4.91 4.73 5.37	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3	N (SPM) (µmolL ⁻¹) 8.8 7.4 9.6 9.9 8.8 17.7	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1	pH 8.07 8.03 8.05 8.05 8.05 8.01 7.97	<i>p</i> CO ₂ (µаtm) 385.1 430.2 411.4 404.1 452.3 507.2
Sample no. 545 546 547 548 549 550 551	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36	DIN (µmolL ⁻¹) 4.93 4.83 4.76 4.91 4.73 5.37 5.56	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3	N (SPM) (µmolL ⁻¹) 8.8 7.4 9.6 9.9 8.8 17.7 14.5	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99	<i>p</i> CO ₂ (µаtm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9
Sample no. 545 546 547 548 549 550 551 552	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36 15:26	$\begin{array}{c} \text{DIN} \\ (\mu\text{mol}L^{-1}) \\ \hline 4.93 \\ 4.83 \\ 4.76 \\ 4.91 \\ 4.73 \\ 5.37 \\ 5.56 \\ 7.08 \end{array}$	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3 248.6/163.5	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array} \\ \begin{array}{c} 8.8 \\ 7.4 \\ 9.6 \\ 9.9 \\ 8.8 \\ 17.7 \\ 14.5 \\ 18.4 \end{array}$	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0 8.9	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6 34.3	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2 3.3/2.1	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99 7.96	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9 520.0
Sample no. 545 546 547 548 549 550 551 552 552 553	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36 15:26 15:42	$\begin{array}{c} \text{DIN} \\ (\mu\text{mol}L^{-1}) \\ \hline 4.93 \\ 4.83 \\ 4.76 \\ 4.91 \\ 4.73 \\ 5.37 \\ 5.56 \\ 7.08 \\ 6.67 \\ \end{array}$	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3 248.6/163.5 257.6/199.3	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array} \\ \begin{array}{c} 8.8 \\ 7.4 \\ 9.6 \\ 9.9 \\ 8.8 \\ 17.7 \\ 14.5 \\ 18.4 \\ 18.3 \end{array}$	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0 8.9 10.9	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6 34.3 41.6	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2 3.3/2.1 3.2/2.1	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99 7.96 7.95	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9 520.0 526.4
Sample no. 545 546 547 548 549 550 551 552 553 554	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36 15:26 15:42 16:04	DIN (µmolL ⁻¹) 4.93 4.83 4.76 4.91 4.73 5.37 5.56 7.08 6.67 7.15	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3 248.6/163.5 257.6/199.3 324.4/271.1	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array}$	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0 8.9 10.9 11.7	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6 34.3 41.6 55.0	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2 3.3/2.1 3.2/2.1 3.4/2.2	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99 7.96 7.95 7.98	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9 520.0 526.4 496.6
Sample no. 545 546 547 548 549 550 551 552 553 554 555	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36 15:26 15:42 16:04 16:38	$\begin{array}{c} DIN\\ (\mu mol L^{-1})\\ \hline 4.93\\ 4.83\\ 4.76\\ 4.91\\ 4.73\\ 5.37\\ 5.56\\ 7.08\\ 6.67\\ 7.15\\ 8.24\\ \end{array}$	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3 248.6/163.5 257.6/199.3 324.4/271.1 440.4/345.2	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \\ \hline \\ 8.8 \\ 7.4 \\ 9.6 \\ 9.9 \\ 8.8 \\ 17.7 \\ 14.5 \\ 18.4 \\ 18.3 \\ 23.2 \\ 29.2 \\ \end{array}$	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0 8.9 10.9 11.7 11.8	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6 34.3 41.6 55.0 75.7	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2 3.3/2.1 3.2/2.1 3.4/2.2 3.2/2.1	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99 7.96 7.95 7.98 7.95	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9 520.0 526.4 496.6 538.0
Sample no. 545 546 547 548 549 550 551 552 553 554 555 555 556	Time (UTC) 10:46 11:19 11:49 12:23 13:35 14:05 14:36 15:26 15:42 16:04 16:38 17:07	$\begin{array}{c} \text{DIN} \\ (\mu\text{mol}L^{-1}) \\ \hline 4.93 \\ 4.83 \\ 4.76 \\ 4.91 \\ 4.73 \\ 5.37 \\ 5.56 \\ 7.08 \\ 6.67 \\ 7.15 \\ 8.24 \\ 8.61 \\ \end{array}$	C/C _{org} (SPM) (µmolL ⁻¹) 86.8/65.1 72.7/42.4 112.4/93.4 108.5/104.6 111.1/97.8 233.0/180.3 193.2/174.3 248.6/163.5 257.6/199.3 324.4/271.1 440.4/345.2 430.5/363.3	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array} \\ \begin{array}{c} 8.8 \\ 7.4 \\ 9.6 \\ 9.9 \\ 8.8 \\ 17.7 \\ 14.5 \\ 18.4 \\ 18.3 \\ 23.2 \\ 29.2 \\ 27.9 \end{array}$	Corg : N (SPM) 7.4 5.8 9.7 10.5 11.1 10.2 12.0 8.9 10.9 11.7 11.8 13.0	SPM (mgL ⁻¹) 12.8 8.7 15.4 16.8 13.9 32.2 29.6 34.3 41.6 55.0 75.7 82.4	Ca/Ar (Ω) 3.8/2.4 3.5/2.3 3.7/2.3 3.7/2.4 3.6/2.3 3.3/2.1 3.5/2.2 3.3/2.1 3.2/2.1 3.4/2.2 3.2/2.1 3.1/2.0	pH 8.07 8.03 8.05 8.05 8.01 7.97 7.99 7.96 7.95 7.98 7.95 7.93	<i>p</i> CO ₂ (μatm) 385.1 430.2 411.4 404.1 452.3 507.2 477.9 520.0 526.4 496.6 538.0 576.6

Table B2. Half-tidal-cycle sample parameters during high tide for comparison. Sample no. 564 is the first sample at low tide and sample no. 578 is the last sample at high tide on 23 May 2019 (53.39° N and 5.63° E, 5.62° E^{*}). Shown are values of temperature (Temp), salinity (Sal), total alkalinity (TA), dissolved inorganic carbon (DIC), silicate (Si), nitrate (NO₃⁻), nitrite (NO₂⁻), ammonium (NH₄⁺), phosphate (PO₄³⁻), dissolved inorganic nitrogen (DIN), the amount of carbon (C) and organic carbon (C_{org}) in SPM, the amount of nitrogen (N) in SPM, the calcite (Ca) and aragonite (Ar) saturation states, the pH, and the seawater partial pressure of CO₂ (*p*CO₂) per sample.

Sample	Time	Temp	Sal	TA/DIC	Si	NO_3^-	NO_2^-	NH_4^+	PO_4^{3-}
no.	(UTC)	(°C)	(PSU)	$(\mu mol kg^{-1})$	$(\mu mol L^{-1})$	$(\mu mol L^{-1})$	$(\mu mol L^{-1})$	$(\mu mol L^{-1})$	$(\mu mol L^{-1})$
564	05:09	14.04	32.66	2431/2246	8.53	1.25	0.47	3.31	0.38
565	05:32	14.02	32.68	2441/2287	9.14	1.26	0.45	3.08	0.37
566	06:01	13.95	32.69	2436/2284	8.88	1.33	0.38	2.46	0.34
567	06:33	14.16	32.69	2443/2284	8.68	0.95	0.37	2.37	0.33
568	07:02	14.21	32.69	2432/2280	6.94	0.75	0.34	2.63	0.32
569	07:31	14.15	32.55	2401/2223	2.12	0.98	0.27	4.12	0.33
570	08:04	14.20	32.55	2403/2218	2.10	1.04	0.27	3.88	0.30
571	08:35	14.27	32.55	2409/2228	2.15	0.92	0.25	4.18	0.32
572	09:04	14.37	32.53	2400/2209	1.88	1.00	0.22	3.86	0.26
573	09:34	14.16	32.52	2398/2200	1.70	1.03	0.21	3.51	0.21
574*	10:02	14.17	32.52	2391/2197	1.72	1.07	0.21	3.40	0.18
575*	10:34	14.11	32.51	2389/2195	1.78	1.18	0.20	3.45	0.16
576	11:04	14.21	32.50	2390/2187	1.76	1.12	0.19	3.29	0.14
577	11:34	14.50	32.51	2399/2193	1.66	1.10	0.20	3.32	0.16
578	12:03	13.96	32.51	2390/2187	1.75	1.41	0.19	3.72	0.11
Sample	Time	DIN	C/Corg (SPM)	N (SPM)	Corg : N	SPM	Ca/Ar	pH	pCO_2
Sample no.	Time (UTC)	$\begin{array}{c} \text{DIN} \\ (\mu mol L^{-1}) \end{array}$	$\begin{array}{c} C/C_{org} \ (SPM) \\ (\mu mol L^{-1}) \end{array}$	N (SPM) (µmol L ⁻¹)	C _{org} : N (SPM)	$\begin{array}{c} \text{SPM} \\ (\text{mg}\text{L}^{-1}) \end{array}$	Ca/Ar (Ω)	pH	pCO ₂ (µatm)
Sample no. 564	Time (UTC) 05:09	DIN (µmolL ⁻¹) 5.03	C/C _{org} (SPM) (µmolL ⁻¹) 353.7/253.2	N (SPM) (μmol L ⁻¹) 27.5	C _{org} : N (SPM) 9.2	SPM (mgL ⁻¹) 52.3	Ca/Ar (Ω) 3.0/2.2	рН 7.99	<i>p</i> CO ₂ (µatm) 490.3
Sample no. 564 565	Time (UTC) 05:09 05:32	DIN (µmolL ⁻¹) 5.03 4.78	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1	N (SPM) (μmol L ⁻¹) 27.5 26.1	C _{org} : N (SPM) 9.2 8.4	SPM (mgL ⁻¹) 52.3 49.7	Ca/Ar (Ω) 3.0/2.2 3.0/1.9	pH 7.99 7.92	<i>p</i> CO ₂ (μatm) 490.3 592.9
Sample no. 564 565 566	Time (UTC) 05:09 05:32 06:01	DIN (µmolL ⁻¹) 5.03 4.78 4.17	C/C _{org} (SPM) (µmolL ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9	N (SPM) (μmolL ⁻¹) 27.5 26.1 25.5	C _{org} : N (SPM) 9.2 8.4 9.1	SPM (mgL ⁻¹) 52.3 49.7 51.7	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9	pH 7.99 7.92 7.91	рСО ₂ (µatm) 490.3 592.9 600.3
Sample no. 564 565 566 566 567	Time (UTC) 05:09 05:32 06:01 06:33	DIN (µmolL ⁻¹) 5.03 4.78 4.17 3.68	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7	N (SPM) (µmolL ⁻¹) 27.5 26.1 25.5 21.8	Corg : N (SPM) 9.2 8.4 9.1 9.0	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9	pH 7.99 7.92 7.91 7.92	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6
Sample no. 564 565 566 567 568	Time (UTC) 05:09 05:32 06:01 06:33 07:02	DIN (µmolL ⁻¹) 5.03 4.78 4.17 3.68 3.72	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2	N (SPM) (µmolL ⁻¹) 27.5 26.1 25.5 21.8 24.5	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9	pH 7.99 7.92 7.91 7.92 7.91	рСО ₂ (µatm) 490.3 592.9 600.3 582.6 601.8
Sample no. 564 565 566 567 568 569	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31	DIN (µmolL ⁻¹) 5.03 4.78 4.17 3.68 3.72 5.37	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1	N (SPM) (µmolL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1	pH 7.99 7.92 7.91 7.92 7.91 7.98	рСО ₂ (µаtm) 490.3 592.9 600.3 582.6 601.8 500.7
Sample no. 564 565 566 567 568 569 570	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04	DIN (µmolL ⁻¹) 5.03 4.78 4.17 3.68 3.72 5.37 5.20	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6	N (SPM) (µmolL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0 8.8	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99	рСО ₂ (µаtm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6
Sample no. 564 565 566 567 568 569 570 571	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35	DIN (µmolL ⁻¹) 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6	N (SPM) (µmolL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0 8.8 9.9	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98	рСО ₂ (µаtm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6
Sample no. 564 565 566 567 568 569 570 571 572	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04	DIN (μ molL ⁻¹) 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35 5.08	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array}$ $\begin{array}{c} 27.5 \\ 26.1 \\ 25.5 \\ 21.8 \\ 24.5 \\ 7.0 \\ 8.8 \\ 9.9 \\ 9.9 \\ 9.9 \end{array}$	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6
Sample no. 564 565 566 567 568 569 570 571 572 573	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04 09:34	DIN (μ molL ⁻¹) 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35 5.08 4.75	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9 82.1/72.7	$\begin{array}{c} \text{N (SPM)} \\ (\mu\text{mol}\text{L}^{-1}) \\ \hline 27.5 \\ 26.1 \\ 25.5 \\ 21.8 \\ 24.5 \\ 7.0 \\ 8.8 \\ 9.9 \\ 9.9 \\ 9.9 \\ 7.2 \\ \end{array}$	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5 10.0	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4 11.8	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2 3.6/2.3	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00 8.02	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6 445.3
Sample no. 564 565 566 567 568 569 570 571 572 573 574*	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04 09:34 10:02	DIN $(\mu mol L^{-1})$ 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35 5.08 4.75 4.68	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9 82.1/72.7 85.2/62.9	N (SPM) (μ molL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0 8.8 9.9 9.9 9.9 7.2 7.2 7.2	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5 10.0 8.7	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4 11.8 9.9	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2 3.6/2.3 3.5/2.3	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00 8.02 8.01	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6 445.3 450.5
Sample no. 564 565 566 567 568 569 570 571 572 573 574* 575*	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04 09:34 10:02 10:34	DIN $(\mu mol L^{-1})$ 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35 5.08 4.75 4.68 4.83	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9 82.1/72.7 85.2/62.9 83.5/65.9	N (SPM) (μ molL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0 8.8 9.9 9.9 9.9 7.2 7.2 7.2 7.2	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5 10.0 8.7 9.2	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4 11.8 9.9 11.1	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2 3.6/2.3 3.5/2.3	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00 8.02 8.01 8.01	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6 445.3 450.5 449.6
Sample no. 564 565 566 567 568 569 570 571 572 573 574* 575* 576	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04 09:34 10:02 10:34 11:04	DIN $(\mu mol L^{-1})$ 5.03 4.78 4.17 3.68 3.72 5.37 5.20 5.35 5.08 4.75 4.68 4.83 4.60	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9 82.1/72.7 85.2/62.9 83.5/65.9 82.7/52.1	N (SPM) (μ molL ⁻¹) 27.5 26.1 25.5 21.8 24.5 7.0 8.8 9.9 9.9 7.2 7.2 7.2 7.2 8.2	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5 10.0 8.7 9.2 6.3	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4 11.8 9.9 11.1 8.5	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2 3.6/2.3 3.5/2.3 3.5/2.3 3.5/2.3	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00 8.02 8.01 8.01 8.03	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6 445.3 450.5 449.6 429.9
Sample no. 564 565 566 567 568 569 570 571 572 573 574* 575* 576 577	Time (UTC) 05:09 05:32 06:01 06:33 07:02 07:31 08:04 08:35 09:04 09:34 10:02 10:34 11:04 11:34	$\begin{array}{c} \text{DIN} \\ (\mu\text{mol}L^{-1}) \\ \hline 5.03 \\ 4.78 \\ 4.17 \\ 3.68 \\ 3.72 \\ 5.37 \\ 5.20 \\ 5.35 \\ 5.08 \\ 4.75 \\ 4.68 \\ 4.83 \\ 4.60 \\ 4.62 \end{array}$	C/C _{org} (SPM) (µmol L ⁻¹) 353.7/253.2 333.5/220.1 330.3/232.9 274.7/195.7 317.8/220.2 88.6/59.1 96.8/73.6 114.2/109.6 107.5/73.9 82.1/72.7 85.2/62.9 83.5/65.9 82.7/52.1 65.8/50.8	$\begin{array}{c} N \ (SPM) \\ (\mu mol \ L^{-1}) \end{array}$	Corg : N (SPM) 9.2 8.4 9.1 9.0 9.0 8.5 8.4 11.0 7.5 10.0 8.7 9.2 6.3 7.8	SPM (mgL ⁻¹) 52.3 49.7 51.7 36.9 46.1 14.7 18.1 14.8 16.4 11.8 9.9 11.1 8.5 7.2	Ca/Ar (Ω) 3.0/2.2 3.0/1.9 2.9/1.9 3.0/1.9 2.9/1.9 3.3/2.1 3.4/2.2 3.3/2.1 3.5/2.2 3.6/2.3 3.5/2.3 3.5/2.3 3.7/2.4	pH 7.99 7.92 7.91 7.92 7.91 7.98 7.99 7.98 8.00 8.02 8.01 8.01 8.01 8.03 8.03	<i>p</i> CO ₂ (μatm) 490.3 592.9 600.3 582.6 601.8 500.7 482.6 497.6 466.6 445.3 450.5 449.6 429.9 430.8

Table B3. Transect parameters of cruise LP20190515 on RV Ludwig Prandtl in the Dutch Wadden Sea in May 2019. Shown are values
of latitude (Lat), longitude (Long), temperature (Temp), salinity (Sal), total alkalinity (TA), dissolved inorganic carbon (DIC), silicate (Si),
nitrate (NO3 ⁻), the calcite (Ca) and aragonite (Ar) saturation states, and pH per sample. Our salinity and temperature data were comple-
mented by data from three Rijkswaterstaat stations, which were close to our stations. Of these three stations, Dantziggat ^a (53°24'4.093" N,
5°43'37.132" E) showed temperatures of 11.4 and 14.9 °C and salinities of 31.9 and 31.2 PSU on 10 and 27 May 2019, respectively; Ter-
schelling 10 ^b (53°27'37.318" N, 5°5'58.129" E) showed temperatures of 11.4 and 12.9 °C and salinities of 32.8 and 33.4 PSU on 15 and
28 May 2019, respectively; and Vliestroom ^c (53°18′48.054″ N, 5°9′35.655″ E) showed a temperature of 11.8 °C and a salinity of 31.1 PSU
on 14 May 2019.

Sample	Time	Day in May	Lat $(\circ \mathbf{N})$	Long	Temp	Sal	TA/DIC	Si/NO_3^-	Ca/Ar	pН
no.	(010)	Way	(N)	(E)	()	(PSU)	(µmorkg -)	(µmort -)	(22)	
535	07:56	20	53.18	5.4	14.72	30.24	2507/2357	10.00/5.10	3.0/1.9	7.92
536	08:26	20	53.19	5.34	14.81	32.51	2458/2296	3.45/0.46	3.1/2.0	7.92
537	08:53	20	53.22	5.29	14.05	32.38	2413/2227	1.92/0.85	3.4/2.2	8.00
538	09:28	20	53.23	5.19	13.36	32.65	2381/2153	0.52/1.02	4.0/2.6	8.10
539	09:53	20	53.27	5.17	13.17	32.65	2389/2161	0.45/1.37	4.0/2.6	8.10
540 ^c	10:36	20	53.33	5.12	12.77	32.97	2375/2118	0.32/0.84	4.4/2.8	8.16
541	11:03	20	53.32	5.0	12.41	33.25	2368/2097	0.34/0.77	4.6/3.0	8.19
542 ^b	11:49	20	53.4	5.1	12.93	32.92	2374/2109	0.44/0.81	4.6/2.9	8.17
543	12:49	20	53.45	5.33	12.95	32.45	2385/2196	6.25/1.85	3.4/2.2	8.02
544	13:31	20	53.46	5.5	13.55	32.51	2388/2169	3.02/1.30	3.9/2.5	8.08
558	11:33	22	53.41	5.65	13.31	32.51	2384/2224	2.57/1.56	3.0/1.9	7.95
559	12:04	22	53.4	5.66	13.45	32.51	2393/2195	1.58/1.41	3.6/2.3	8.03
560	12:40	22	53.39	5.69	13.67	32.52	2391/2183	1.52/1.33	3.7/2.4	8.05
561	13:09	22	53.4	5.73	14.23	32.48	2418/2242	2.04/1.04	3.3/2.1	7.97
562	13:32	22	53.42	5.77	14.71	32.51	2417/2237	3.23/1.04	3.3/2.1	7.97
563 ^a	13:56	22	53.42	5.82	15.33	32.45	2421/2242	4.68/0.86	3.3/2.1	7.96
579	09:05	24	53.42	5.77	15.26	32.65	2417/2215	2.71/0.86	3.7/2.4	8.01
580	09:31	24	53.4	5.73	14.99	32.66	2426/2249	3.52/1.46	3.3/2.1	7.95
581	10:01	24	53.4	5.66	13.87	32.58	2396/2205	1.69/1.60	3.5/2.2	8.01
582	10:25	24	53.43	5.61	14.36	32.51	2389/2193	2.23/2.52	3.6/2.3	8.02
583	10:59	24	53.5	5.61	13.48	32.58	2382/2187	3.69/3.41	3.5/2.2	8.03
584	11:31	24	53.51	5.76	13.50	32.59	2390/2172	1.93/2.96	3.9/2.5	8.07
585	12:00	24	53.52	5.9	13.65	32.59	2390/2173	2.34/2.23	3.9/2.5	8.07
586	12:30	24	53.52	6.04	13.50	32.48	2384/2179	2.00/1.53	3.7/2.3	8.05
587	13:02	24	53.45	6.07	15.13	32.40	2389/2169	0.70/1.19	3.9/2.5	8.05
588	13:31	24	53.42	6.38	15.63	31.96	2396/2182	1.33/0.62	3.9/2.5	8.04
589	07:20	25	53.42	6.18	15.73	28.31	2430/2245	4.16/5.13	3.6/2.3	8.02
590	07:52	25	53.44	6.09	15.80	30.90	2407/2225	1.58/1.39	3.4/2.2	7.98
591	08:21	25	53.48	6.08	15.34	31.82	2395/2222	4.16/5.09	3.3/2.1	7.96
592	08:51	25	53.52	6.05	14.80	32.41	2386/2178	0.71/0.67	3.8/2.4	8.04
593	09:22	25	53.55	6.18	13.96	32.30	2379/2175	0.36/1.52	3.7/2.3	8.04
594	09:53	25	53.58	6.31	13.43	32.14	2332/2148	0.34/5.75	3.3/2.1	8.01
595	10:24	25	53.61	6.45	13.47	32.10	2347/2113	0.26/5.19	4.1/2.6	8.12
596	11:05	25	53.58	6.63	14.50	29.99	2381/2184	0.78/20.25	3.7/2.3	8.05
597	11:33	25	53.52	6.75	14.94	29.17	2383/2214	3.04/27.84	3.3/2.1	7.99
598	12:00	25	53.47	6.85	15.28	27.82	2395/2249	8.90/37.93	3.0/1.9	7.94
599	12:30	25	53.4	6.95	15.46	26.39	2423/2284	17.63/36.54	2.9/1.8	7.94
600	12:59	25	53.33	7.02	15.76	23.01	2460/2343	41.93/37.68	2.7/1.7	7.92
601	13:29	25	53.33	7.16	15.96	20.25	2517/2430	56.32/37.94	2.3/1.4	7.86

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Data availability. The data generated and presented in this study are given in Appendix A and B.

Author contributions. MN wrote the manuscript, carried out the carbon sampling and sample measurement, analyzed and evaluated the data, and led the study. JEEvB led the research cruise. JEEvB and HT provided editorial and scientific recommendations. MN prepared the manuscript with contributions from all co-authors.

Competing interests. The contact author has declared that none of the authors has any competing interests.

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